

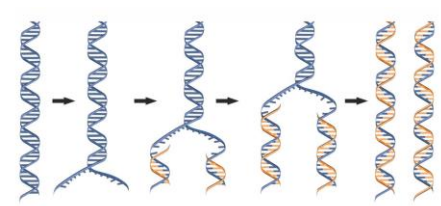
# **Lecture 3:**

## **Nucleic Acid Technologies**

- Review of DNA Polymerase activity
- Nucleic Acid Technologies – PCR & Sequencing

## **Immunology and Immunotherapies**

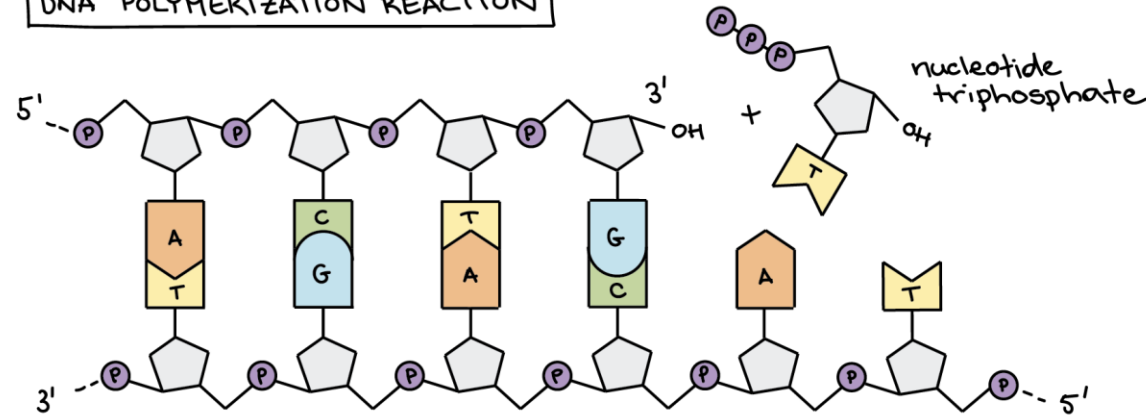
- Overview of immune system
- Antibody Response
- Antibody based therapies for cancer (Final presentations)
- Cell-based Response
- Cell-based cancer therapy (Final presentations)
- Vaccines & Vaccine development



# DNA Polymerase – Fundamental Activity.

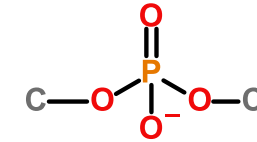
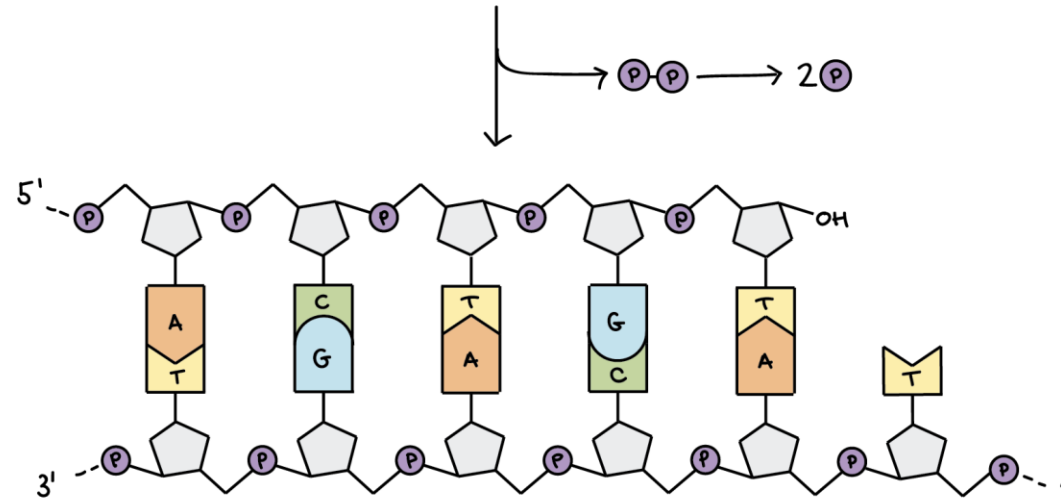
5' to 3'  
polymerization

DNA POLYMERIZATION REACTION

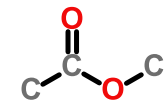


1. Where on the deoxyribose is the new base added?

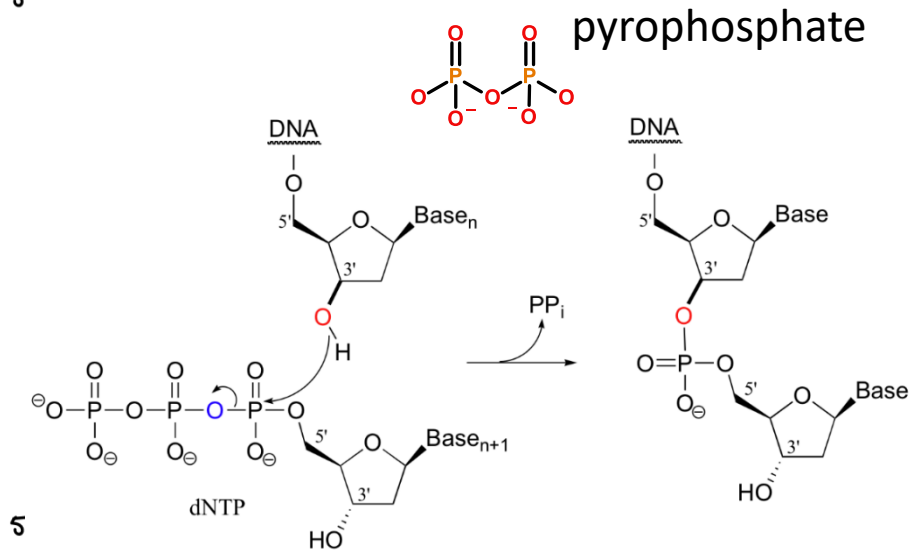
2. What determines which base is to be added?



Phosphodiester linkage



ester linkage



# DNA Polymerase – Fundamental Activity.

5' A-T-C-A

3' A-T-G-C-C-G-T-A-G-T-C-G-T-A-C-A-G-T-A-C-G-T-G-C-A  
1 2 3 4 5 6 7 8 9 0 1 2 3 4 5 6 7 8 9 0 1 2 3 4 5  
1 2

1. *Where (what position) will this primer (ATCA) anneal?*
2. *What base will be added first?*
3. *What is the last base added?*

# DNA Sequencing – Sanger (dideoxy) Sequencing

DNA Sequencing - Determining the Order of Bases in the DNA.

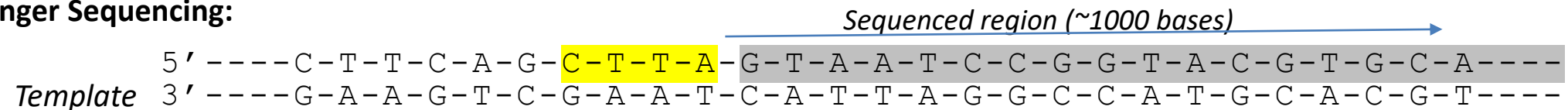
Sanger Sequencing:

- Second method to generate long (~1000 base) sequence information (an earlier chemical method developed by Gilbert proved to be impractical for most laboratories (hydrazine = rocket fuel was required))
- Sanger was awarded his 2<sup>nd</sup> Nobel prize for this work in 1980, shared with Gilbert.



Determine the position of all four bases in a DNA strand = Sequence (video)

**Sanger Sequencing:**

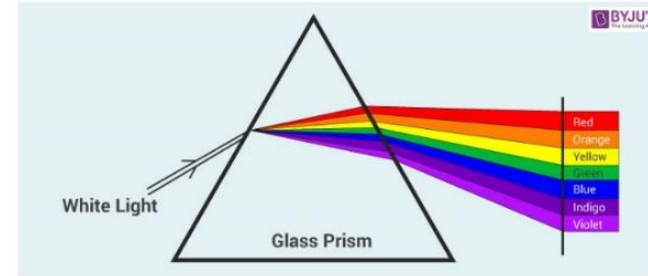
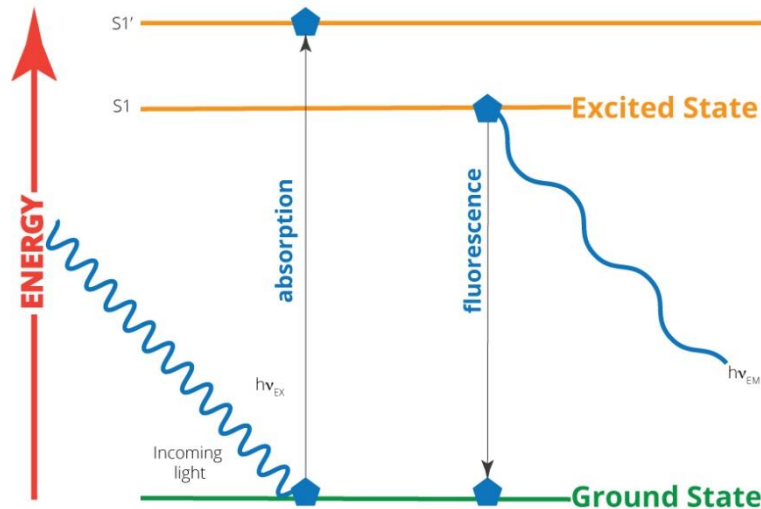


*Primer*                      5' **C-T-T-A**<sup>OH</sup>

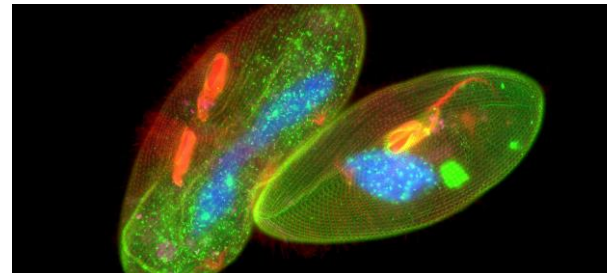
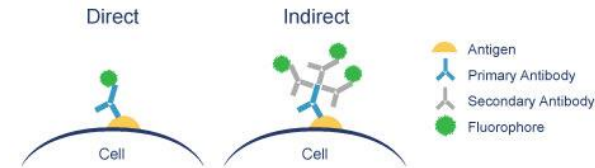
*Template*   3' ----G-A-A-G-T-C-G-A-A-T-C-A-T-T-A-G-G-C-C-A-T-G-C-A-C-G-T----

# What is fluorescence?

- When molecules absorb light an electron goes from a lower shell to a higher shell. This is where the energy from the light goes.
- In most molecules the electron goes back down to its original shell with the release of heat.
- Fluorescent molecules emit the energy as light of a longer wavelength (different color).
- The color that is emitted depends on the molecule.

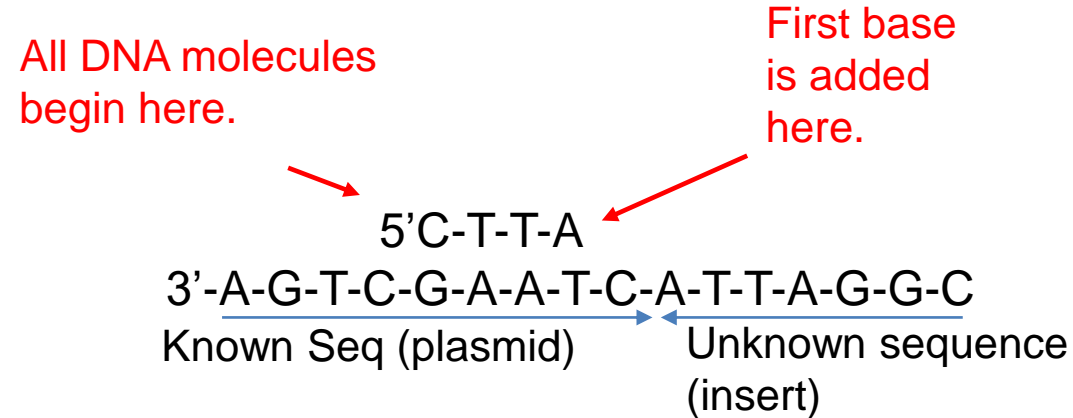


Fluorescently tagged antibodies can be used to stain components of cell with fluorophores.



# DNA Sequencing - Determining the Order of Bases Added by DNA Polymerase

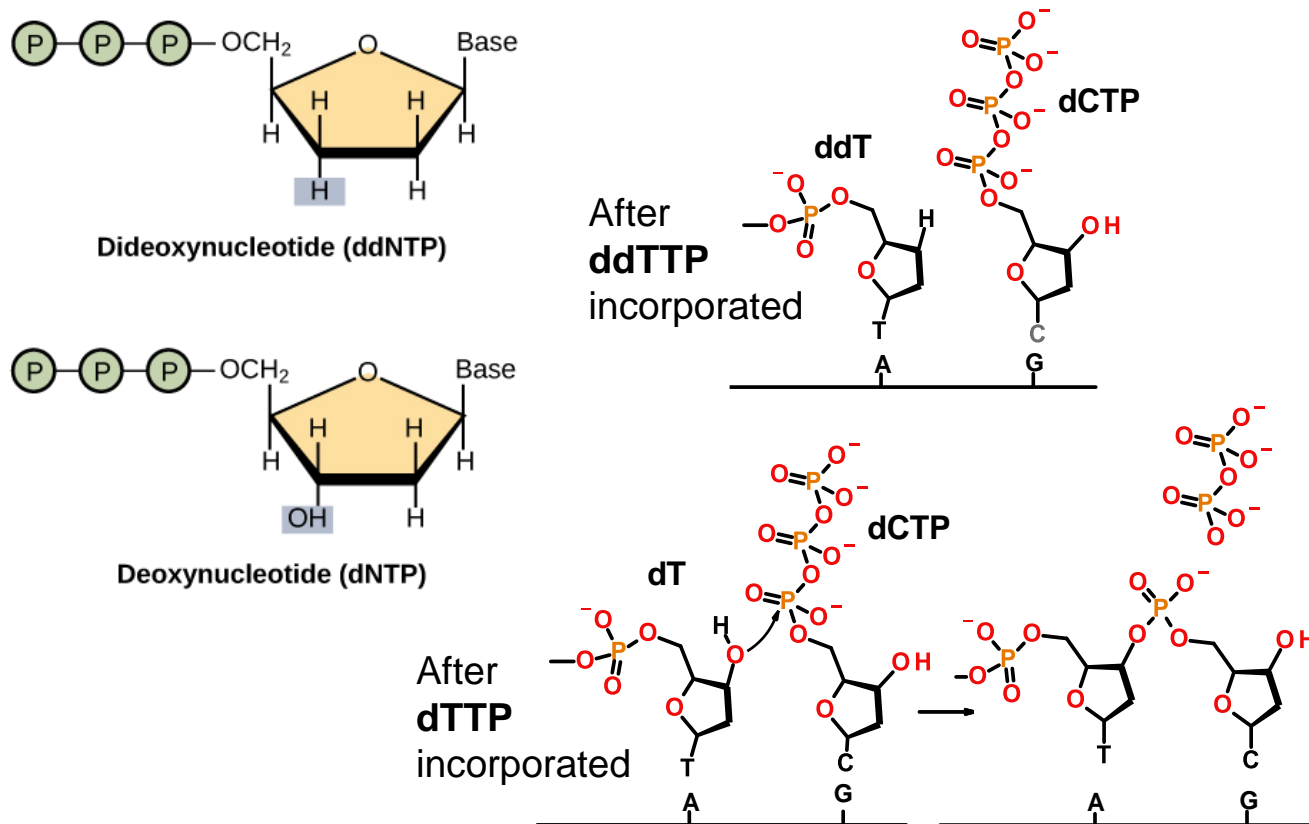
1. Start sequencing at known location with primer that anneals at a **unique** location on the plasmid, “upstream” from the region to be sequenced.



# DNA Sequencing - Determining the Order of Bases Added by DNA Polymerase

2. Use a mixture of normal bases (dNTPs) and dideoxy bases (ddNTP) for polymerization. Ratio of dNTP to ddNTP is (100:1), most of the time elongation occurs.

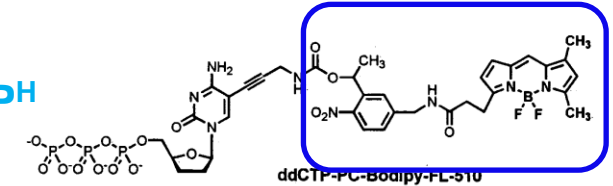
- ddNTPs can be added to the DNA since they have a 5'-triphosphate but **terminate** the chain due to the lack of a 3'-OH. ~ 1 in 100 chains terminate at each base addition



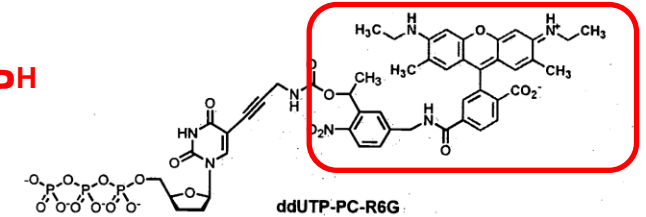
3. The ddNTPs are color coded by different fluorescent emission wavelengths.

*The ddNTP that terminated the chain is known from its fluorescent color.*

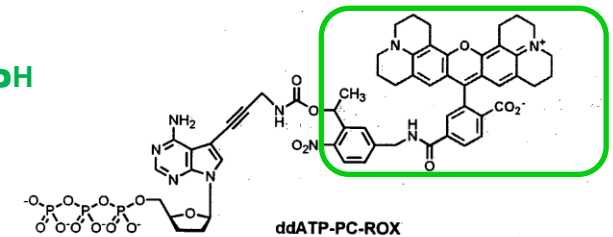
ddCTP<sup>H</sup>



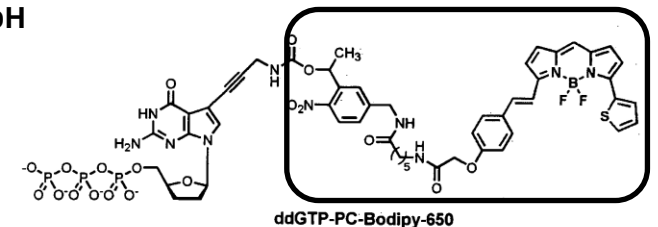
ddTTP<sup>H</sup>

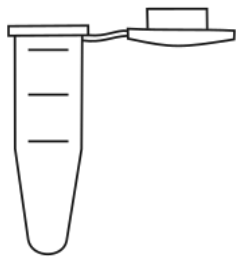


ddATP<sup>H</sup>



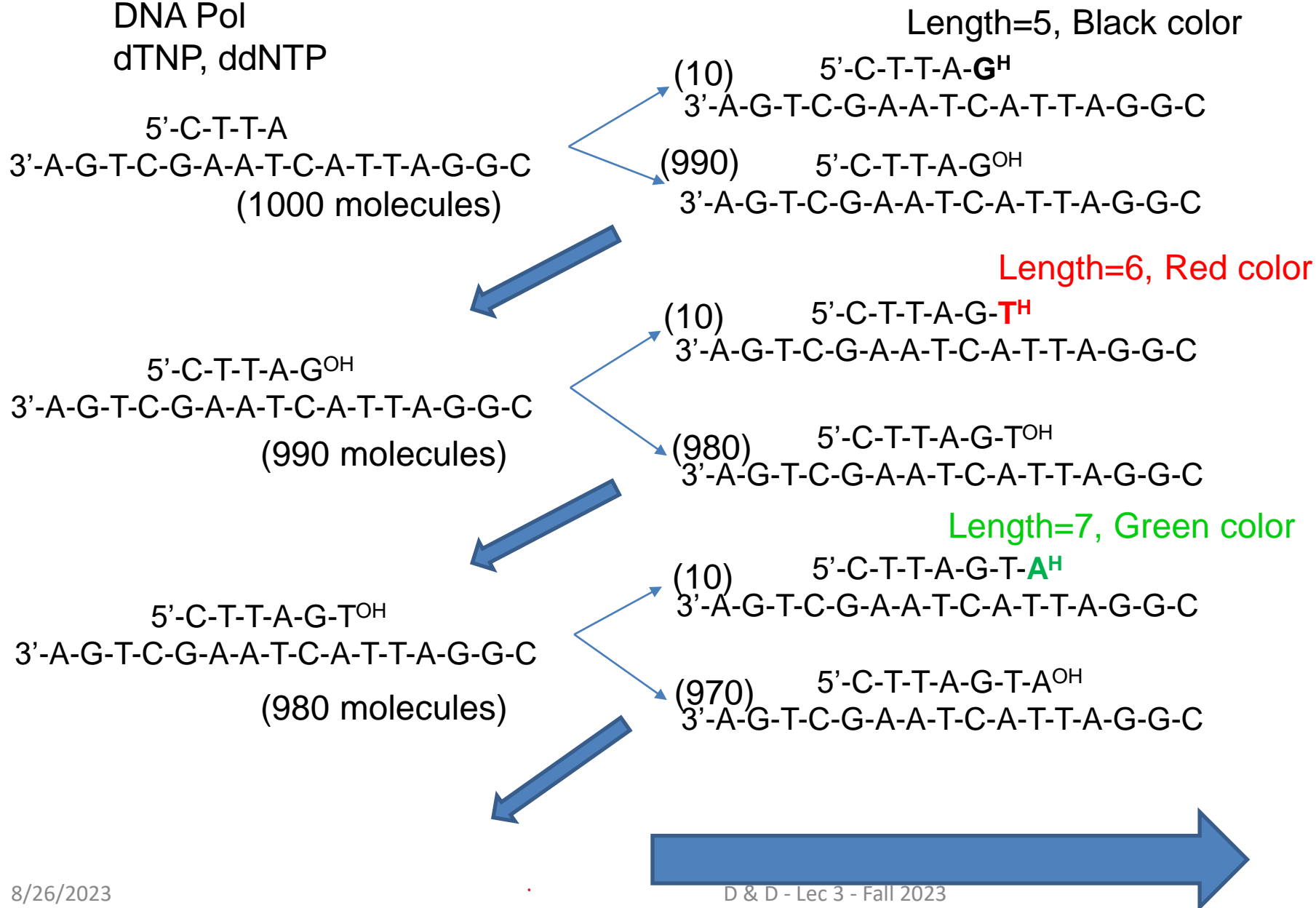
ddGTP<sup>H</sup>





Template  
Primer  
DNA Pol  
dTNP, ddNTP

# DNA Sequencing – Generation of Fragments



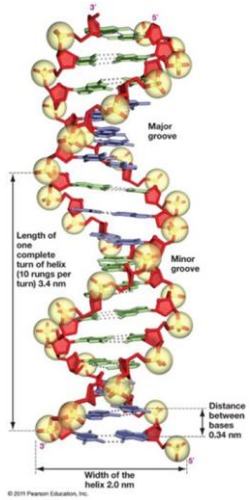
## All Possible Fragments are Made:

1. Each begins with the primer
2. Each ends with a *known* ddNTP, based on the color of the fluorescence.
3. Each is one longer than the previous.

C-T-T-A-G  
C-T-T-A-G-T  
C-T-T-A-G-T-A  
C-T-T-A-G-T-A-A  
C-T-T-A-G-T-A-A-T  
C-T-T-A-G-T-A-A-T-C  
C-T-T-A-G-T-A-A-T-C-C  
Primer      Added by Pol.



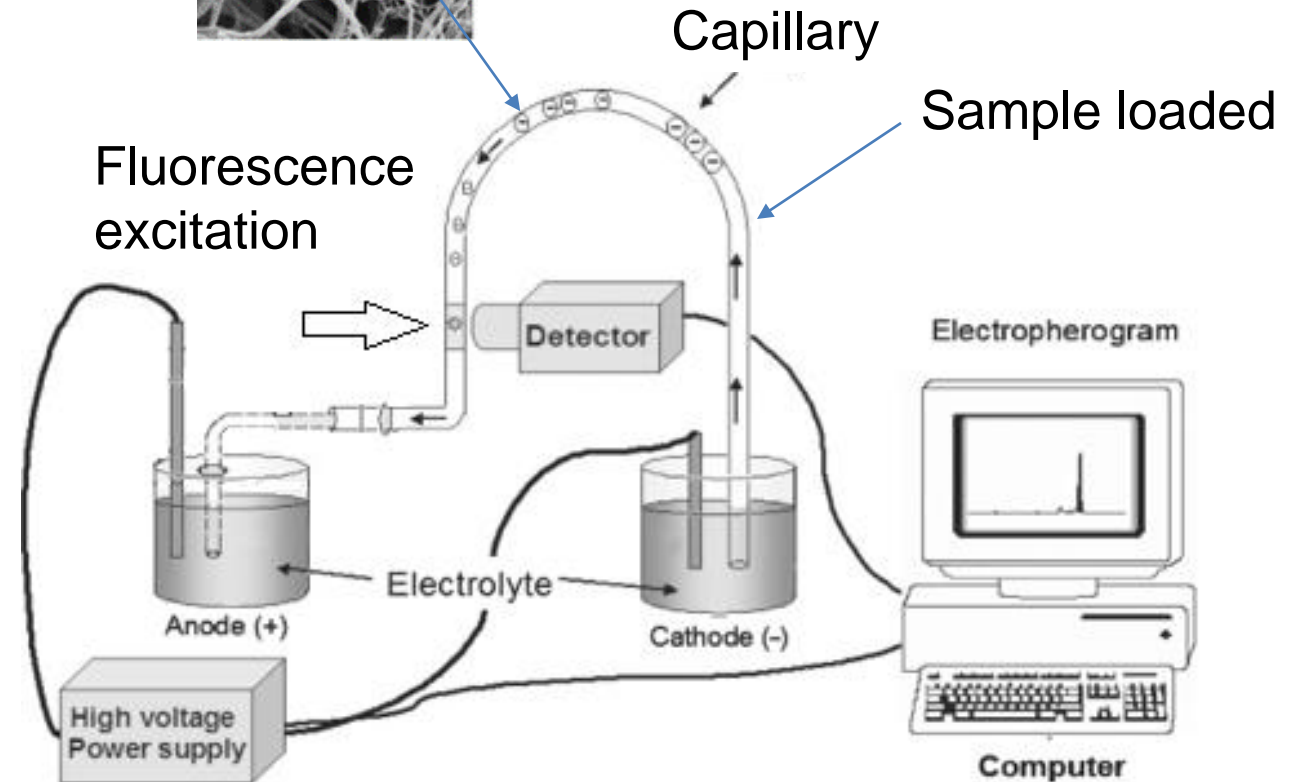
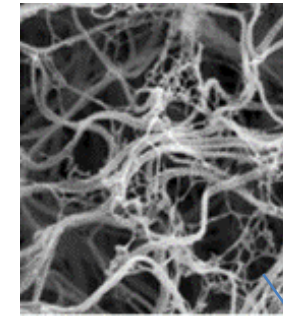
# Size Determination of Fragments from DNA Sequencing Capillary Electrophoresis



DNA has a negative charge.  
It will migrate towards the anode.

Capillary is filled with a gel that causes  
separation by size.

DNA molecules that are smaller migrate

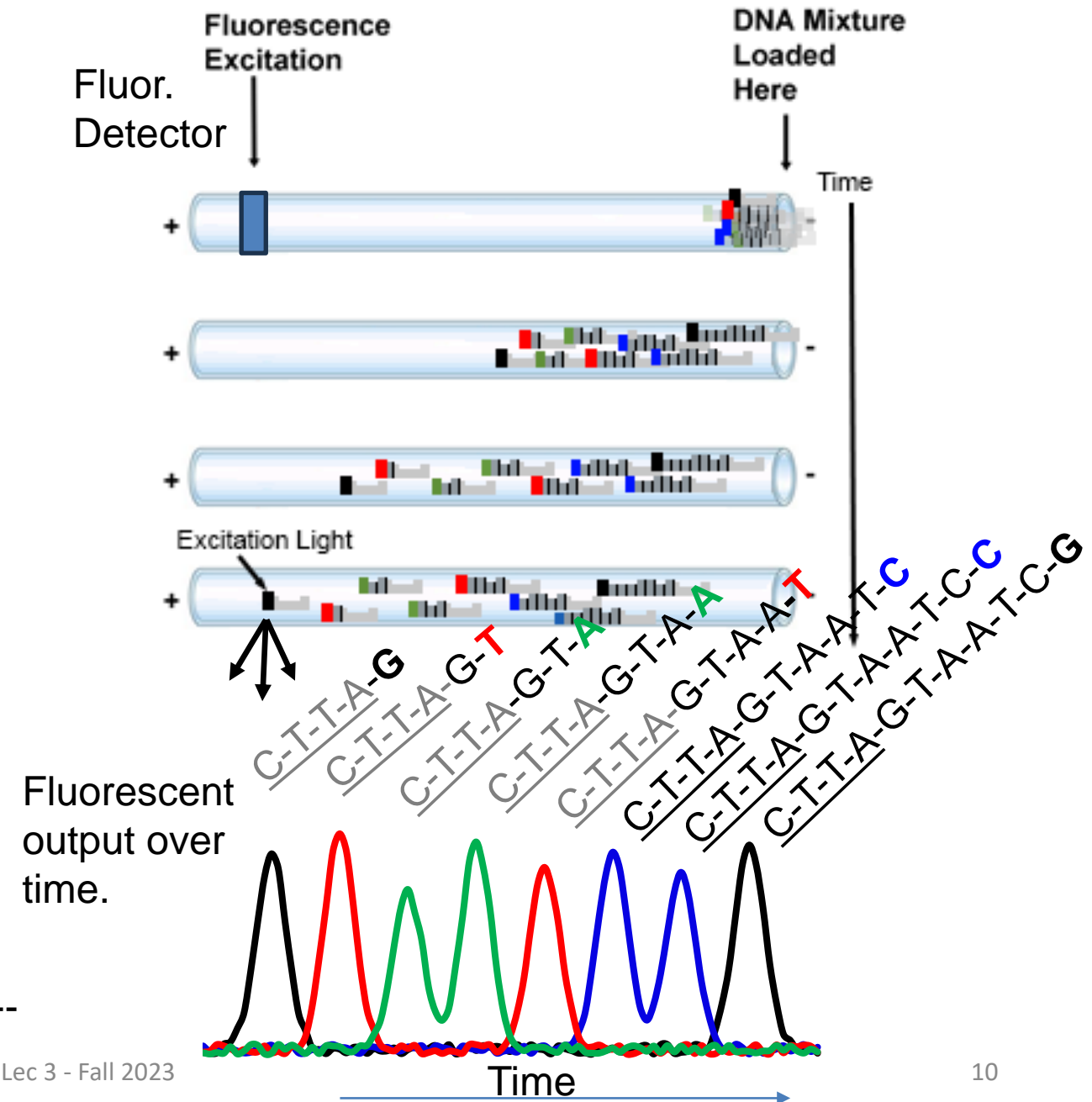


# DNA Sequencing – Analysis of Fragments to Determine Order of Addition

## 4. Capillary Gel Electrophoresis

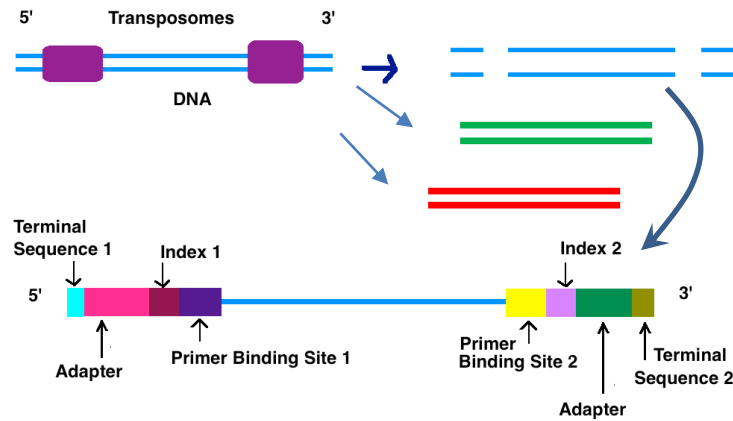
- Migration due to the voltage because of the neg. charge on DNA phosphates
- Separation of DNA molecules by size, smaller travel through gel faster.
- Fragments reach the detector in the order of their length: primer+1 first, primer+2 second, etc.
- At the detector, a laser excites the fluorescence.
- Only fluorescent DNA molecules (terminated with ddNTP) give a signal.
- The color of the emitted fluorescence gives the dideoxy base at the 3' end of the DNA fragment.
- The order of peaks gives the sequence that is complementary to the template (= strand with primer).

5'-C-T-T-A G-T-A-A-T-C-C-G  
3'-A-G-T-C-G-A-A-T-C-A-T-T-A-G-G-C---



# Newer Sequencing Methods-Illumina Dye Sequencing – Next Generation High Throughput

## A. Obtaining the DNA

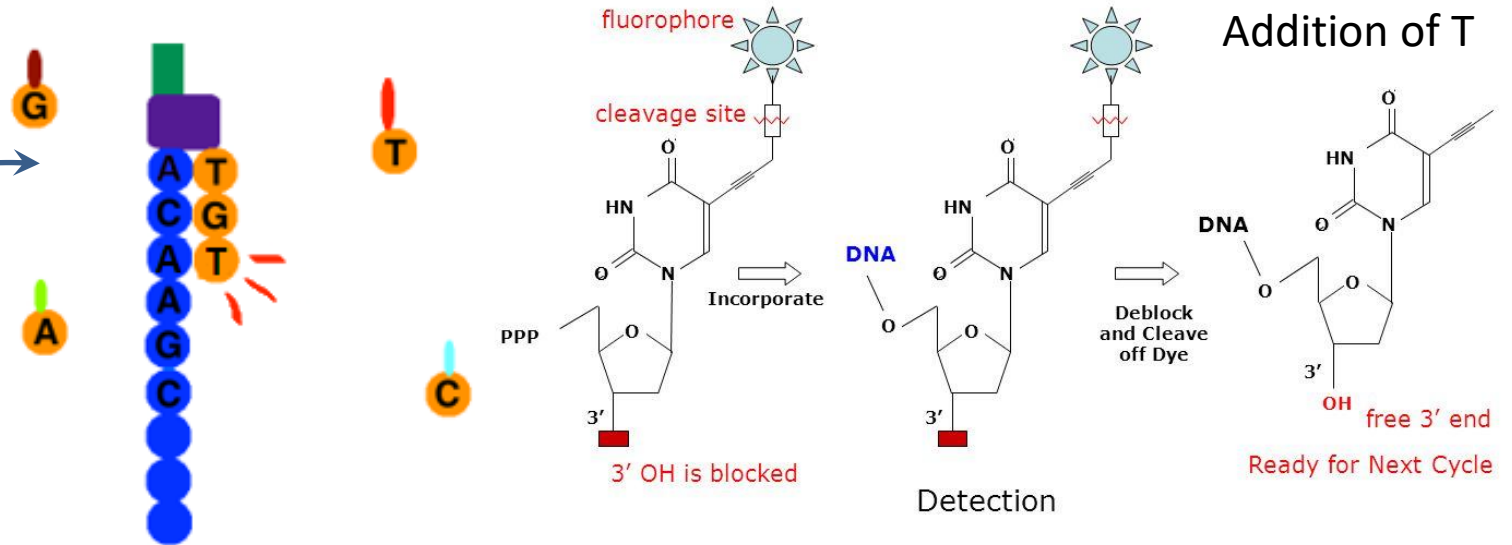


- The entire genome can be sequenced.
- The DNA is fragmented into small 100 base pieces.
- Synthetic DNA is added to the ends (sites for primers for sequencing)
- Different fragments are bound to different location on a chip.
- All fragments are sequenced at the same time on a chip.



Cluster formation

## B. Sequencing by synthesis – Fluorescent labeling & reversible 3'-OH blocking



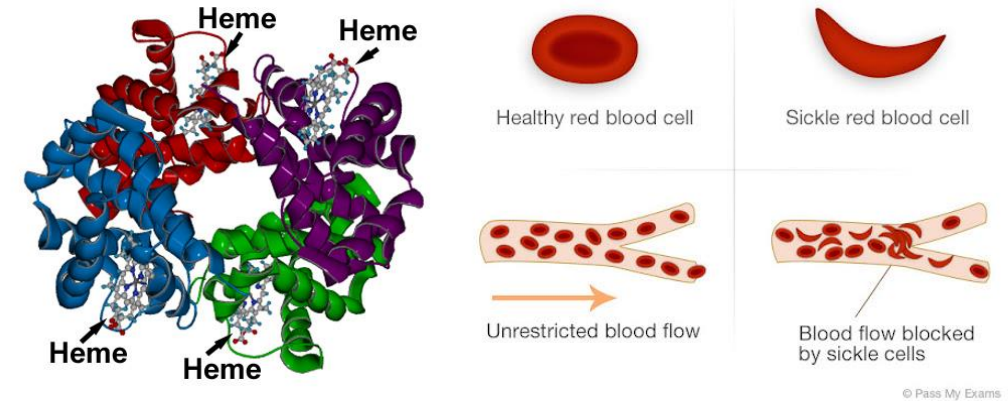
1. Only one base is added at a time (3'-OH is blocked)
2. The base that is added is determined by the color of the fluorescent base.
3. 3'-OH blocking group and the fluorescent group are removed prior to the next addition. ~100 cycles can be performed.

By DMLapato - Own work, CC BY-SA 4.0,  
<https://commons.wikimedia.org/w/index.php?curid=43777596>

Method	Read Length	Samples Processed
Sanger	~1000	1
Illumina	~100	~1000s

# Genotyping at the Molecular Level with DNA Sequencing.

- Sickle cell anemia is caused by a single mutation in the beta chain of hemoglobin
- This mutation causes the hemoglobin to form long polymers that distort the shape of the red blood cell.
- Determining whether someone has the mutation can be useful for treatment.

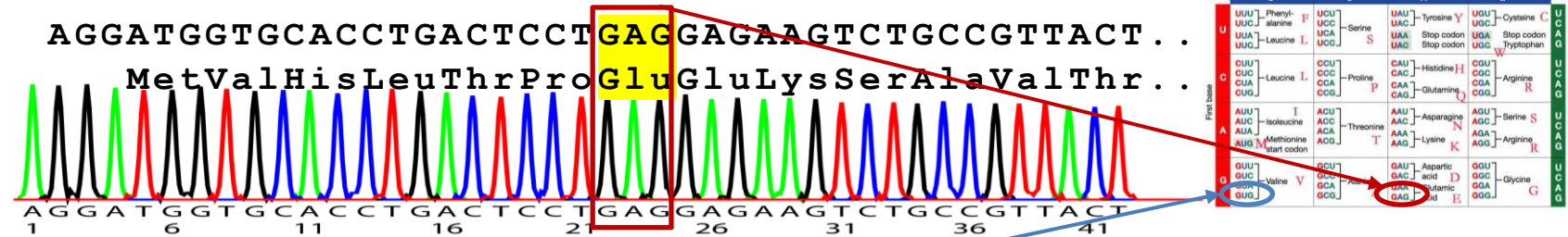


The 5' end of the Hb gene is shown on the right (ATG=start). Using **GGTGCCAG** as a sequencing primer gives the following sequences for the normal and mutant genes:

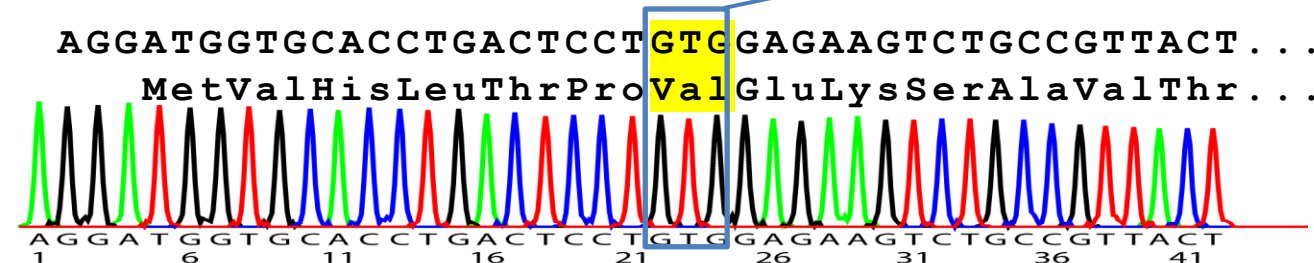
First dd-base added by polymerase

**GGTGCCAGAGGATG**GTGCACCTGACTCCTGAGGAGAAGTC...  
CCACGGTCTCCTACCACGTGGACTGAGGACTCCTCTTCAG...

The sequencing data for the normal beta chain is:



The sequencing data for the mutation with sickle cell is:



False color code:

A=Green  
G=Black  
T=Red  
C=Blue

# Sequencing Summary & Expectations

## Sanger Sequencing:

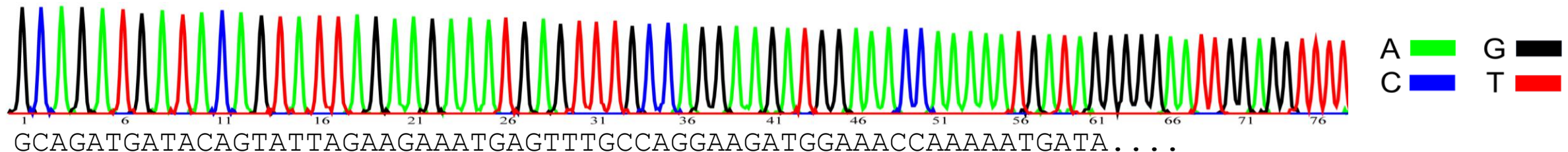
- Gives the sequence that is complementary to the template strand = “top” strand, same strand at the primer.
- The start of the sequencing information is defined by a primer that anneals to the template (therefore some of the sequence has to be known, how this is done will be described later)
- Dideoxy sequencing is carried out by adding both dideoxynucleotide triphosphates (ddNTPs) and deoxyribonucleotide triphosphates (dNTPs) to the synthesis reactions, at a ratio of 1:100. Most growing chains do not terminate.
- ddNTPs are identical to dNTPs except that they lack the 3' hydroxyl group. Because of the missing 3'-OH, DNA polymerization stops once one ddNTP is added to a growing strand.
- The type of the added base is determined by “color coding” each base.
- The location of added bases is determined by measuring the size of the DNA fragment that was terminated by the ddNTP.
- It is possible to sequence approximately 1000 bases by this method.

## Next Gen-Sequencing:

- Simultaneous sequencing of a large number of fragments
- Shorter “reads” 100 versus 1000 bases/template

## Expectations:

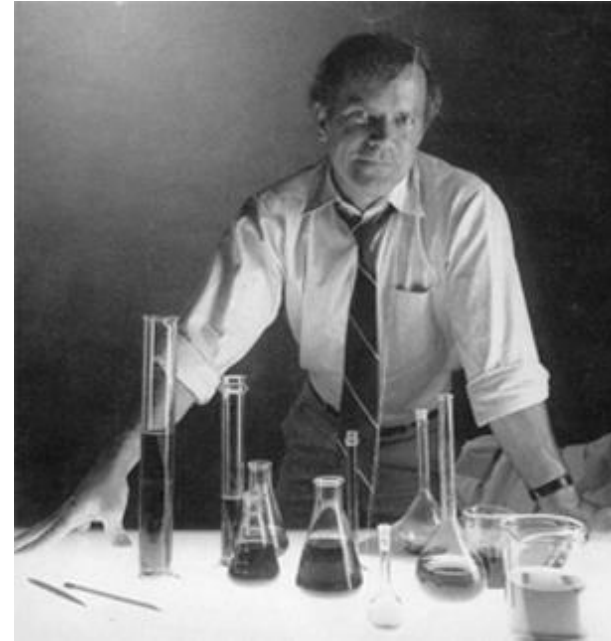
- Can you explain how the colored peaks are generated by random termination using dideoxybases in Sanger Sequencing?
- Can you compare and contrast major features of Sanger and Next-Gen Sequencing?





# Polymerase Chain Reaction -PCR

- In 1983, Kary Mullis developed the molecular biology technique that has since revolutionized genetic research, earning him the Nobel Prize in 1993.
- PCR had an impact on four main areas of biotechnology: gene mapping, cloning, DNA sequencing, and gene detection (e.g. coronavirus).
- PCR is now used as a medical diagnostic tool to detect specific mutations that may cause genetic disease, in criminal investigations and courts of law to identify suspects on a molecular level, and in the sequencing of the human genome.

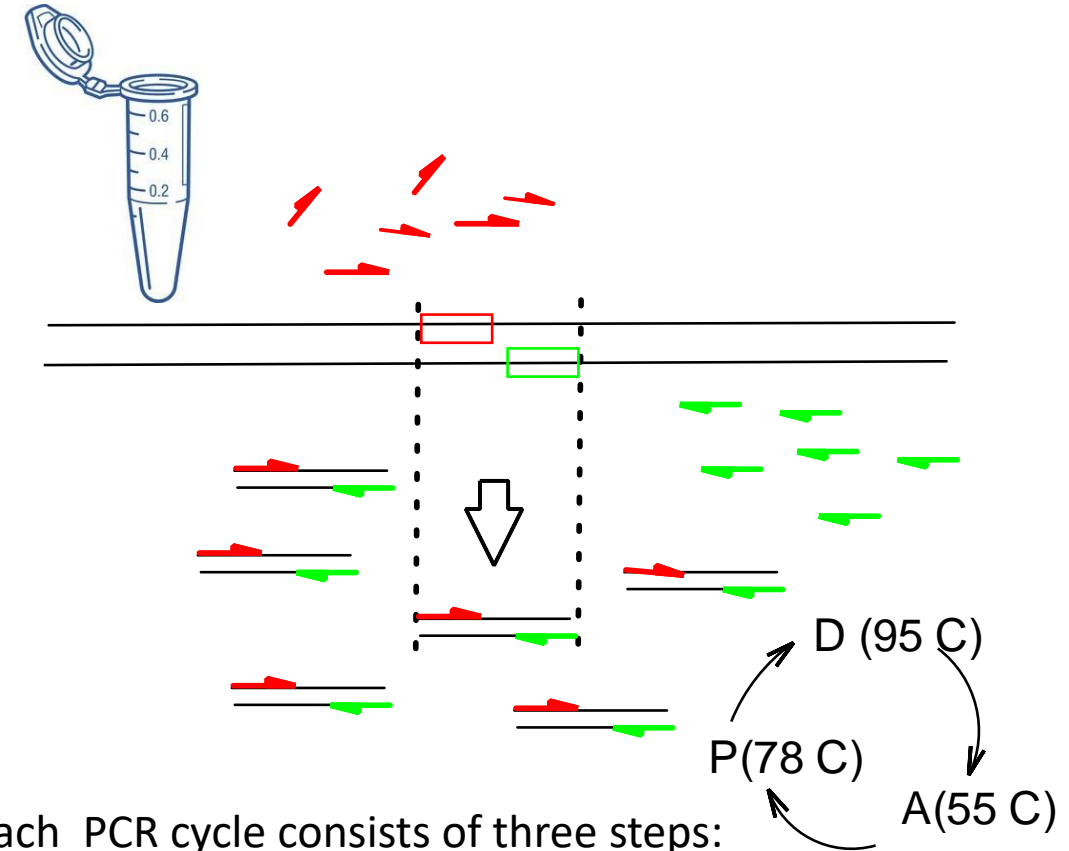


## Expectations:

1. Be able to explain how PCR works to amplify a segment of DNA.
2. Be able to give the left and right primers.
3. Apply PCR approaches to determine genotype and detection of viruses.

# Polymerase Chain Reaction

- PCR is an *in vitro* DNA synthesis reaction in which a specific section of DNA is replicated over and over generating exponentially large amounts of a specific piece of DNA from trace amounts of starting material (template).
- Template can be trace amounts of DNA from a drop of blood, a single hair follicle, or a cheek cell.
- The region of DNA that is copied is specified by the sequence of two primers, which are short ssDNA that initiate polymerase activity. The primers are in vast excess over the DNA.
- The location of the amplified segment is *defined* by two primers (**left = upstream, right = downstream**):
  - they anneal to their templates according to Watson-Crick pairing rules (A-T, G-C),
  - initiate polymerization from those sites,
  - they are incorporated into the final PCR product.
- **Left primer = sequence of top strand at left boundary**
- **Right primer = sequence of bottom strand at right boundary**
- **The primers are DNA and are synthesized chemically, they can be any desired sequence.**
- If there is no homology between the primers and the input DNA, then no PCR product will be formed.



Each PCR cycle consists of three steps:

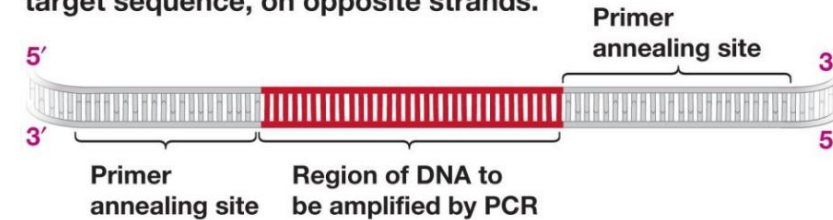
1. Denaturation of the DNA to make it single stranded (2 min at 98 C)
2. Lowering of temperature to let the primers form double-stranded DNA (1 min at 55 C)
3. Elongation by DNA polymerase (1 min/kb at 78 C)

# PCR – Primer Design

- Before a region of DNA can be amplified, one must identify and determine the sequence of a piece of DNA upstream and downstream of the region of interest.
- These areas are then used to determine the sequence of oligonucleotide primers that will be synthesized and used as starting points for DNA replication.
- Primers are complimentary to the up- and down-stream regions of the sequence to be amplified, so they stick, or anneal, to those regions.
  - Left primer = sequence of top strand on the left. This primer will anneal to the bottom strand.
  - Right primer = sequence of bottom strand on the right. This primer will anneal to the top strand.
- Primers are in large excess over the template DNA, they are never used up and they are incorporated into the final PCR product.

Note: Actual primer lengths are 20-30 bases, in the illustrations here and on problem sets, much shorter primers are used.

(a) PCR primers must bind to sequences on either side of the target sequence, on opposite strands.



(b) When target DNA is single stranded, primers bind and allow DNA polymerase to work.



Amplified region

5' --AAG CTGAC TAGTCGATGCGAATGTGCGGTGC--3'  
3' --TTCGACTGATCAGCTACGCTT ACACG CCACG--5'

5' CTGAC

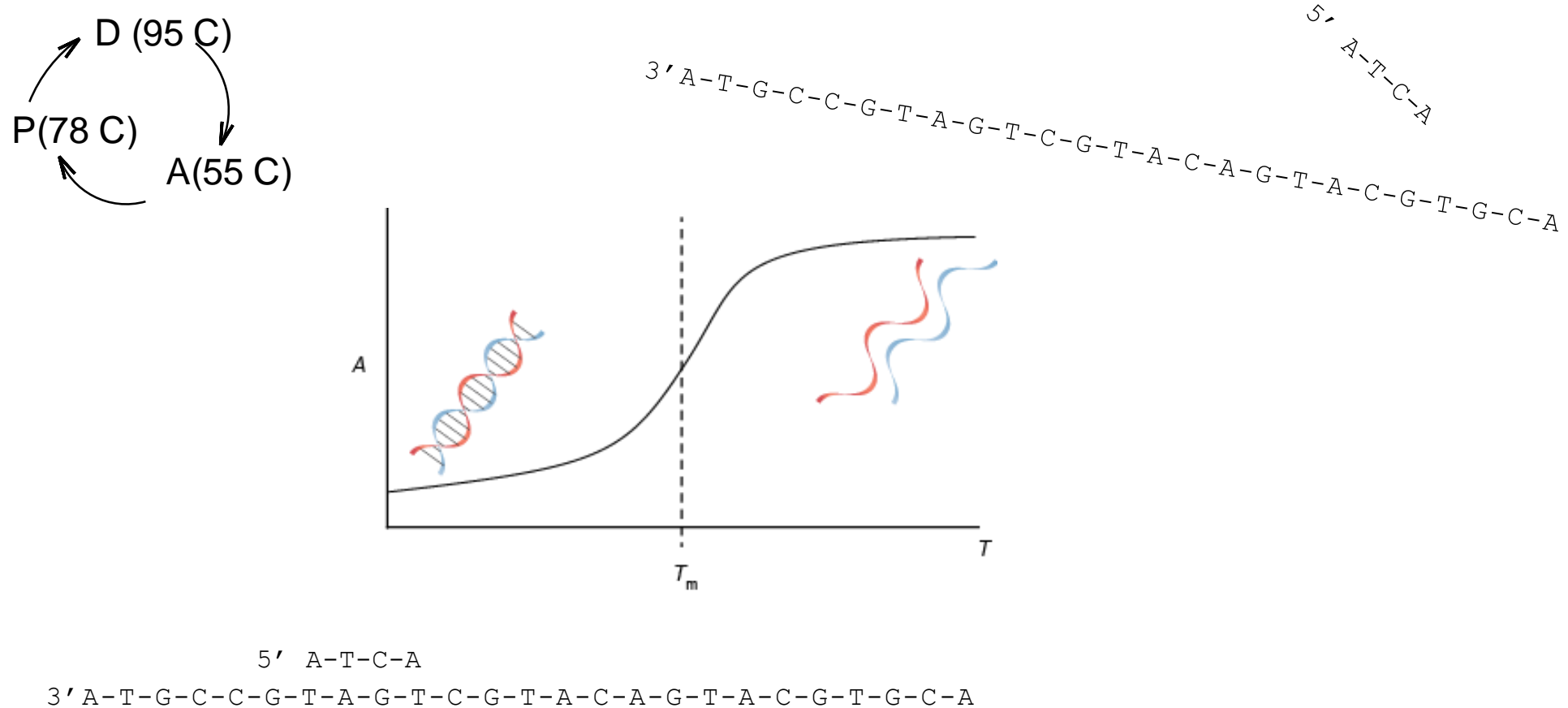
ACACG5' = 5' GCACA

CTGACTAGTCGATGCGAATGTGC  
GACTGATCAGCTACGCTTACACG

Know these rules!



# PCR Step 1 - Thermal Stability of Double Stranded DNA (dsDNA)

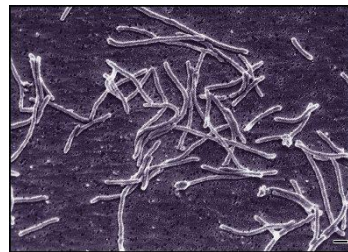


# PCR Step 1 - Thermostable Polymerases

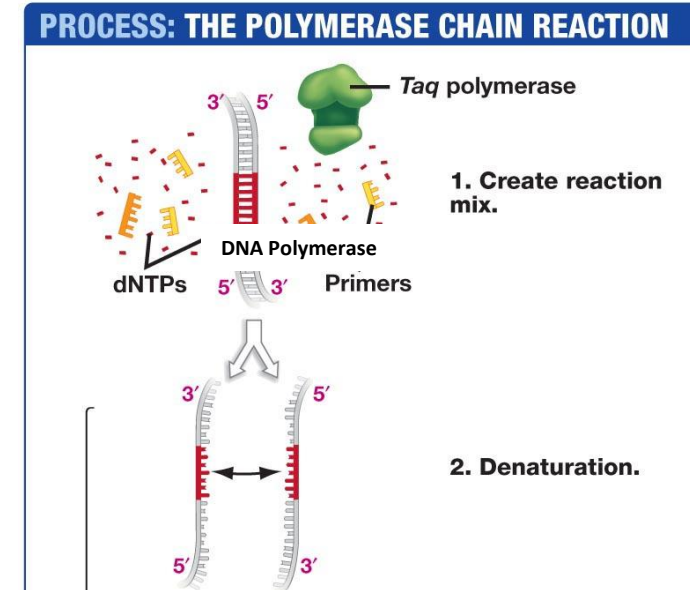
- If we heat up the DNA to temperatures high enough that it denatures into single stranded form, (temperatures of between 60°C and 94°C) what will happen to our DNA polymerases?
- Most DNA polymerases are destroyed at this high temperature.
- How can we synthesize DNA if all of our DNA polymerases are destroyed?
- Utilize a thermostable polymerase



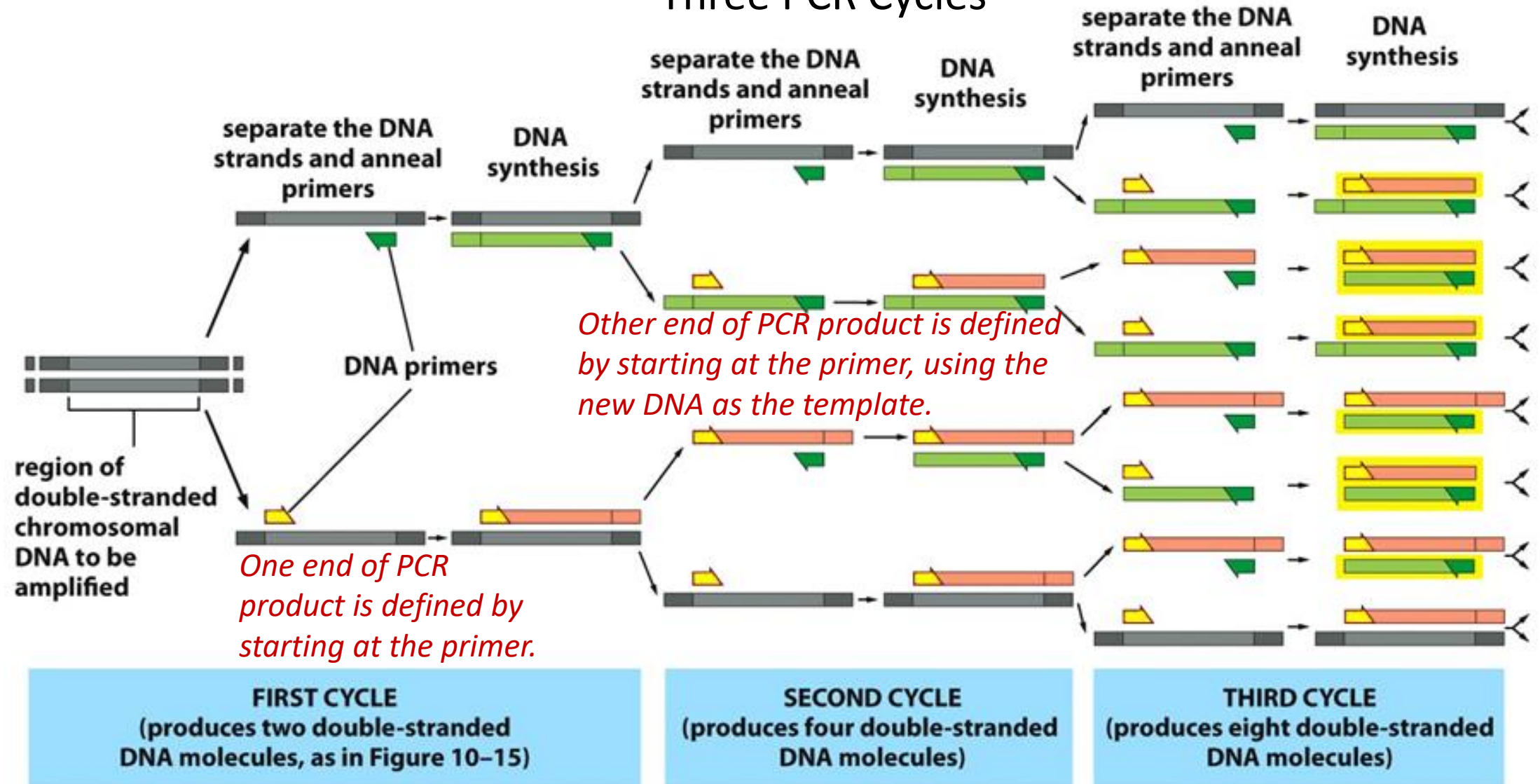
*Thermus Aquaticus*



[http://www.mun.ca/biology/scarr/Thermus\\_aquaticus.html](http://www.mun.ca/biology/scarr/Thermus_aquaticus.html)



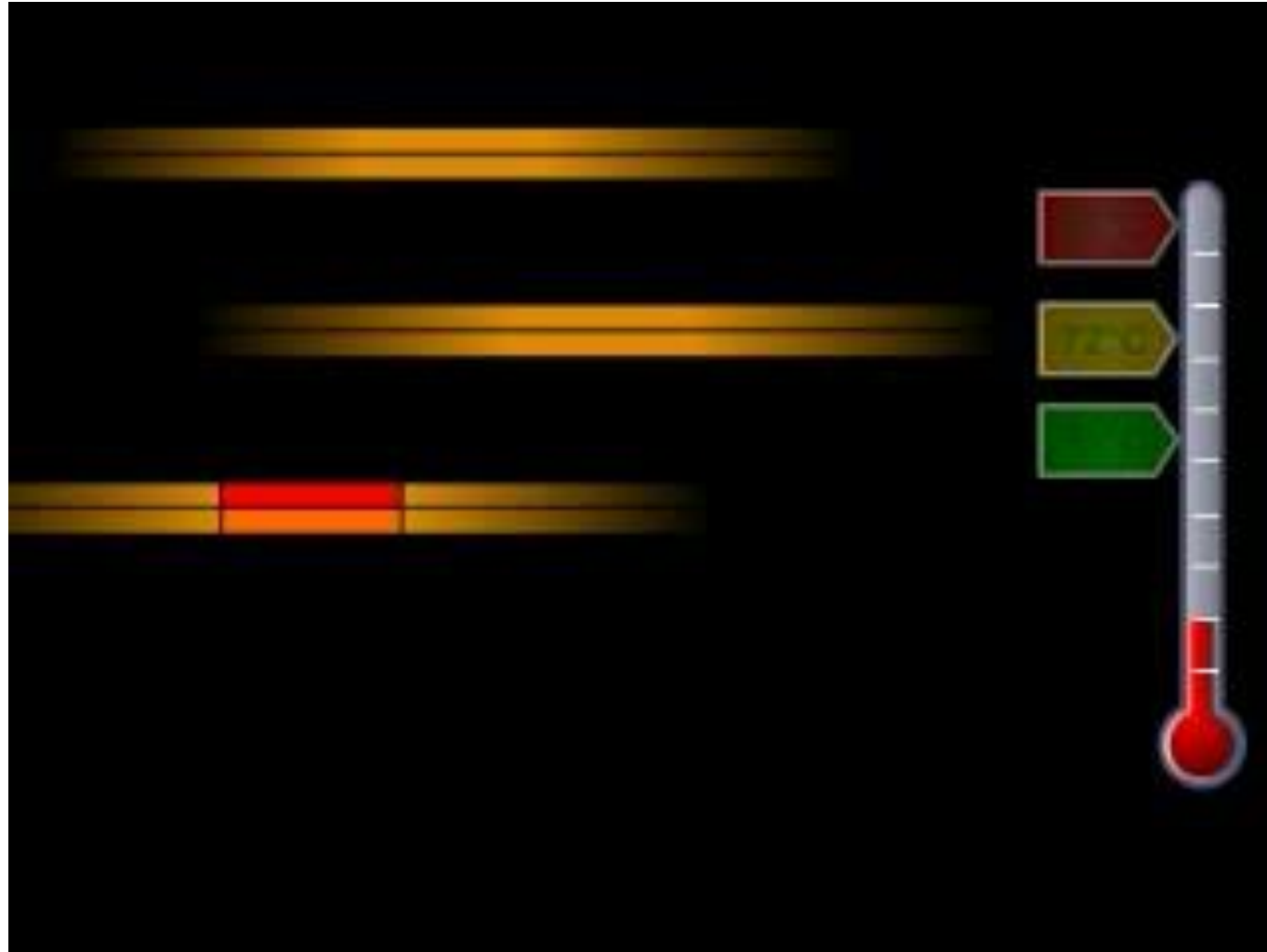
# Three PCR Cycles



After 30 cycles there will be  $2^{30}$ , or over 1 billion times more copies than at the beginning!!!

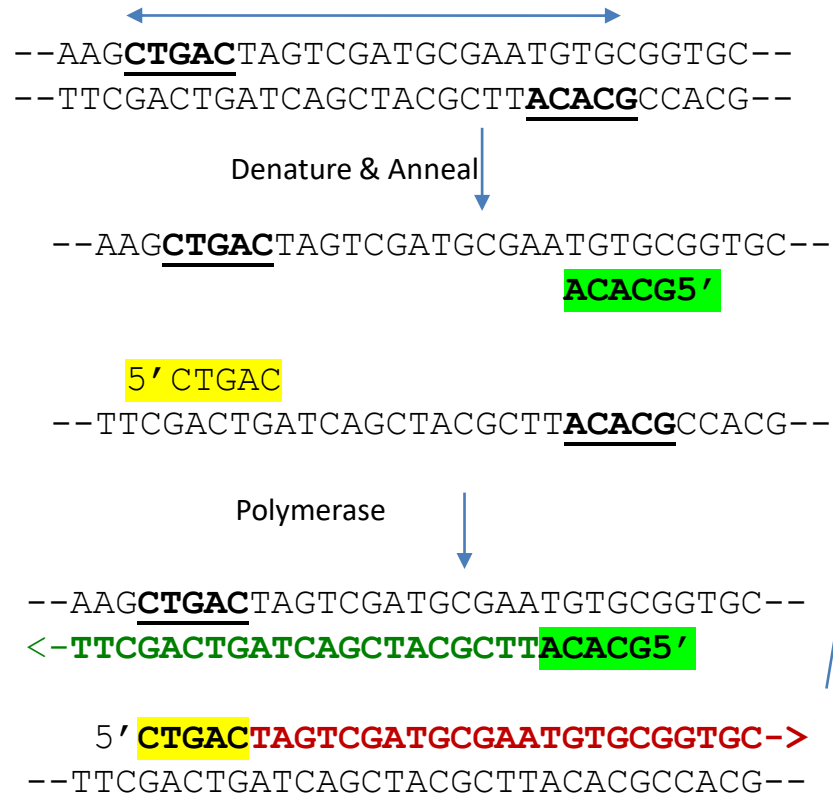
# PCR Animation

Watch Me!



# Detailed Events during first Three PCR Cycles

Cycle I

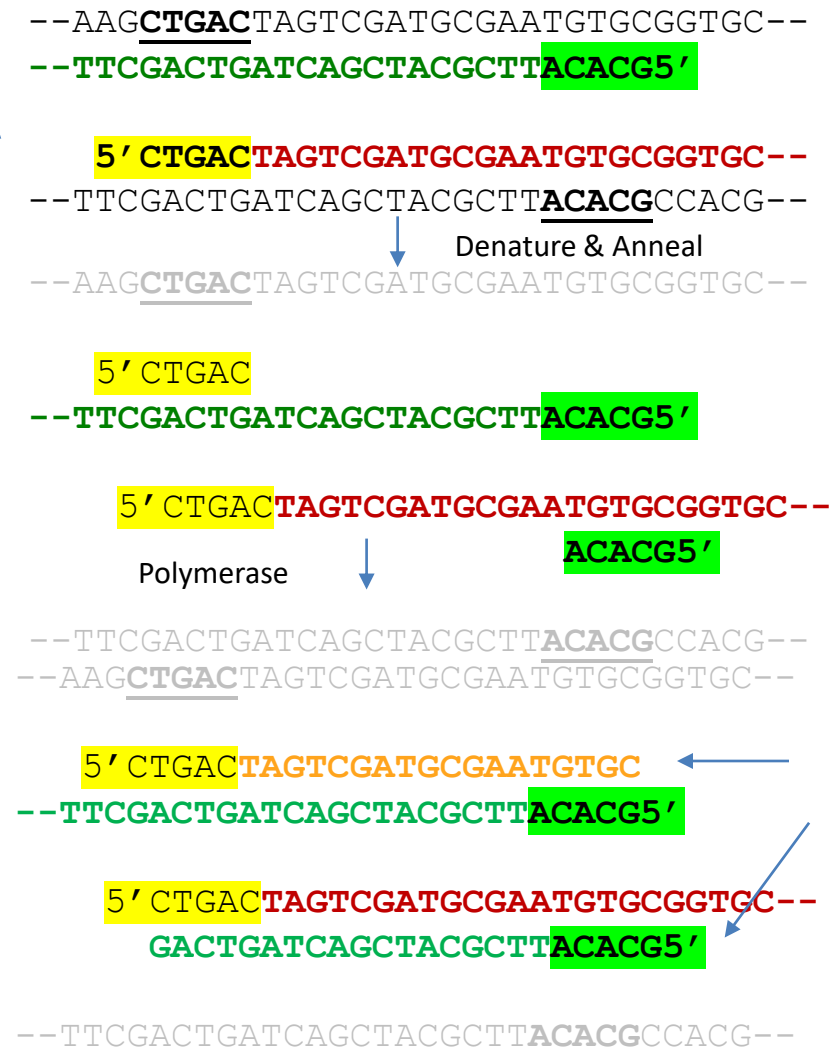


So far - defined one end of the product

Final Product

CTGACTAGTCGATGCGAATGTGC  
 GACTGATCAGCTACGCTTACACG

Cycle II



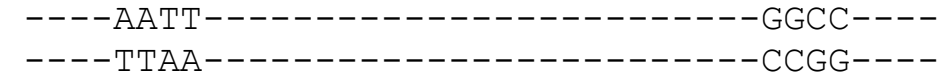
Now have one strand of the product

# Detailed Events during first Three PCR Cycles

Cycle 3



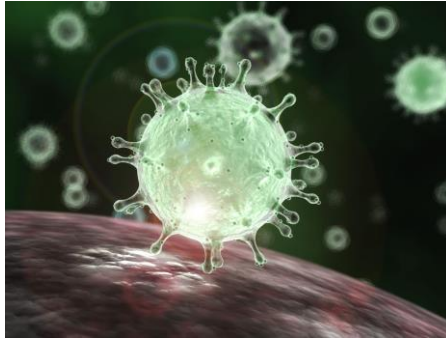
Example – follow the PCR cycles for the following template with primers 5' AATT (left) and 5' GGCC (right)



Now have complete PCR product. This is doubled in each of the following cycles. Note that the primers are the first bases at the beginning of each strand.

# PCR & Detection of Viruses

## Coronavirus



## Sequence of Covid-19 (top strand only)

```

1      attaaaggtt tataccttcc caggtaacaa accaaccaac tttcgatctc ttgtagatct
61     gttctctaaa cgaactttaa aatctgtgtg gctgtcactc ggctgcatgc ttagtgcaact
121    cacgcagtat aattaataac taattactgt cgttgacagg acacgagtaa ctcgtctatc
181    ttctgcaggc tgcttacggg ttctgtccgtg ttgcagccga tcatcagcac atctagggtt

28261  cgaacaaact aaaatgtctg ataatggacc ccaaaatcag cgaaatgcac cccgcattac
28321  gtttggtgga ccctcagatt caactggcag taaccagaat ggagaacgca gtggggcgcg
28381  atcaaaacaa cgtcggcccc aaggtttacc caataatact gcgtcttggg tcaccgctct
28441  cactcaacat ggcaaggaag accttaaatt ccctcgagga caaggcggtc caattaacac

29701  gggaggactt gaaagagcca ccacattttc accgaggcca cgcggagtac gatcgagtgt
29761  acagtgaaca atgctaggga gagctgccta tatggaagag ccctaattgtg taaaattaat
29821  tttagtagtg ctatcccat gtgattttaa tagcttctta ggagaatgac aaaaaaaaaa
29881  aaaaaaaaaa aaaaaaaaaa aaa.
    
```

## CDC Recommended PCR Primers

2019-Novel Coronavirus (2019-nCoV) Real-time rRT-PCR Panel Primers and Probes				
Name	Description	Oligonucleotide Sequence (5'>3')	Label <sup>1</sup>	Working Conc.
2019-nCoV_N1-F	2019-nCoV_N1 Forward Primer	5'-GAC CCC AAA ATC AGC GAA AT-3'	None	20 µM
2019-nCoV_N1-R	2019-nCoV_N1 Reverse Primer	5'-TCT GGT TAC TGC CAG TTG AAT CTG-3'	None	20 µM

*dsSeq of above bold & circled region*

28271 aaaatgtctgataatg **GACCCAAAATCAGCGAAAT**gcaccccgcatctacgttttgggtggaccctcagattcaactggcagtaaccagaatggagaacgca  
 ttttacagactattacctgggggttttagtcgctttacgtggggcgtaatgcaaaccacctggga **GTCTAAGTTGACCGTCATTGGTCT**tacctcttgcgt

## PCR Product

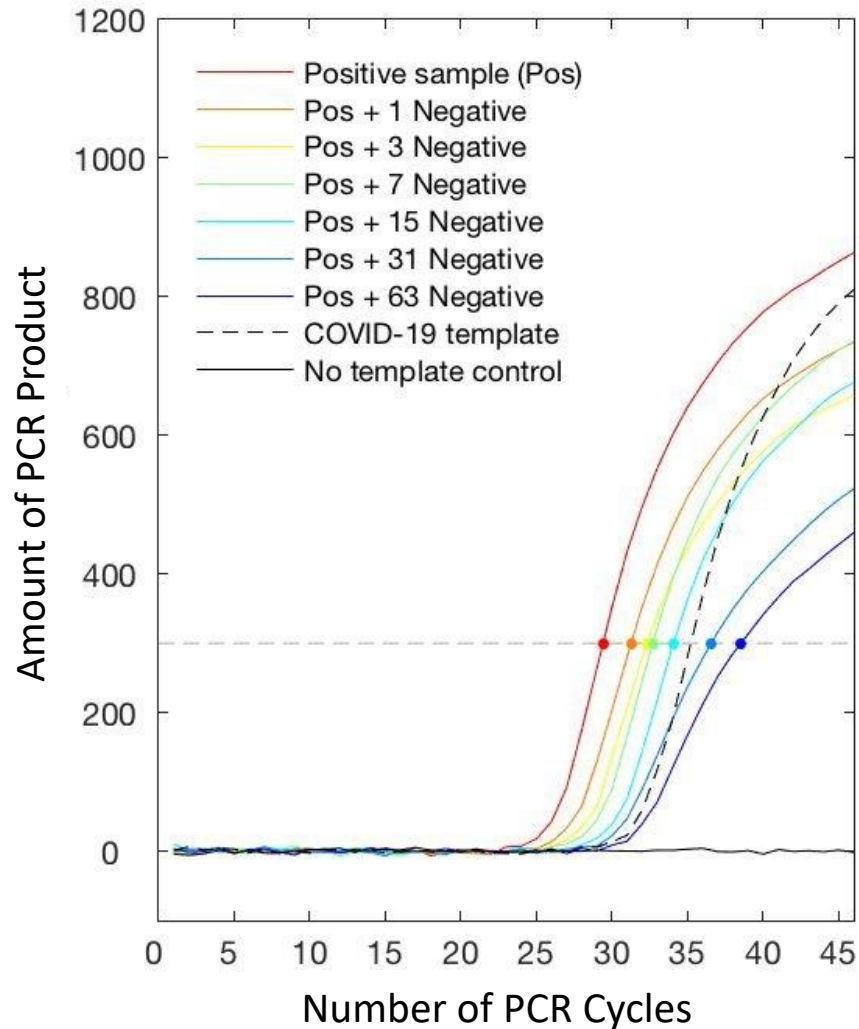
**GACCCAAAATCAGCGAAAT**GCACCCCGCATTACGTTTGGTGGACCCTCAGATTCAACTGGCAGTAACCAGA  
 CTGGGGTTTTAGTCGCTTTACGTGGGGCGTAATGCAAACCACCTGGGA**GTCTAAGTTGACCGTCATTGGTCT**

*Will PCR generate products if the viral DNA is not present?*



# Covid 19 PCR Test: Detection of the PCR Product.

<https://www.medrxiv.org/content/10.1101/2020.03.26.20039438v1>

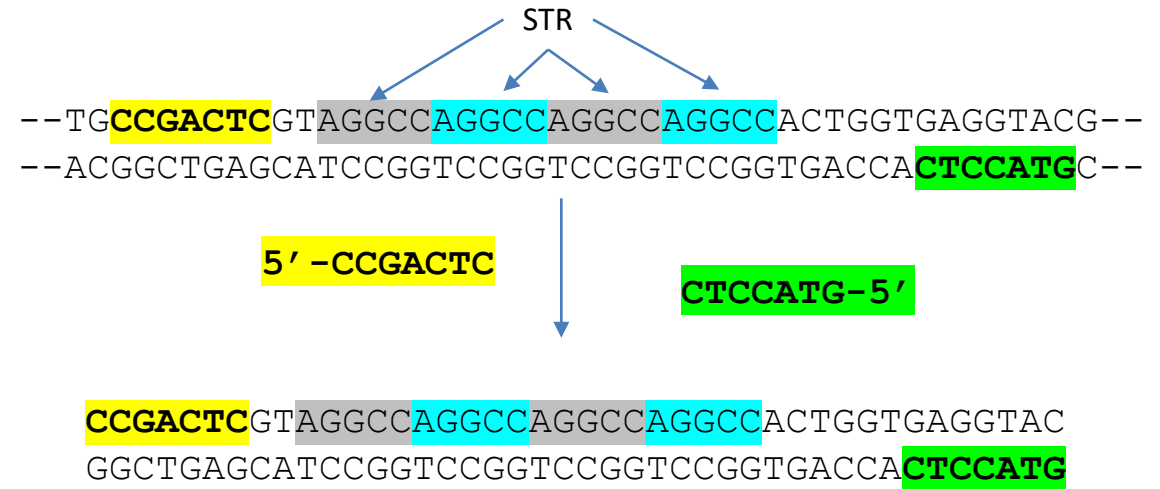


- Production of PCR products (double stranded DNA) causes an increase in signal (fluorescence)
- Dots represent the cross point of the fluorescence threshold (threshold = 300, gray dashed line).
- Red curve (Positive sample) shows a threshold level of PCR product after 27 cycles.
- Next 6 samples are the positive sample mixed with up to 63 negative samples, showing that it is possible to test pooled samples.
- - - - is a **positive control** amount of Covid template. It shows that you can detect a PCR product if the covid genome is present.
- Solid black line is a **negative control**, no Covid DNA. It shows that addition of covid template will lead to a signal.



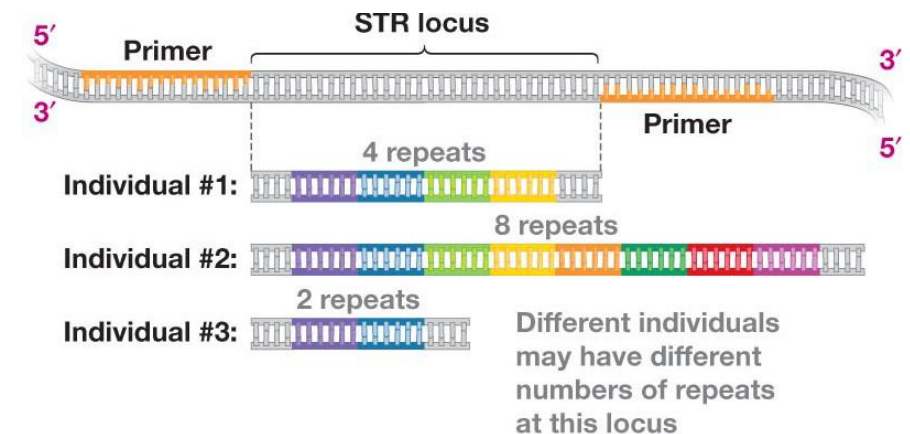
# Application of PCR – Identification of Individuals

- Regions of DNA have variable numbers of repeated DNA sequences (Short tandem repeats, STR) that differ from one person to the next.
- Individuals will inherit one copy of the repeat from each parent. The length of the inherited DNA can be the same or different.
- PCR Primers are designed to be outside the repeated region, so that they will anneal to a single location on the chromosome and then amplify the region containing the STR
- PCR Product length = primer lengths + number of tandem repeats (+ any DNA between the primers and the repeats). *Individuals can be differentiated by the length of the PCR product if they have different numbers of STR*



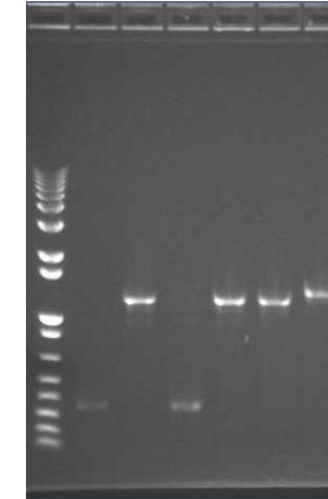
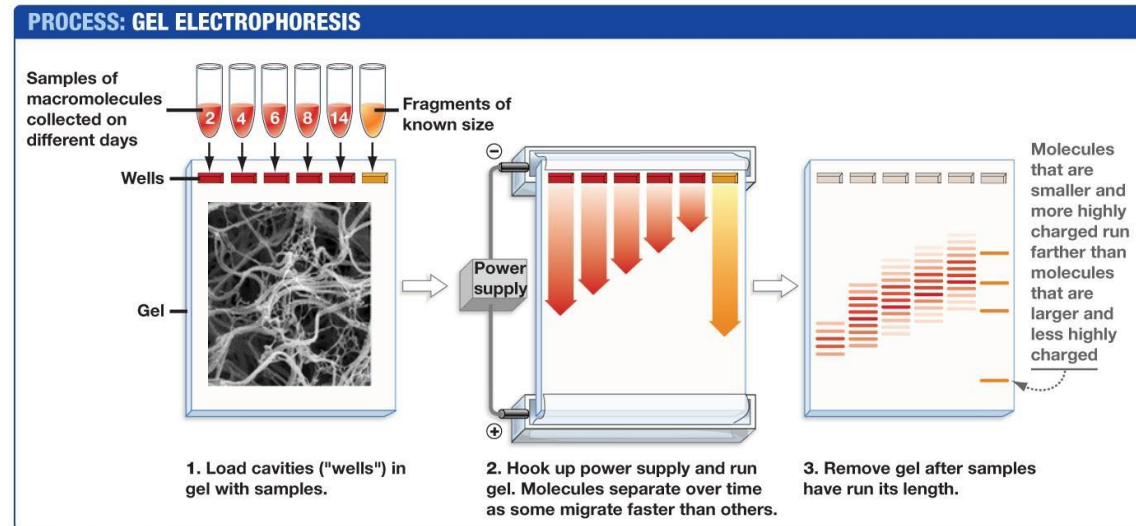
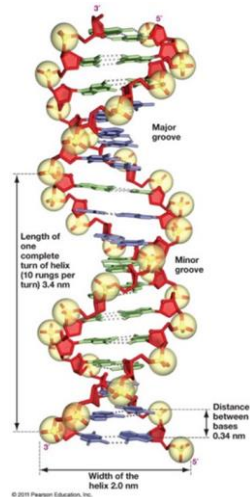
*Which individual has the shortest PCR product?*

*Which has the longest?*

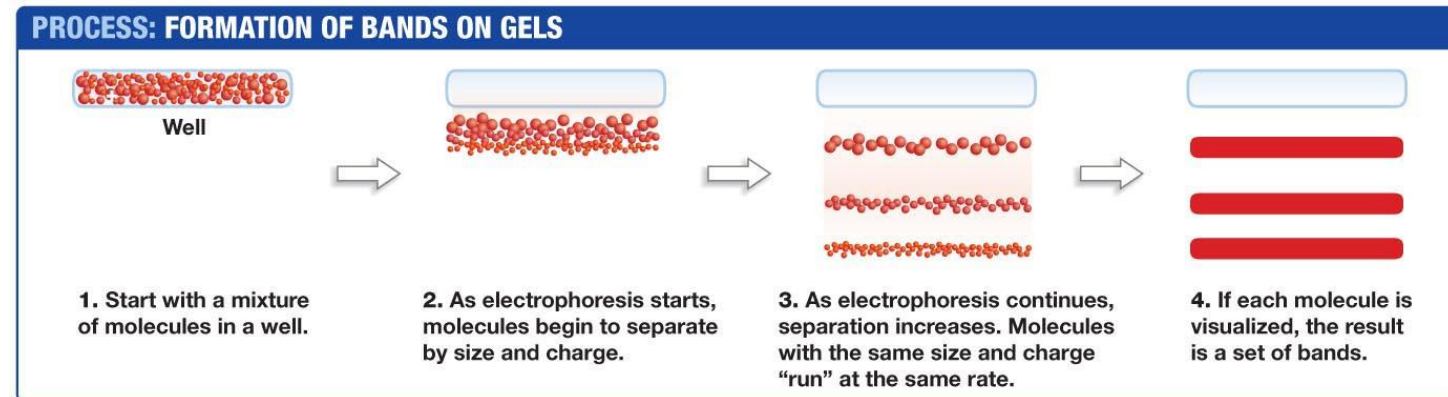


# Size Determination of PCR products - Agarose Gel Electrophoresis.

<https://dnalc.cshl.edu/resources/animations/gelectrophoresis.html>

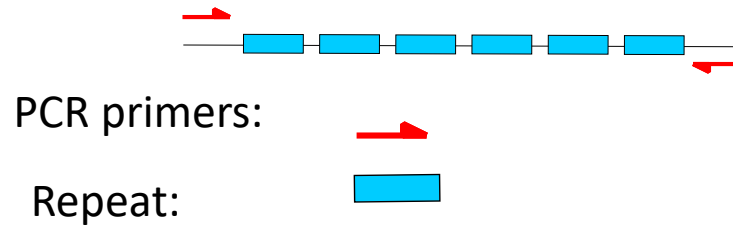


*Which are the smallest PCR fragments?*



# Short Tandem Repeats to Test Paternity

1. DNA samples (blood, cheek cells) would be obtained from:
  - Mother
  - Child
  - Candidate fathers.
2. PCR would be performed using primers that amplify a segment of the chromosome containing repeats.
3. Each individual would show 2 bands on the gel, corresponding to the PCR product from each chromosome (we have two copies of each chromosome).
4. The child would inherit one copy from the mother and the other from the father:
  - One of the child's PCR product would match one of the mothers.
  - The other PCR product from the child would match one of the PCR products from the father.



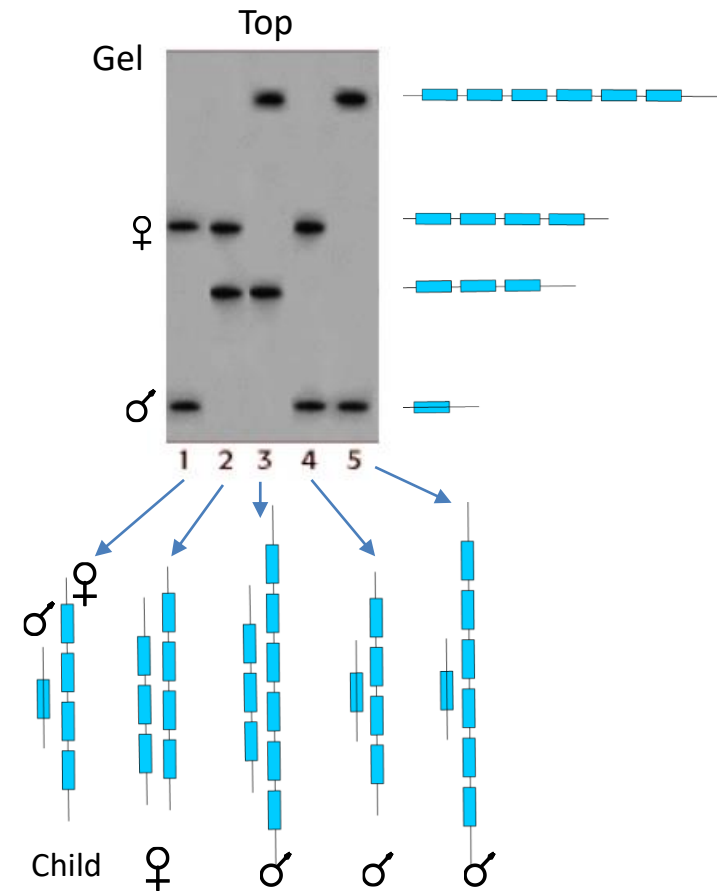
Lane 1: Child

Lane 2: Mother

Lanes 3-5: Possible Fathers

2. Who is not the father?

3. Who *may* be the father?





# Repeat Expansions Related to Diseases

## Chapter 9 - Repeat expansion diseases

Henry Paulson

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<https://doi.org/10.1016/B978-0-444-63233-3.00009-9>

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		Second base		
		U	C	A
t base	U	UUU } Phenylalanine UUC } UUA } Leucine UUG }	UCU } Serine UCC } UCA } UCG }	UAU } Tyrosine UAC } UAA } Stop codon UAG } Stop codon
	C	CUU } Leucine CUC } CUA } CUG }	CCU } Proline CCC } CCA } CCG }	CAU } Histidine CAC } CAA } Glutamine CAG }

- The number of repeats can be detected by:
  - DNA sequencing
  - PCR

- CAG – at least 10 diseases (Huntington disease, spinal and bulbar muscular atrophy, dentatorubral-pallidoluysian atrophy and seven SCAs)
- CGG – fragile X, fragile X tremor ataxia syndrome, other fragile sites (GCC, CCG)
- CTG – myotonic dystrophy type 1, Huntington disease-like 2, spinocerebellar ataxia type 8, Fuchs corneal dystrophy
- GAA – Friedreich ataxia
- GCC – FRAXE mental retardation
- GCG – oculopharyngeal muscular dystrophy
- CCTG – myotonic dystrophy type 1
- ATTCT – spinocerebellar ataxia type 10
- TGGAA – spinocerebellar ataxia type 31
- GGCCTG – spinocerebellar ataxia type 36
- GGGGCC – C9ORF72 frontotemporal dementia/amyotrophic lateral sclerosis
- CCCCGCCCGCG – EPM1 (myoclonic epilepsy)

# Introduction to Immunology

1. Branches of the immune system (Innate and acquired)
2. Properties of antibodies (Quaternary structure, antigen recognition)
3. How antibodies are produced:
  - Genome DNA changes
  - mRNA splicing
  - Cellular export
4. How antibodies eliminate pathogens

## Key Questions:

1. Why is the innate system important?
2. What is the origin of diversity in acquired immunity?
3. How are antibodies made.

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## The Nobel Prize in Physiology or Medicine 2018

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Ill. Niklas Elmehed. © Nobel Media

James P. Allison

Prize share: 1/2



Ill. Niklas Elmehed. © Nobel Media

Tasuku Honjo

Prize share: 1/2

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The Nobel Prize in Physiology or Medicine 2018 was awarded jointly to James P. Allison and Tasuku Honjo "for their discovery of cancer therapy by inhibition of negative immune regulation."

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**Some Important Definitions:**

**Antigen** = something that is recognized by the immune system, e.g. bacteria, virus, pollen.

**Epitope** = the part of the antigen that is contacted by the antibody.

**Antibody** (Ab) = Y-shaped protein that recognizes antigens, found on the surface of B-cells or secreted by plasma cells. When bound to antigen, it can initiate a process that results in the destruction of the antigen. *Specificity is high due to AA sequence in the variable segments.*

**Immunoglobulin (Ig)** = antibody.

**B-cell** = involved in antibody production and recognition of pathogen. Has antibody molecule on its surface (as part of the B-cell receptor). Develops into plasma cells after activation by T<sub>H</sub> cells. Called B-cells because they are generated in the organ called the Bursa in birds.

**Plasma cell** = derived from B-cell after activation of the B-cell, produces secreted antibodies with the *same specificity as the original B-cell.*

**T<sub>H</sub> cell** = T-helper: *Required* to activate both B and T<sub>C</sub> cells, as well as other cells in the immune system. Called T-cells because they mature in the thymus.

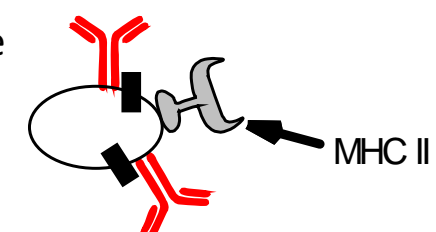
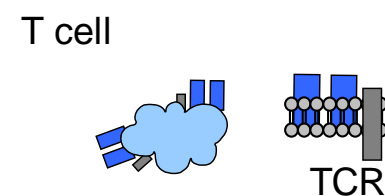
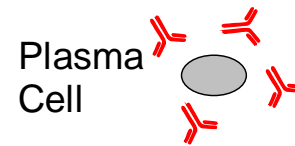
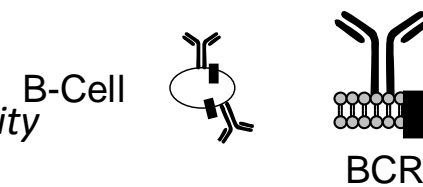
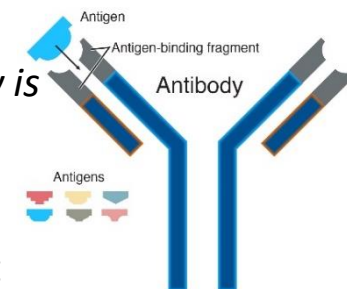
**T<sub>C</sub> cell** = T-cellular: Involved in defense against viruses and cancer.

**TCR** = T-cell receptor – found on the surface of T-cells, recognizes MHC proteins + bound peptide, RTK.

- **T<sub>C</sub> cell** = recognizes MHC I + peptide
- **T<sub>H</sub> cell** = recognizes MHC II + peptide

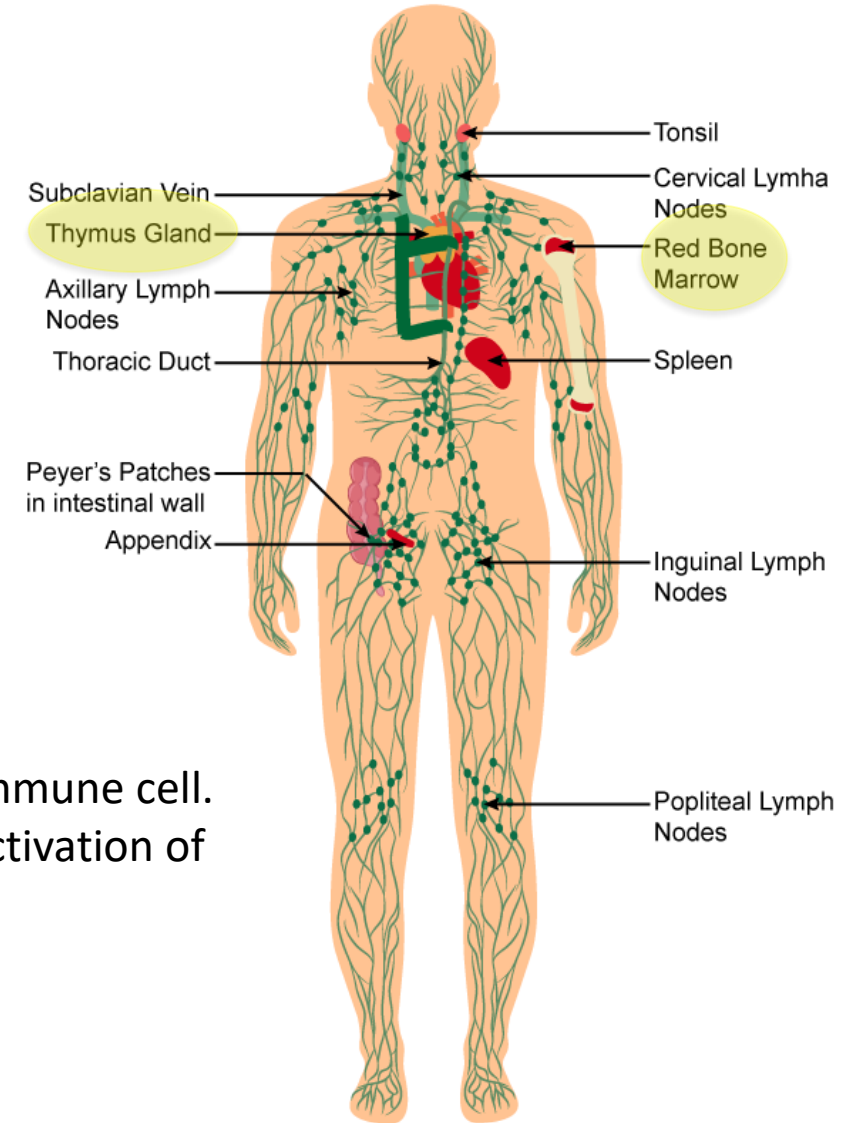
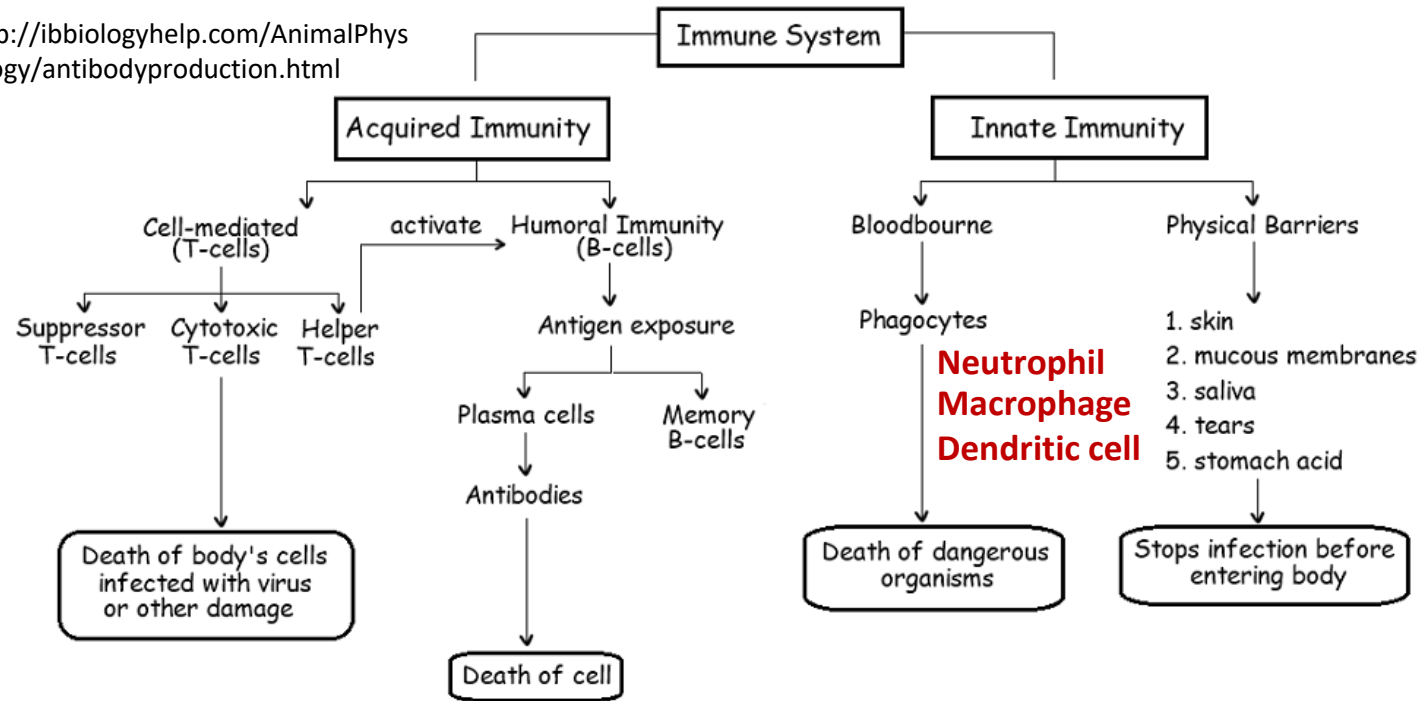
**MHC** = major histocompatibility complex – required for acquired immunity (basis of transplant rejection)

- MHC I = protein found on the surface of **all** cells, “presents” peptides derived from the proteins that were made by the cell. The MHC-peptide complex is recognized by T<sub>C</sub> cells. **Only foreign peptides produce a response.**
- MHC II = on the surface of B-cells, macrophages, and dendritic cells. Presents external peptides to T<sub>H</sub> cells, leading to activation of the cell by T<sub>H</sub> cells. **Only foreign peptides produce a response.**



# Branches of the Immune System:

<http://ibbiologyhelp.com/AnimalPhysiology/antibodyproduction.html>



<https://www.topperlearning.com/>

Important **primary** lymphatic organs: bone marrow (B), thymus (T)-Generate all immune cell.  
Important **secondary** lymphatic organs: lymph nodes, spleen, Peyer's patches – Activation of immune cells.

## Why is the innate system essential?

- A pathogen doubles every hour.
- It takes 7 days to produce antibody (1<sup>st</sup> exposure)
- How many bacteria would be present if they grew uncontrolled for 7 days ( $=2^{24 \times 7}$ )

(there are approximately  $10^{13}$  cells in the human body)

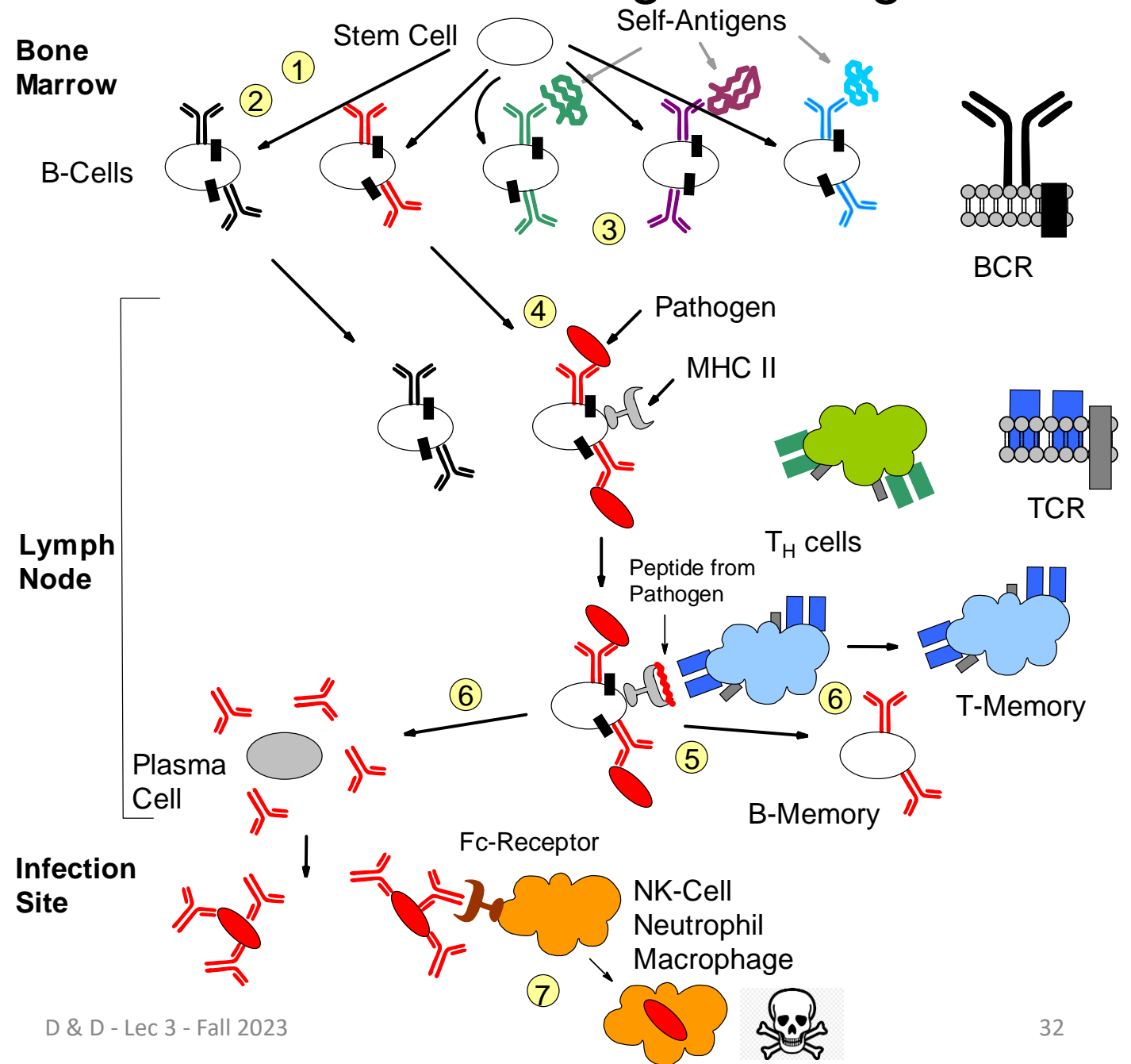
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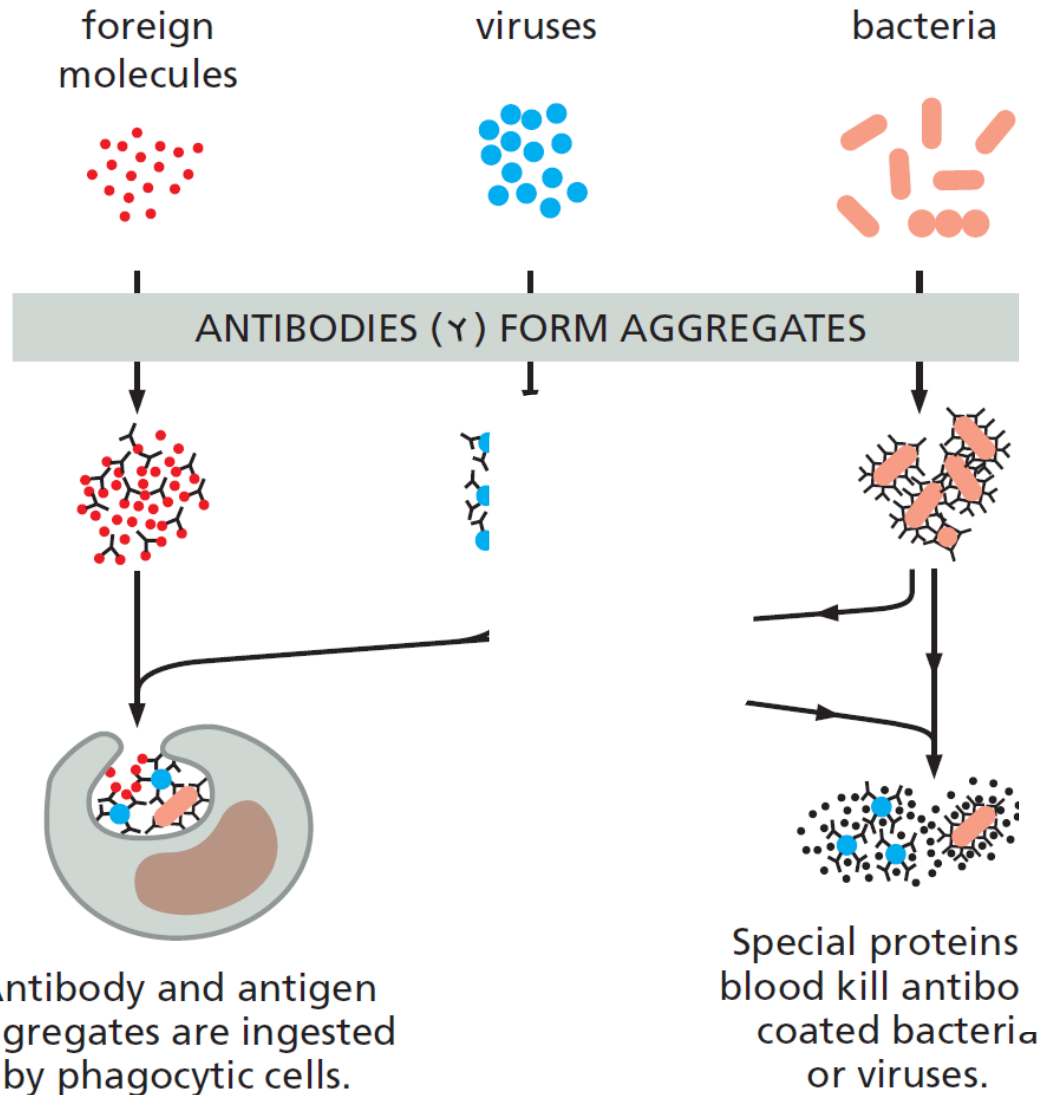
# B-Cell Biology - From Stem Cells to Pathogen Killing.

1. Generation of high diversity of chains during development of stem cells to B-cells in bone marrow.
    - **DNA rearrangements** to generate functional exons for variable segments of both light and heavy chain.
  2. Molecular & cellular biology of **membrane bound antibodies** on cell surface = B-cell receptor (BCR)
    - Transcriptional enhancers, mRNA splicing
    - Light chain and heavy chain exported to surface of B-cells.
  3. **Self tolerance** test to prevent autoimmune diseases, autoreactive B-cells eliminated.
  4. Encounter and **capture of antigen** in lymph nodes
  5. Activation of **B-cells by T<sub>H</sub> cells**
    - Peptides from pathogen presented on major histocompatibility proteins (MHC II).
    - T-cell activation by tyrosine kinase receptors (T-cell Receptor, TCR), secretion of signaling molecules.
  6. Development of
    - **Plasma cells** - Production of soluble antibodies of the same specificity as the parent B-cell.
    - **B-memory** cells (basis of immunity)
    - **T-memory** cells (basis of immunity)
  7. Destruction of Pathogens
    - Fc region of antibody binds to Fc Receptor on NK cells, neutrophils, macrophages
    - Pathogen internalized and destroyed.
- BCR** – B-cell receptor = antibody + signaling chains.  
**TCR** – T cell receptor = MHC-peptide recognition + signaling.

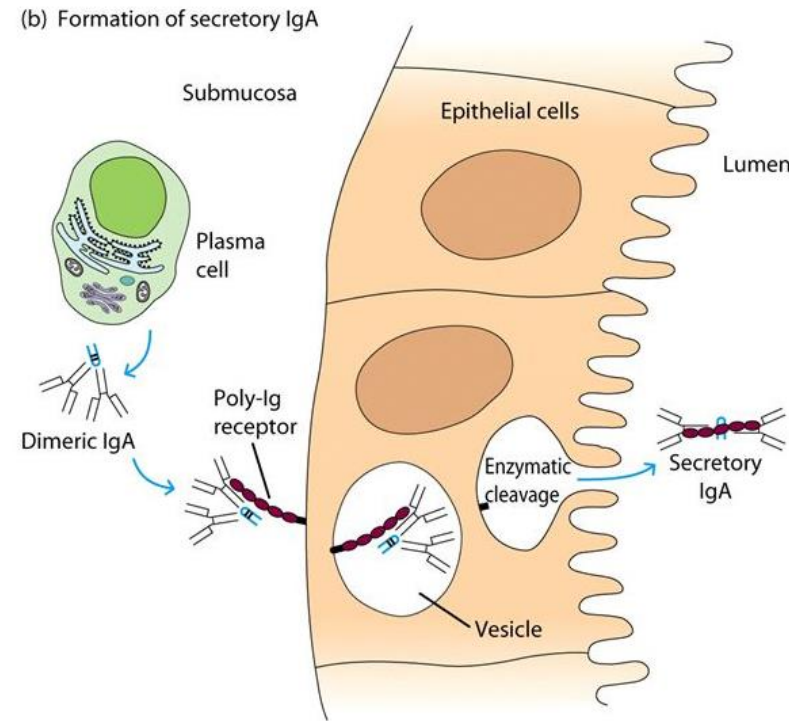




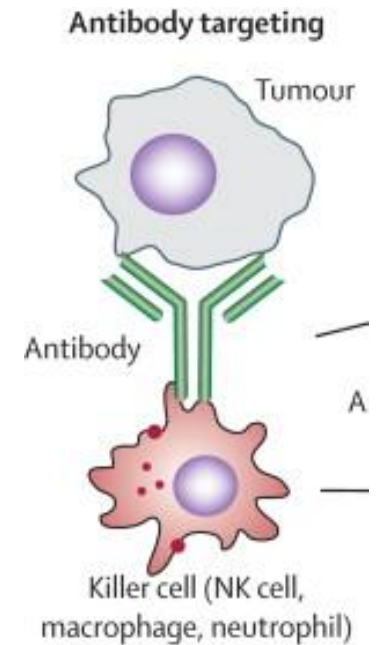
# Antibodies Inactivate Pathogens by Many Mechanisms



Antibodies can be Secreted Outside the Body (10 g/day).



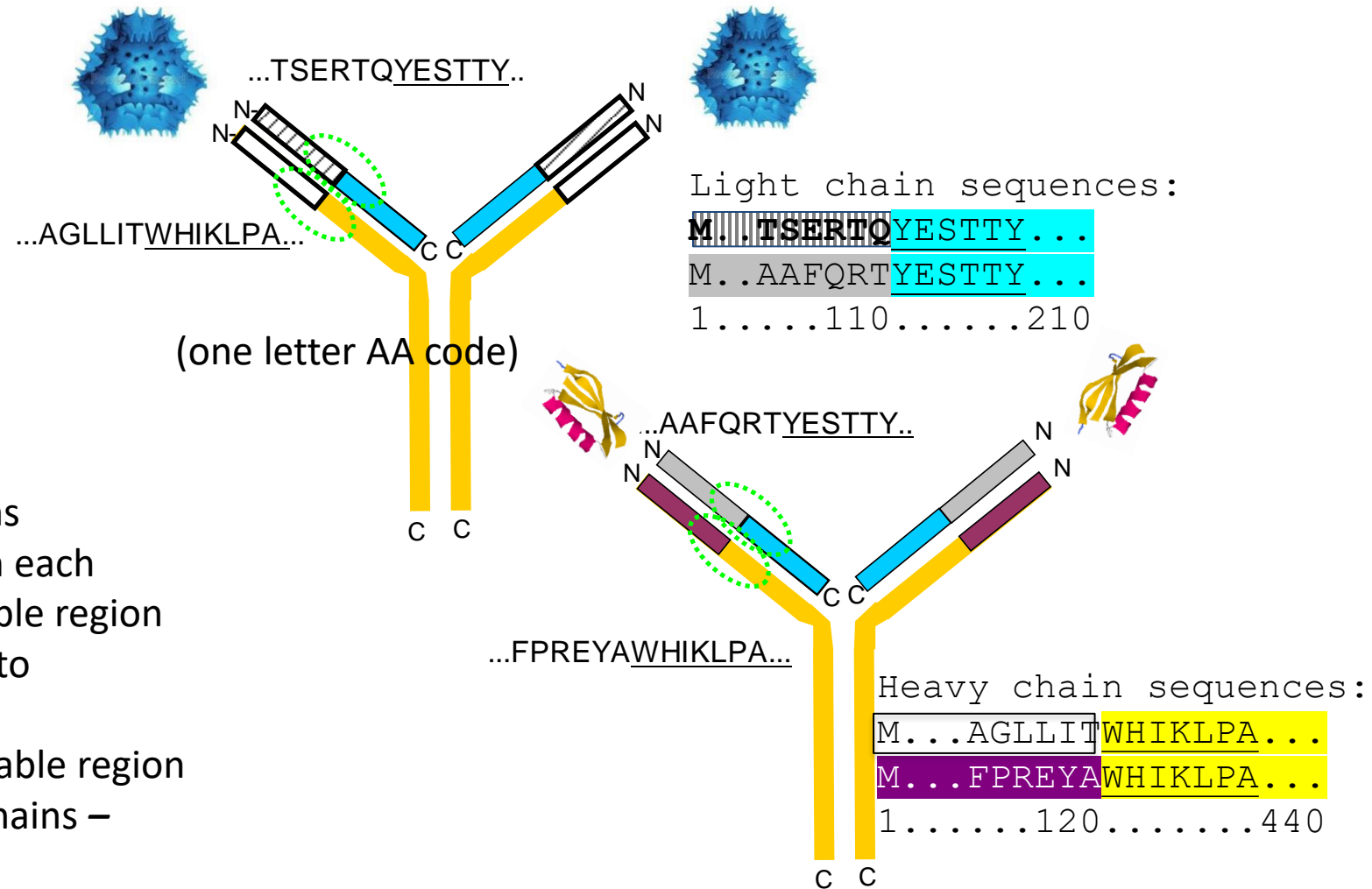
Antibodies can target tumor cells.



Lancet, 373, 966B, P1033

# Production of Antibodies by B-cells

## Primary Structure of Antibodies:



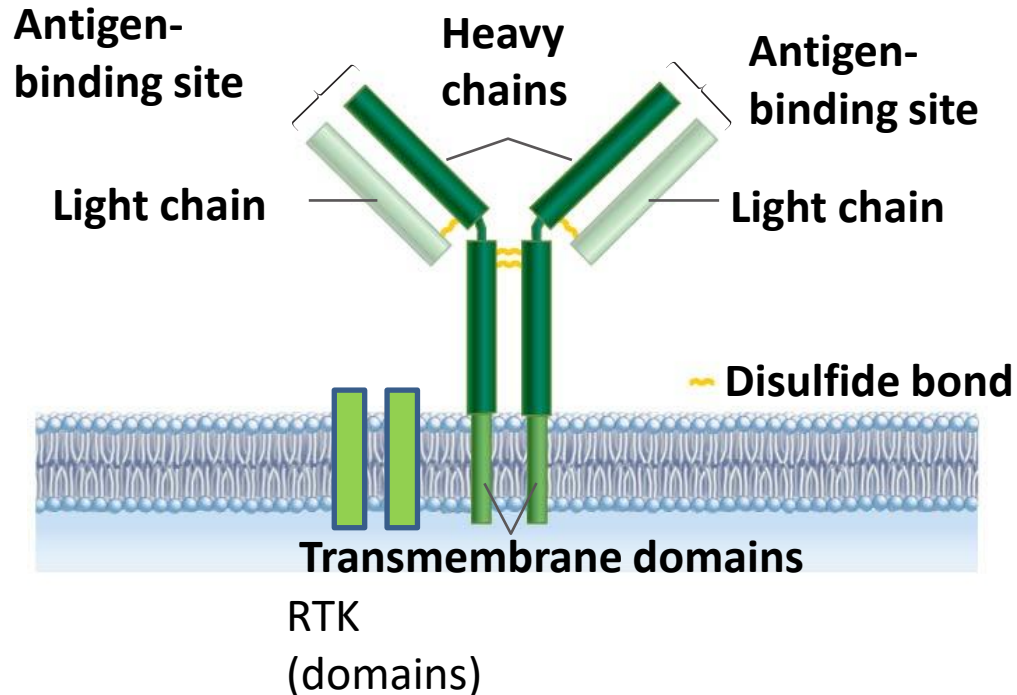
### Each Antibody:

- Two identical light chains
- Two identical heavy chains
- First ~100 Amino acids on each chain are called the variable region and differ from antibody to antibody.
- Unique sequence for variable region of both heavy and light chains – ***defines specificity***
- Constant regions - same protein sequence for all.

# Production of Antibodies by B-cells & Plasma Cells

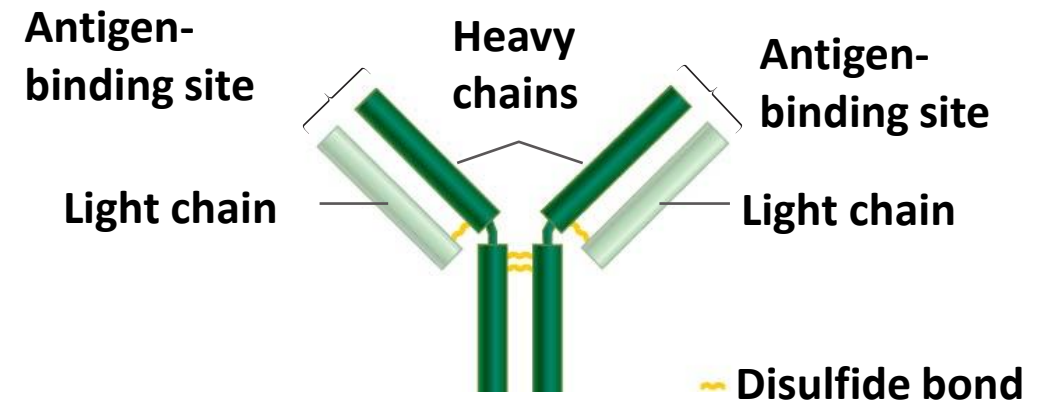
## B- Cells & B-cell Receptor (BCR)

- Each B-cell has only one type of antibody as part of its BCR (B-cell receptor), i.e. the  $10^5$  BCRs are *homogeneous* on the same cell.
- Approximately  $10^8$  different specificities at any one time. i.e.  $10^8$  different B-cells!



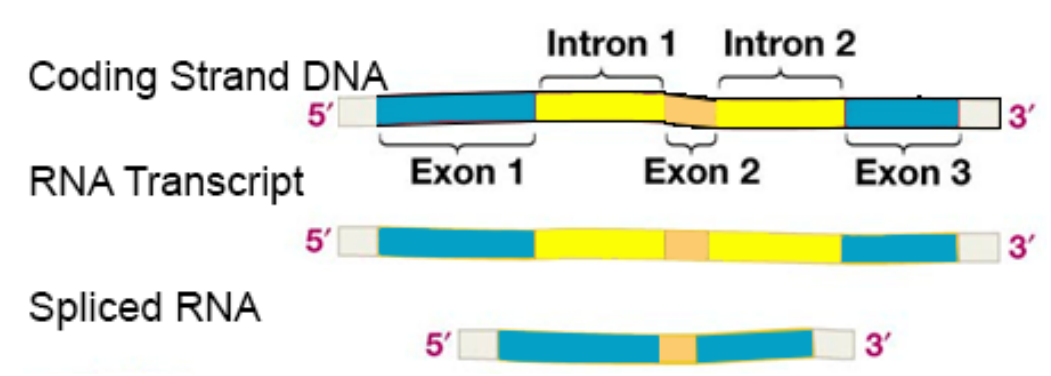
## Plasma Cells:

- After activation, a B-cell develops into a plasma cell.
- The antibody is secreted.
- The same light chains are produced.
- The heavy chains differ only in the absence of the transmembrane domains.

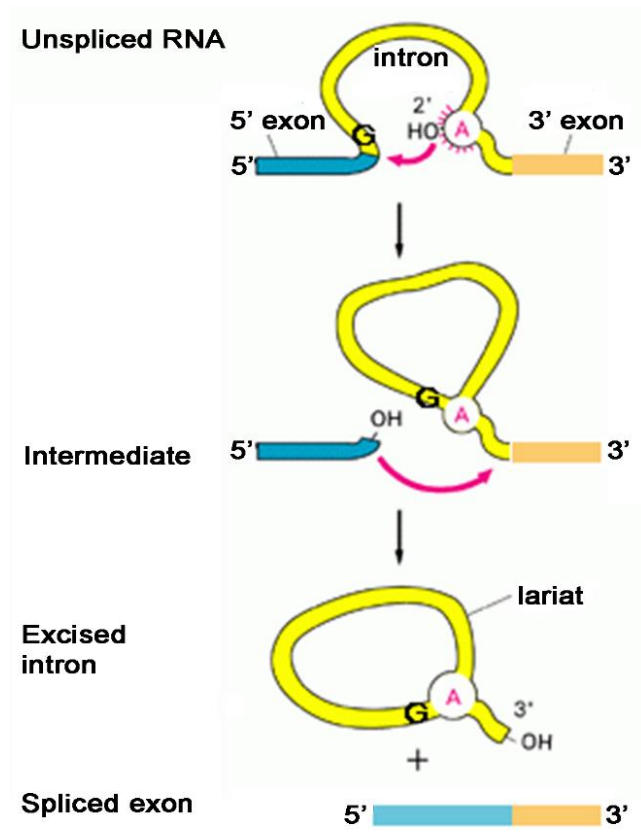
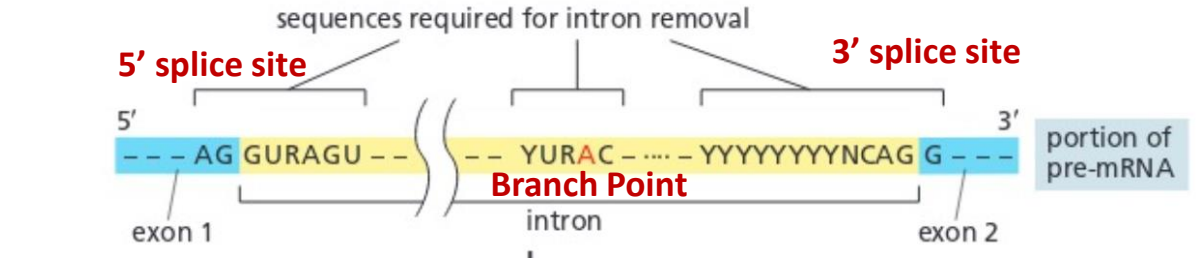


# Antibody Genes are Modular – mRNA Splicing Required to Produce Final mRNA

- When DNA sequences were aligned to RNA sequences, it was found that segments were deleted in the final RNA.
- This suggested that the gene encoding a protein was coded by segments of the DNA:
  - Those to be in the final mRNA were called **exons**.
  - Those sections not in the mRNA were called introns (intervening sequences).



Splice sites are recognized due to specific sequences at the exon-intron boundaries. *Sequences in both the exon and intron are recognized.*



- There is a 5' splice site with a conserved sequence:  
(A/C)AG|GU(A/G)AGU
- There is a 3' splice site with a conserved sequence:  
CAG|G
- There is an A in the intron (branch point) required for splicing.

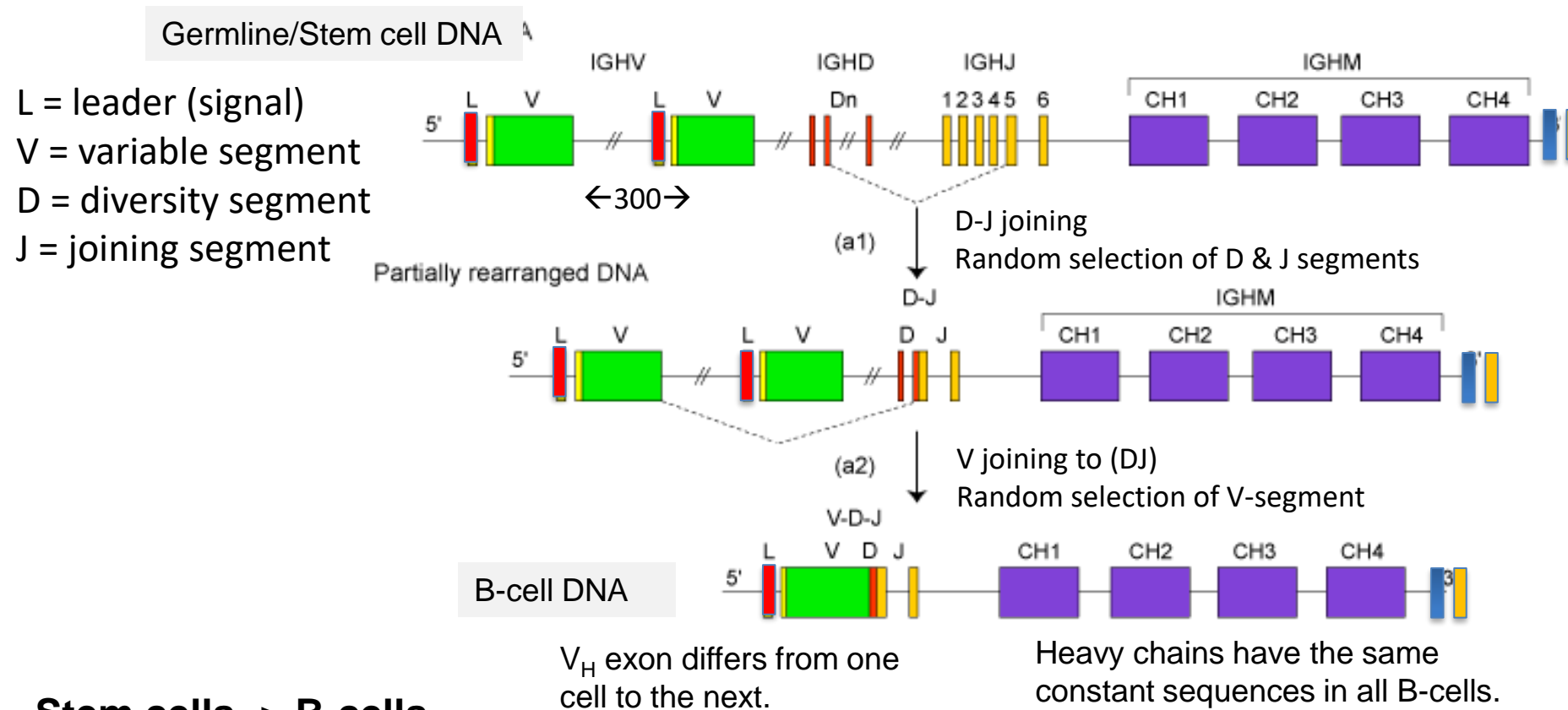
## Steps:

- The branch A breaks at the 5' splice site, forming a lariat.
- The 5'-OH is joined to the 5' end of the downstream 3' exon.

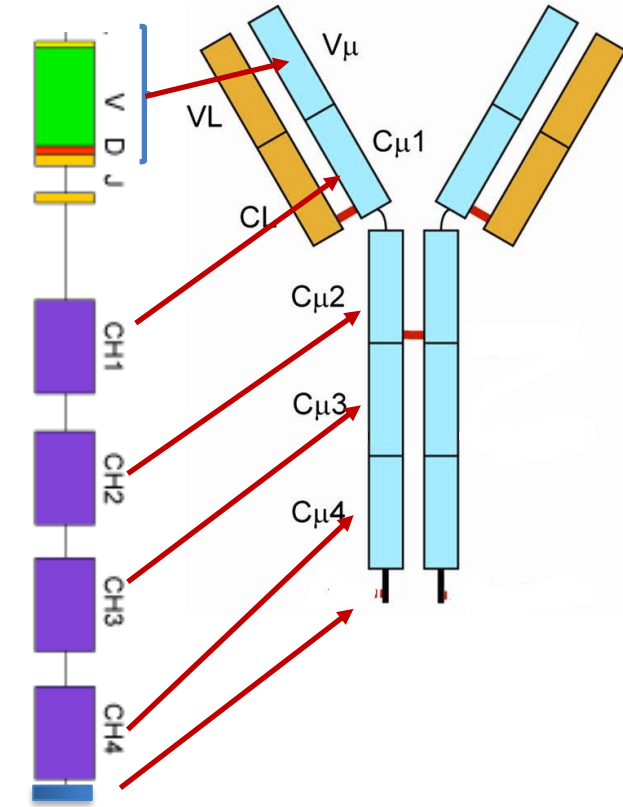
# Antibody genes are assembled from DNA segments, giving many different sequences.

## Production of Heavy Chain Gene:

$V_H$  exon = V+D+J segment (selected at random)



The mRNA coding for antibodies contains 5 exons.



## Stem cells -> B-cells

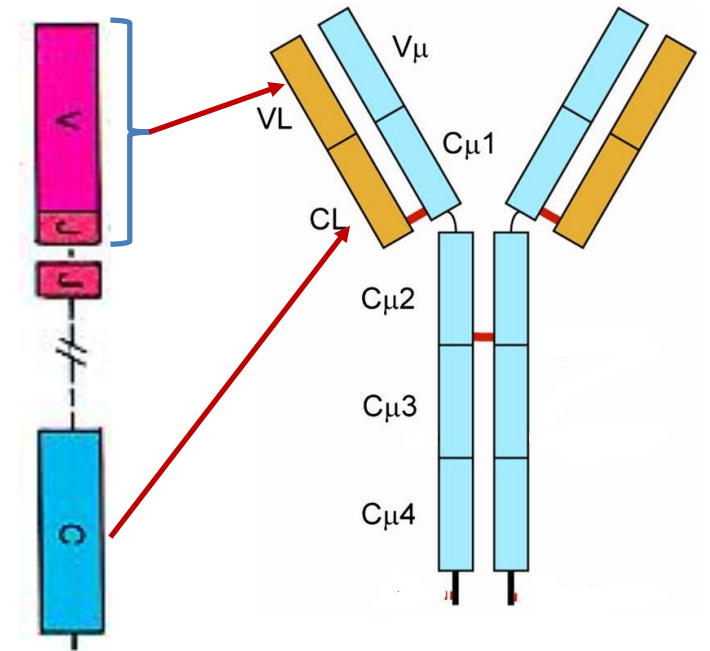
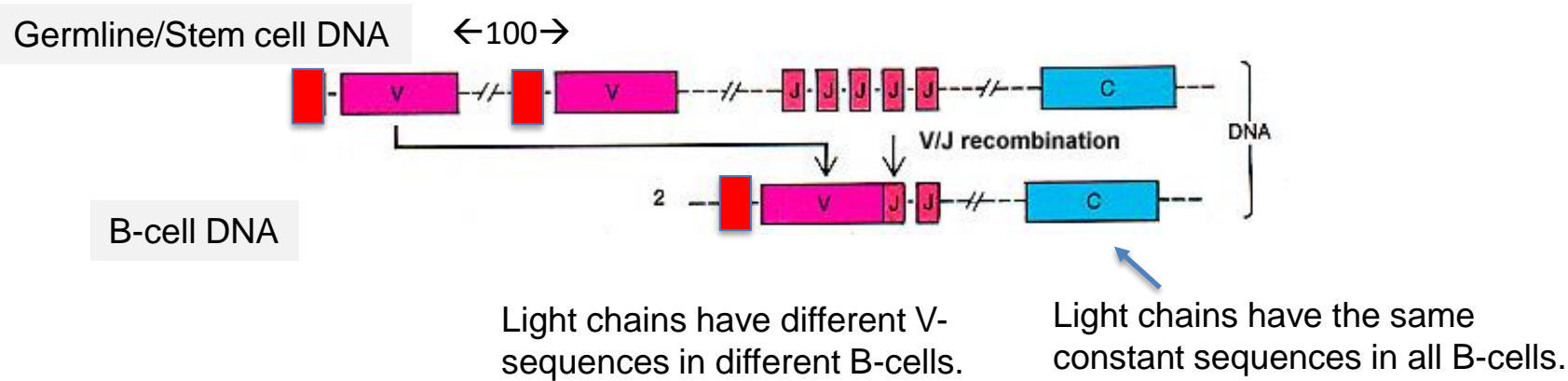
- The exon that codes for the variable region of the heavy chain is generated by the random joining of a V, D, and J DNA segments.
- Each B-cell will generate a unique sequence for its heavy and light chain DNA.
- This is a permanent change to the DNA (**genome**) of the B-cell.

*1. If there are 300 possible V-heavy segments, 10 possible D segments, and 6 possible J segments, how many different heavy chains can be made?*



Light-chain genes are assembled from DNA segments, giving many different sequences.

### Production of Light Chain Gene



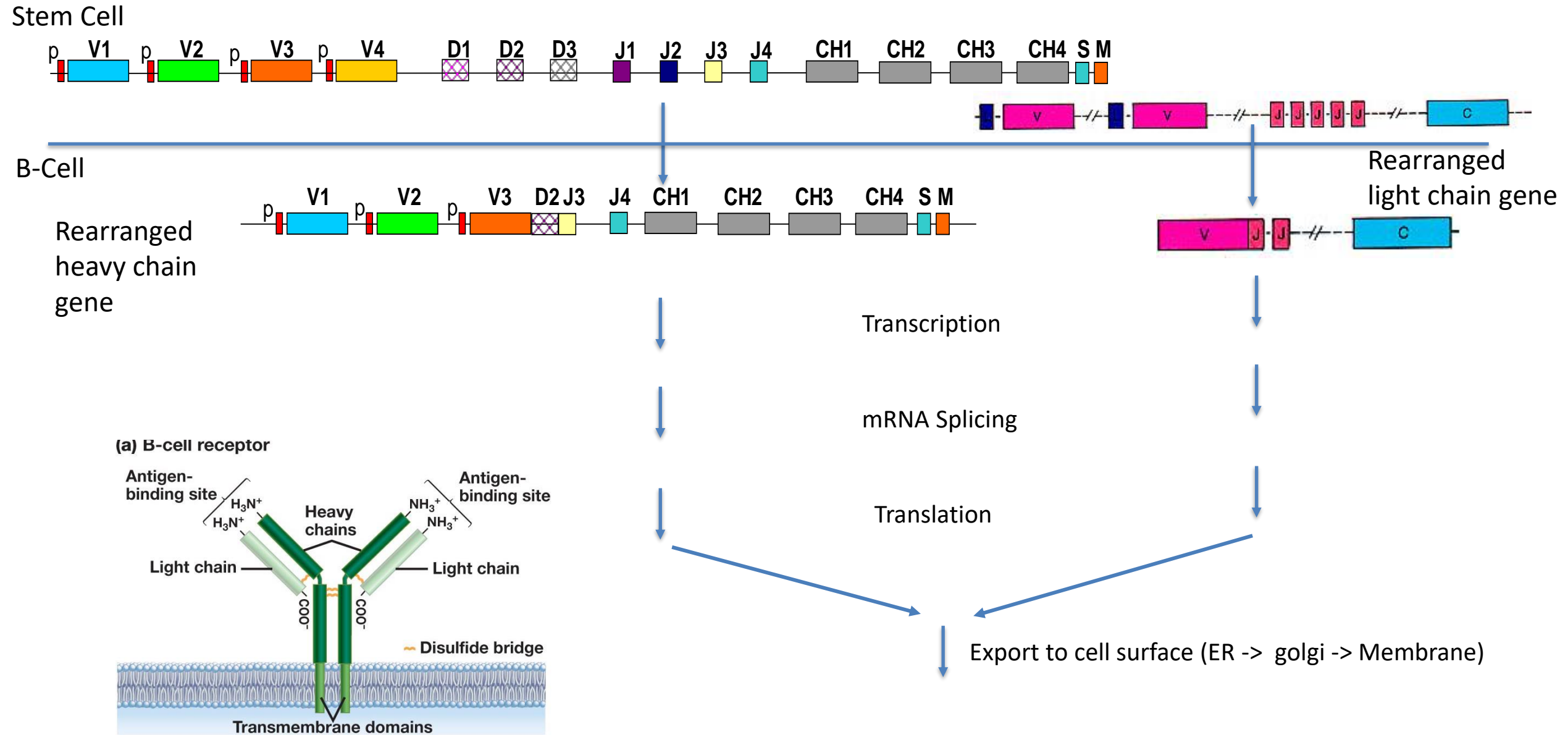
### Antibody Diversity

1. If there are 100 possible V-heavy segments and 5 possible J segments, how many different light chains can be made?
2. If any possible heavy chain can pair with any possible light chain, how many different antibodies can be generated, assuming there are 10,000 possible heavy chains and 500 different light chains?

### Stem cells -> B-cells

- In the case of the light chain, the variable region is generated by VJ joining.
- Each B-cell will generate a unique sequence for its heavy and light chain DNA.
- This is a permanent change to the DNA (**genome**) of the B-cell.

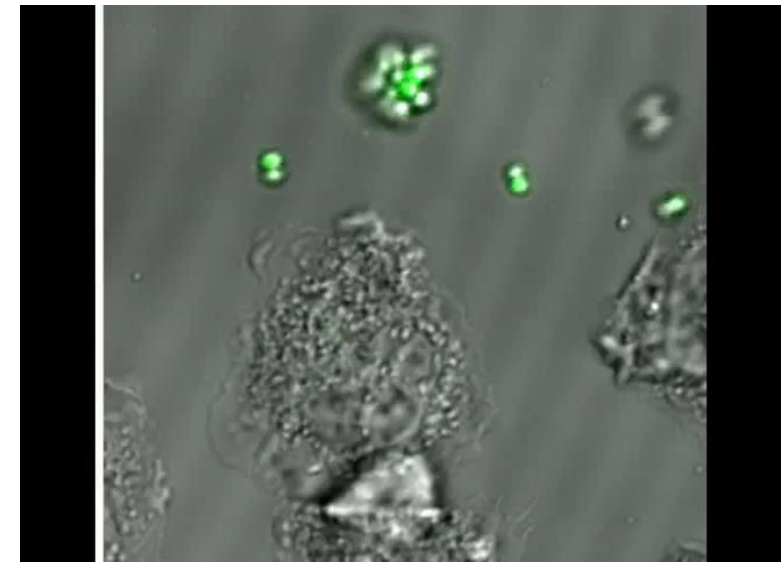
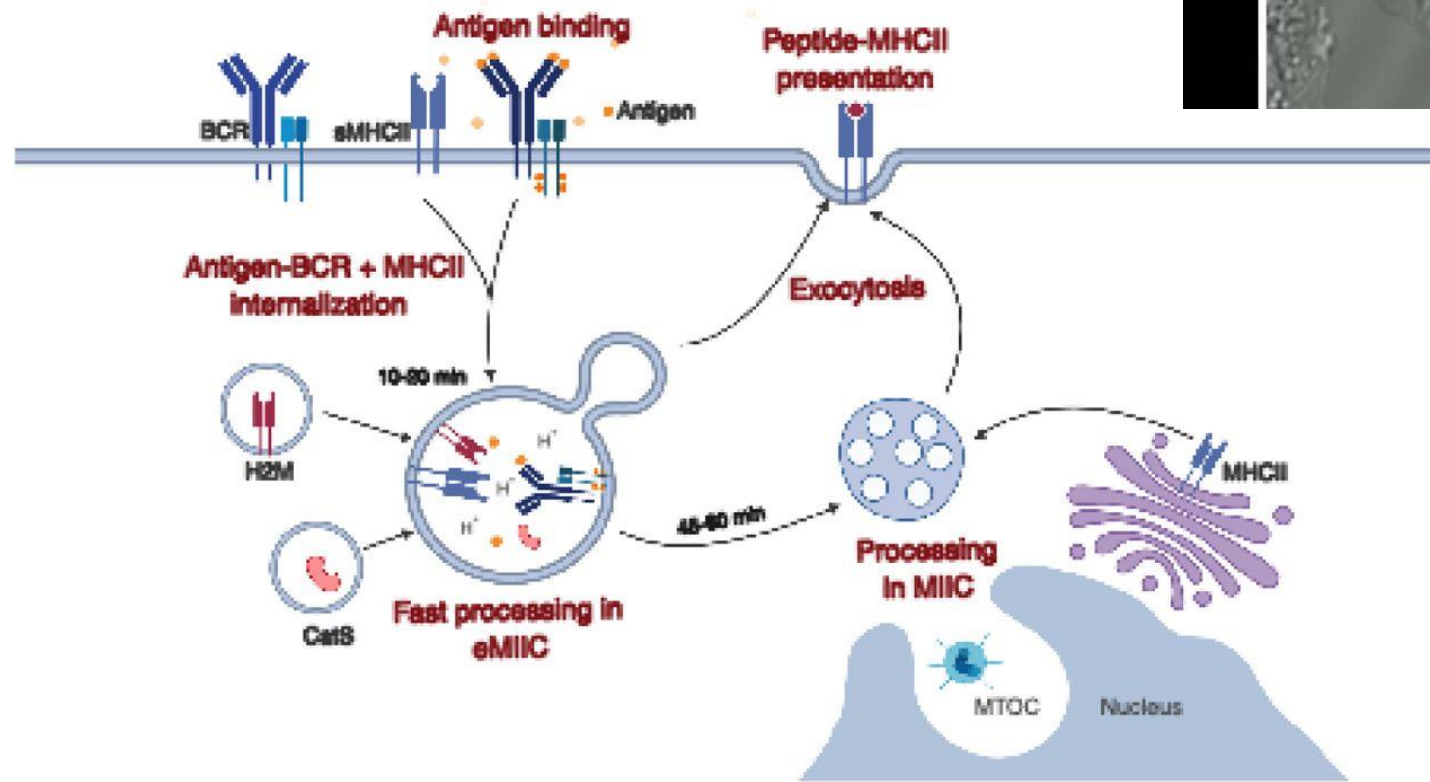
# Antibody Production – From Stem Cells to B-Cells



# Antigen Capture by B-Cells - Endocytic Pathways

1. Antigen binds to variable domains of antibody on the BCR (B-cell receptor)
2. Antigen is internalized and digested into peptides
3. Peptides are loaded on to class II MHC for presentation to T-cells

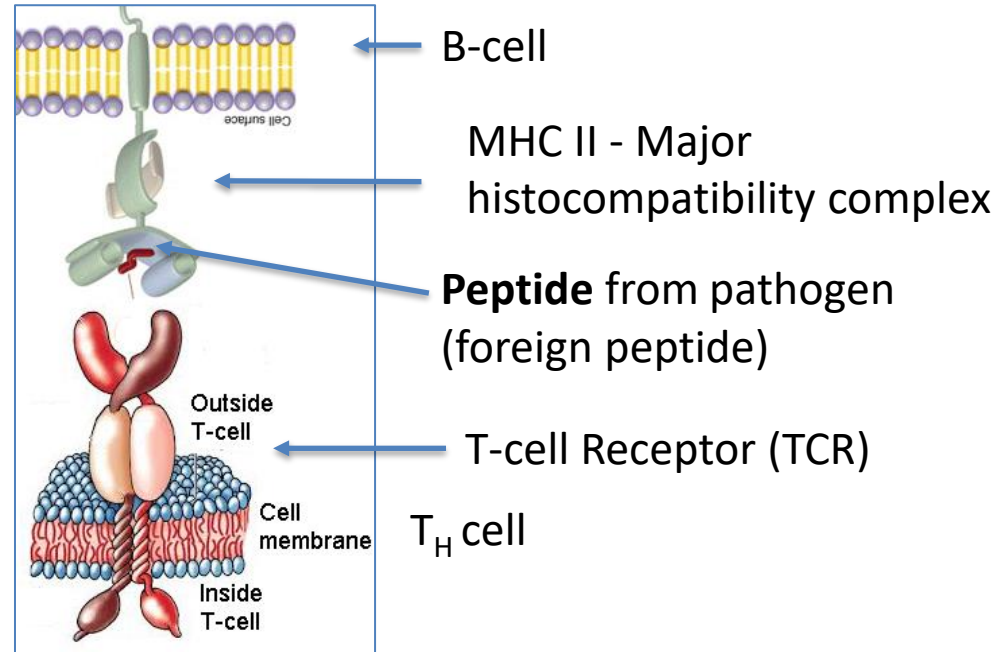
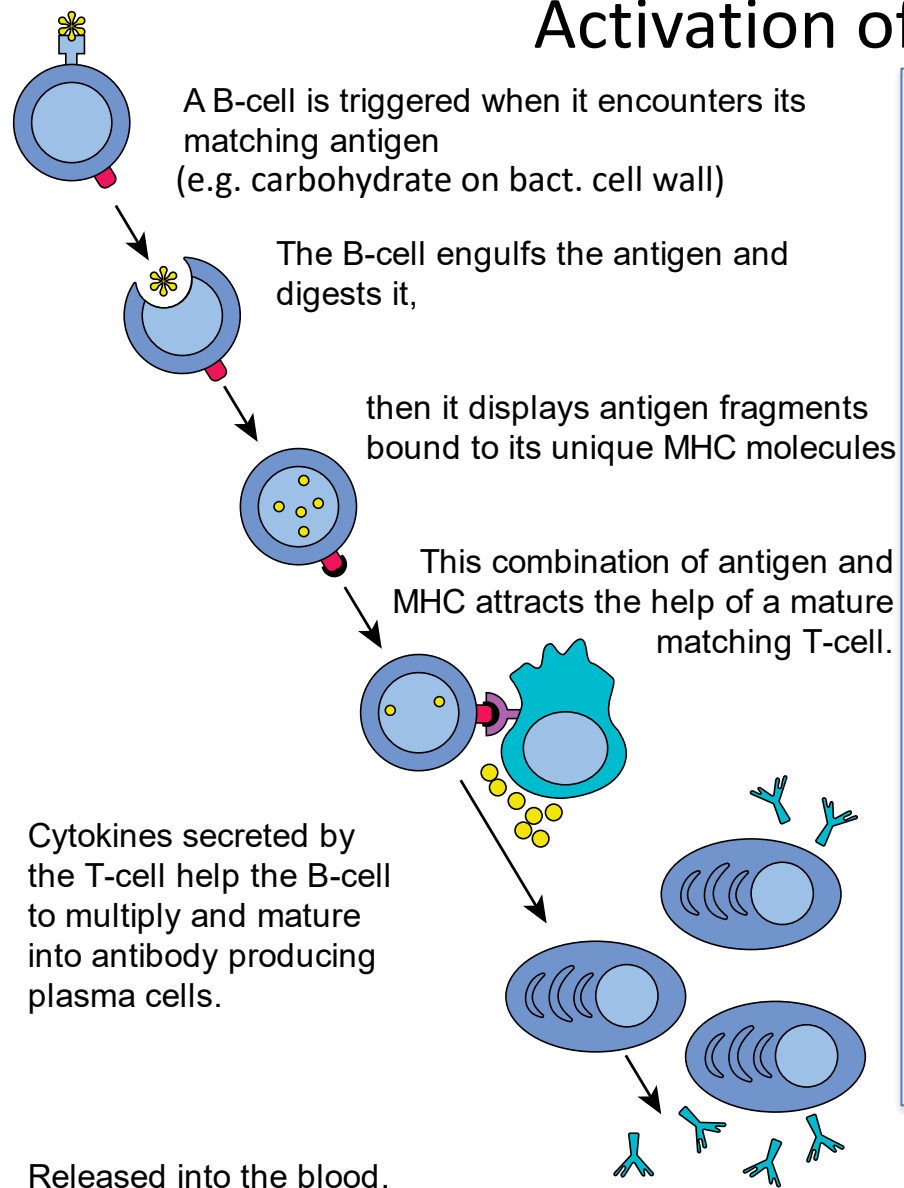
## Endocytosis of bacteria by a B-cell



*Journal of Cell Science* doi: [10.1242/jcs.235192](https://doi.org/10.1242/jcs.235192)



# Activation of B cells by Antigen - Lymph Node



## Events:

1. Recognition of MHC II-peptide by TCR
2. Tyrosine kinase signaling in T<sub>H</sub> cell
3. Cytokines (protein messengers) produced.
4. Cytokines activate B-cells (paracrine)

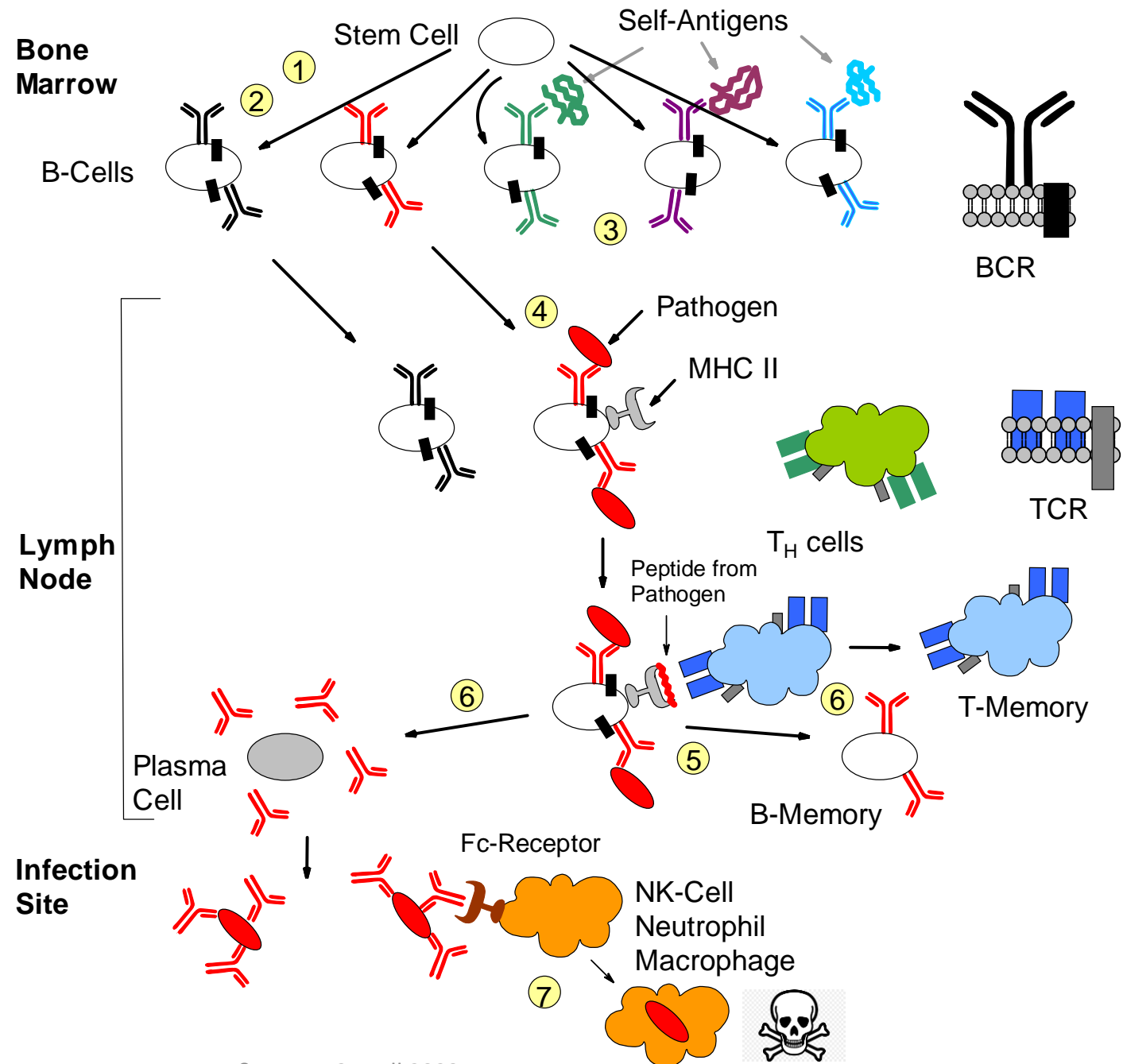
- B-cells develop into antibody secreting *plasma cells*.
- B and T-helper cells develop into **memory** cells, that are long-lived and are quickly activated by the same pathogen. ***This is the basis of vaccination.***

Soluble antibody from plasma cells has the same light and heavy chains as the original B-cell.

Membrane anchors are missing, so antibody is secreted outside the cell.

Can you:

- Describe how the genes for the heavy and light chain are generated, and how this give rise to many different antibodies?
- Do you understand the process of B-cell activation, including presentation of foreign peptides on MHC II and the role of the T-helper cell.
- Describe how antibodies inactivate pathogens?



# Immunology – Part B

## Cell Based Immunology

### Key Questions:

1. How does your immune system fight viruses?
2. How does your immune system detect and destroy cancer cells?
3. How can the immune response be engineered to fight cancer?

### Cell Types:

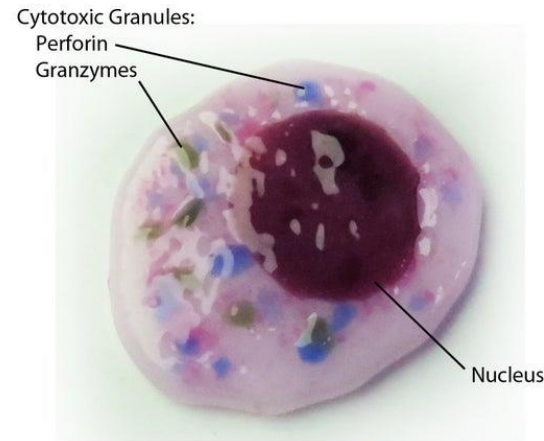
#### Innate

- Natural Killer (NK) cell

#### Acquired

- $T_H$
- $T_C$ ,  $T_{CTL}$

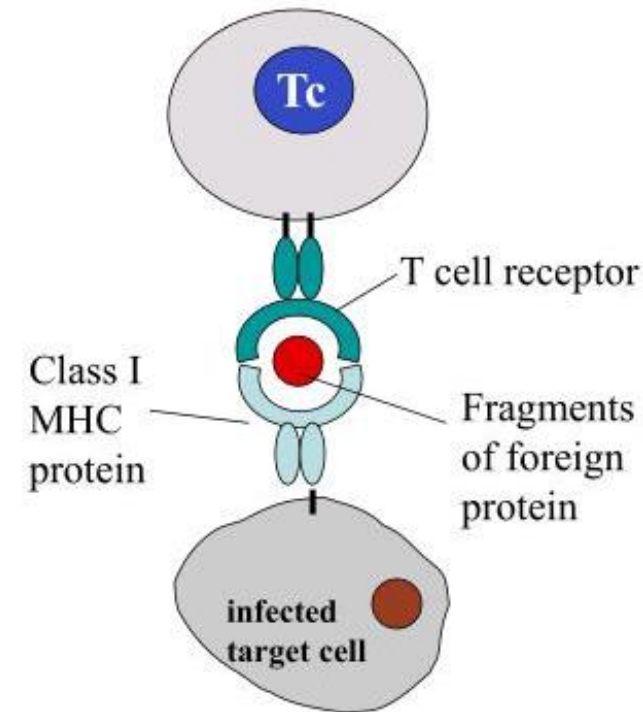
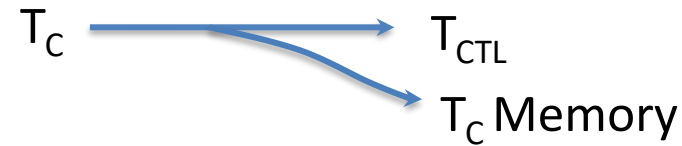
Natural Killer Cell



NK: Innate

- Kill virally infected cells
- Kill cancer cells

5



Activation of Tc cells requires:

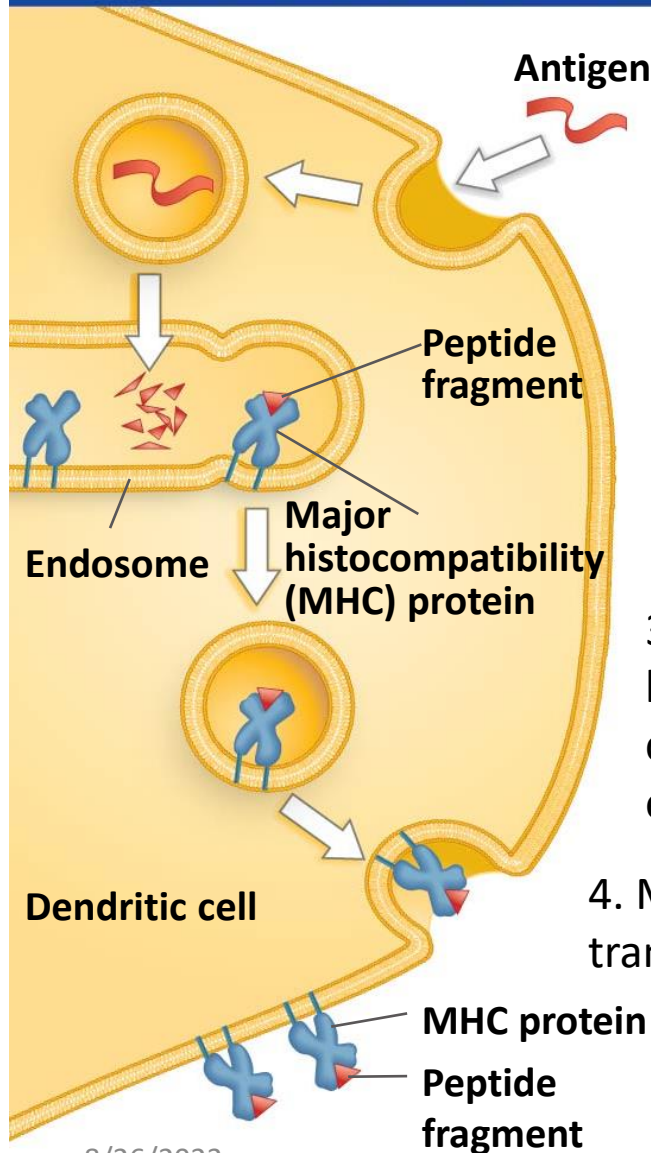
1. Recognition of foreign peptide on MHC I.
2. Assistance from T-helper cells.

Activated Tc cell becomes a cytotoxic T-lymphocyte  $T_{CTL}$

- $T_{CTL}$
- Kill virally infected cells
  - Kill cancer cells

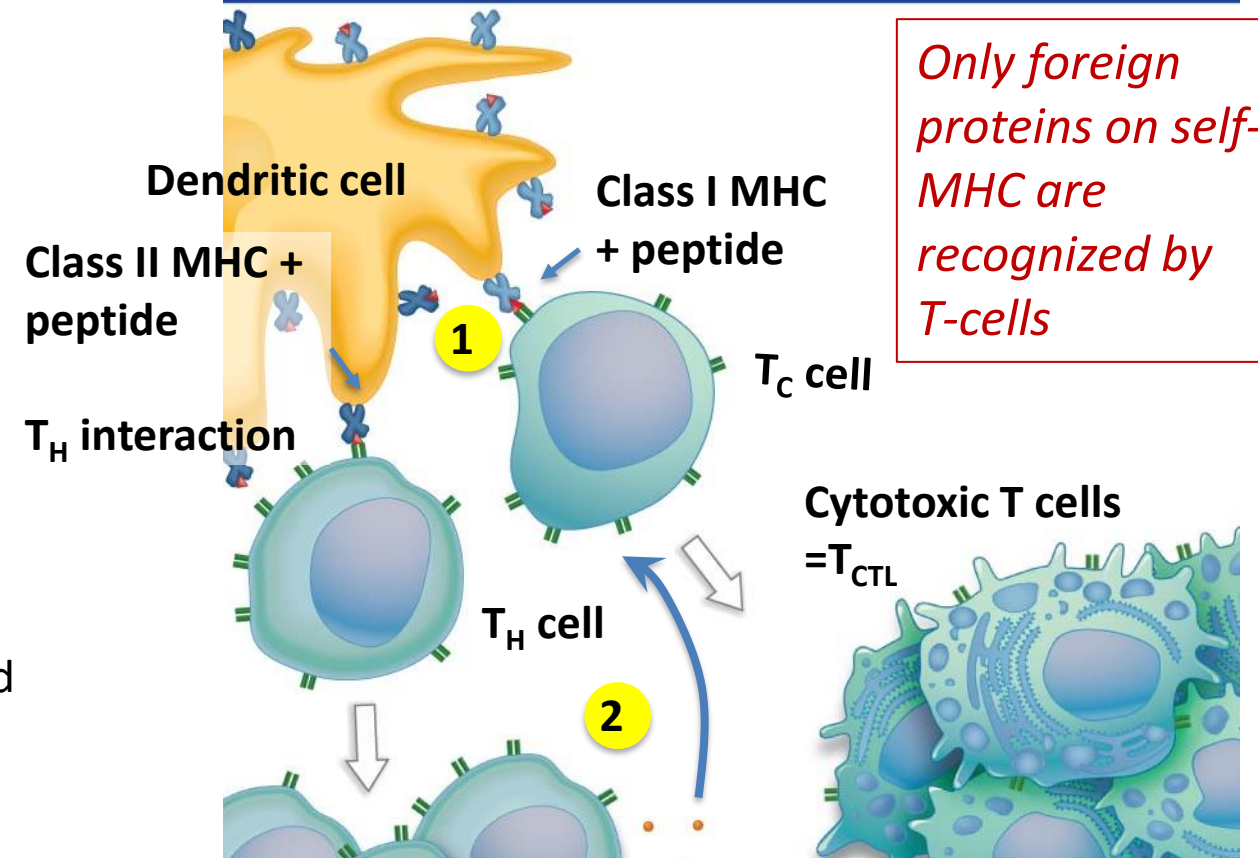
# Dendritic cells acquire antigen from viruses and cancerous cells, activating T-cells

## PROCESS: MHC ANTIGEN PRESENTATION



1. Dendritic cell ingests antigen via **phagocytosis** (intact virus, cell debris from cancer cell).
2. Enzymes break antigen proteins into peptide fragments.
3. Peptide fragments are loaded onto both class I and class II MHC proteins in endosomes.
4. MHC I & II –peptide complex is transported to cell surface.
5. MHC protein presents peptide fragment on cell surface.

## PROCESS: T-CELL ACTIVATION



Activation of T<sub>C</sub> cells requires:

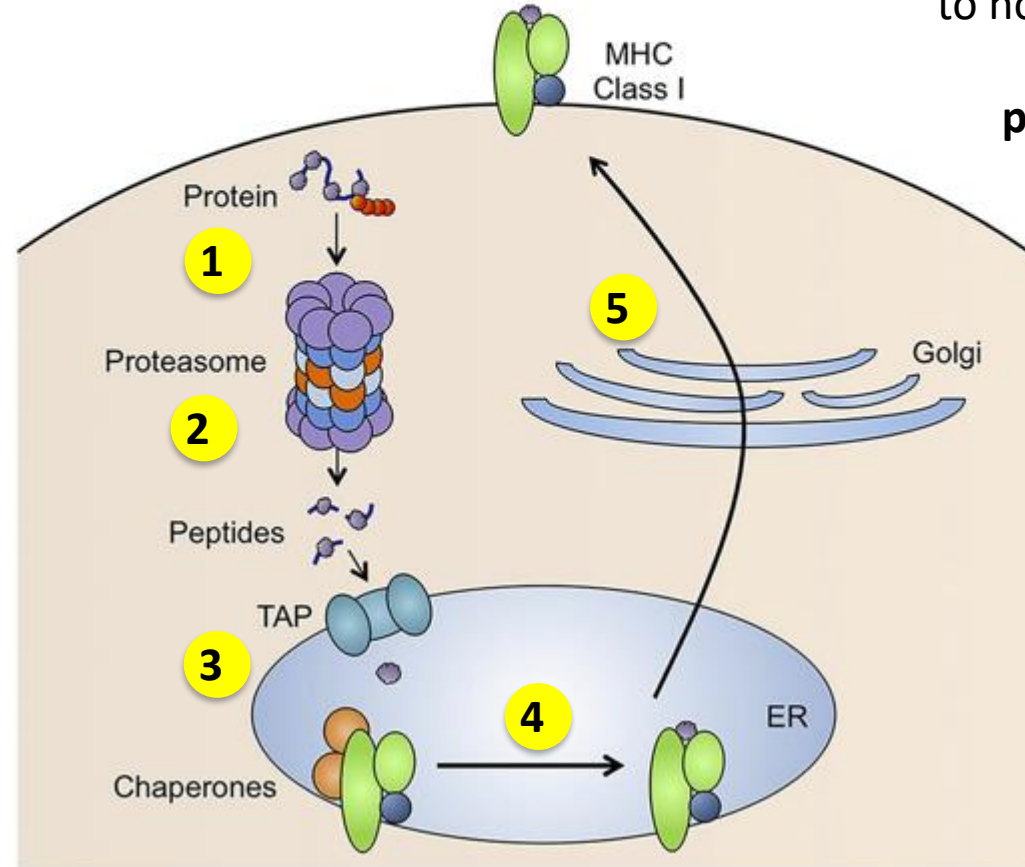
1. Recognition of foreign peptide on MHC I.
2. Assistance from T-helper cells via secreted messengers (small proteins called cytokines)



# Acquired Cellular Immunity - Role of MHC I in Presentation of Peptides

- MHC I present peptides
- Peptides are generated from of **all** of the proteins that are made in the cell:

1. protein targeted by ubiquitin
2. Protein digested by proteasome
3. Peptides transported into ER
4. Peptides loaded on to MHC I
5. Peptide/MHC complex transported to cell membrane.



## Foreign Peptide Source:

1. From replication of viruses in the cell
2. From replication of intracellular bacteria (e.g. TB)
3. New coding sequences in cancer cells due to genetic changes (e.g. mutations in p53 may lead to novel sequences).

## p53 Protein Sequence

Zn Fingers (DNA binding)				
10	20	30	40	50
MEEPQSDPSV	EPPLSQETFS	DLWKLLPENN	VLSPLPSQAM	DDLMLSPDDI
60	70	80	90	100
EQWFTEDPGP	DEAPRMPEAA	PPVAPAPAAP	TPAAPAPAPS	WPLSSSVPSQ
110	120	130	140	150
KTYQGSYGFR	LGFLHSGTAK	SVTCTYSPAL	NKMFCQLAKT	CPVQLWVDST
160	170	180	190	200
PPPGTRVRAM	AIYKQSQHMT	EVVRRCPHHE	RCSDSDGLAP	PQHLIRVEGN
210	220	230	240	250
LRVEYLDDRN	TFRHSVVVFY	EPPEVGSDCT	TIHYNMCMNS	SCMGMNRRP
260	270	280	290	300
ILTIITLEDSE	SGNLLGRNSF	EVRVCACPGR	DRRTEENLR	KKGEPHHELP
310	320	330	340	350
PGSTKRALPN	NTSSSPQPKK	KPLDGEYFTL	QIRGRERFEM	FRELNEALEL
360	370	380	390	
KDAQAGKEPG	GSRAHSSHLK	SKKGQSTSRH	KKLMFKTEGP	DSD

EVVRRCPHHE

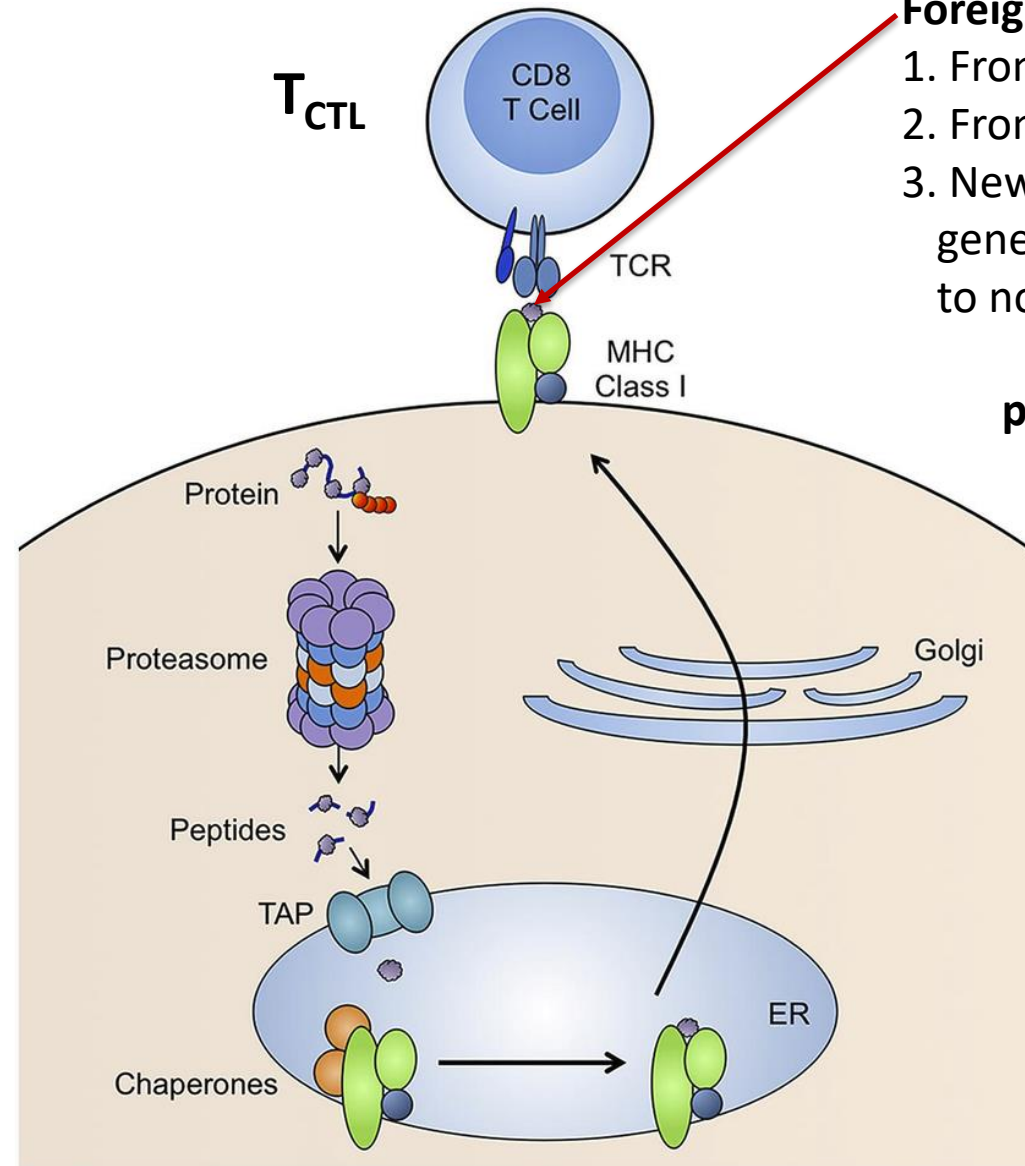
Normal seq., **ignored** by TCR

EVVGGCPHHE

Mutant seq. in cancer, **detected** by TCR

# Acquired Cellular Immunity – Recognition of MHC I + Peptide by Tc Cells

- Tc cells will be activated **only** by **foreign peptides** on MHC class I.
- Tc cells develop into:
  - Tc memory cells
  - cytotoxic T-lymphocytes ( $T_{CTL}$ )
- $T_{CTL}$  cells kill the infected/cancer cell presenting the same peptide that activated it.



## Foreign Peptide Source:

1. From replication of viruses in the cell
2. From replication of intracellular bacteria (e.g. TB)
3. New coding sequences in cancer cells due to genetic changes (e.g. mutations in p53 may lead to novel sequences).

## p53 Protein Sequence

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10	20	30	40	50
MEEPQSDPSV	EPPLSQETFS	DLWKLLPENN	VLSPLP SQAM	DDLMLSPDDI
60	70	80	90	100
EQWFTEDPGP	DEAPRMPEAA	PPVAPAPAAP	TPAAPAPAPS	WPLSSSVPSQ
110	120	130	140	150
KTYQGSYGFR	LGFLHSGTAK	SVTCTYSPAL	NKMFCQLAKT	CPVQLWVDST
160	170	180	190	200
PPPGTRVRAM	AIYKQSQHMT	EVVRRCPHHE	RCSDSDGLAP	PQHLIRVEGN
210	220	230	240	250
LRVEYLDDRN	TFRHSVVVFY	EPPEVGSDOT	TIHYNM CNS	SCMGGMNRRP
260	270	280	290	300
ILTIITLED S	SGNLLGRNSF	EVRVCACPGR	DRRTEENLR	KKGEPHHELP
310	320	330	340	350
PGSTKRALPN	NTSSSPQPKK	KPLDGEYFTL	QIRGRERFEM	FRELNEALEL
360	370	380	390	
KDAQAGKEPG	GSRAHSSHLK	SKKGQSTSRH	KKLMFKTEGP	DSD

EVVRRCPHHE

Normal seq., **ignored** by TCR

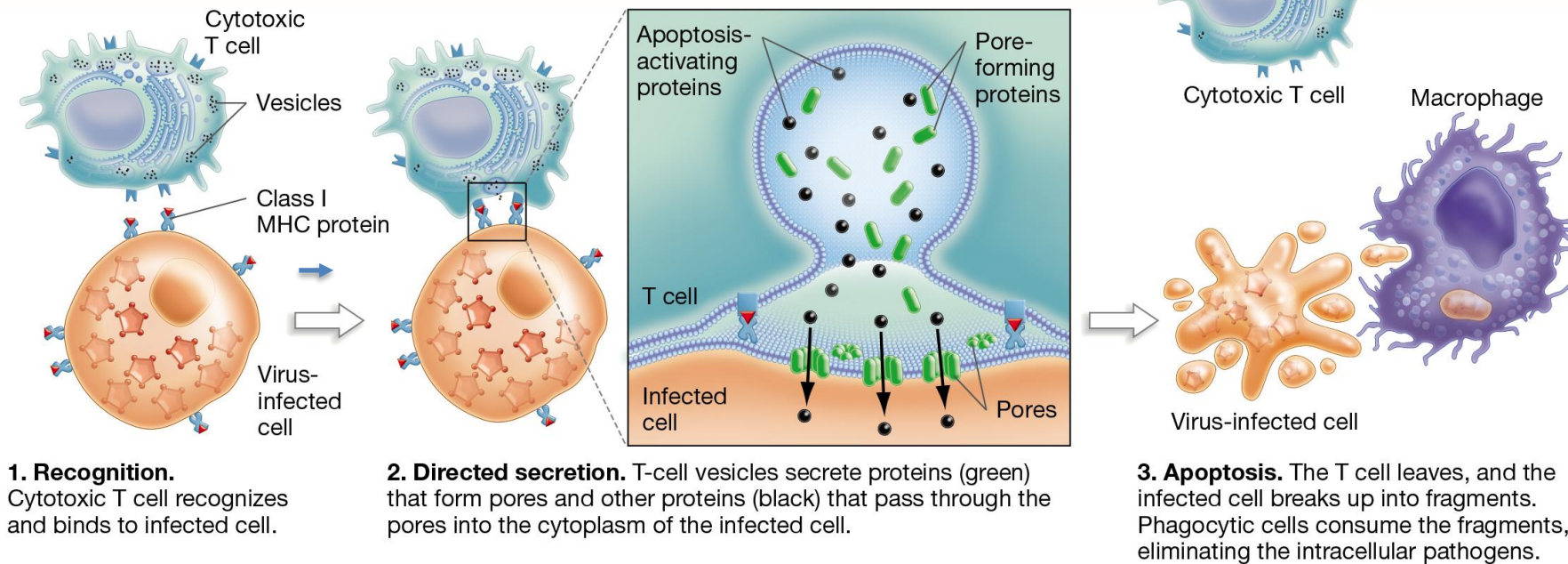
EVVGGCPHHE

Mutant seq. in cancer, **detected** by TCR



# Detection and Killing of Virally Infected or Cancer Cells:

## PROCESS: CELL-MEDIATED RESPONSE



**Cancer cell or  
Infected cell**

- Granzymes enter through perforin pore and cause cell undergo programmed cell death (apoptosis)

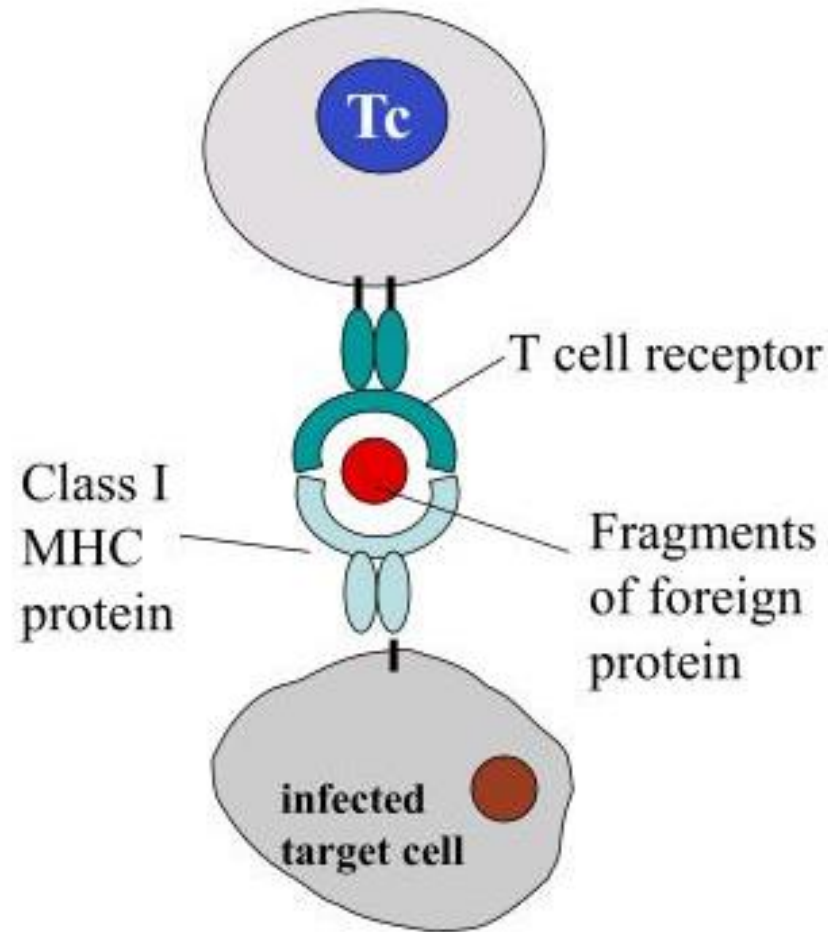
**Cytotoxic  
T-Lymphocyte  
Killing Target**

© James A. Sullivan  
Quill Graphics  
Charlottesville, VA USA

# Cancer Evasion Mechanism I - Loss of MHC I on Tumor Cell

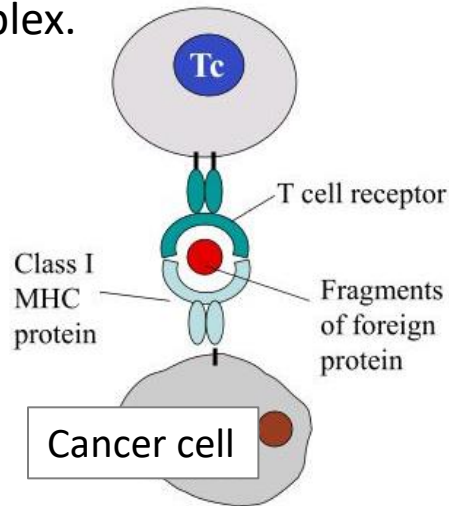
Loss of MHC I expression means that  $T_{CTL}$  cells can no longer recognize and kill cancer cells because T-cell activation requires recognition of the MHC-peptide complex.

*How to re-establish  $T_C$  contact with tumor cell and activation of the T-cell so that the cancer cell is killed?*

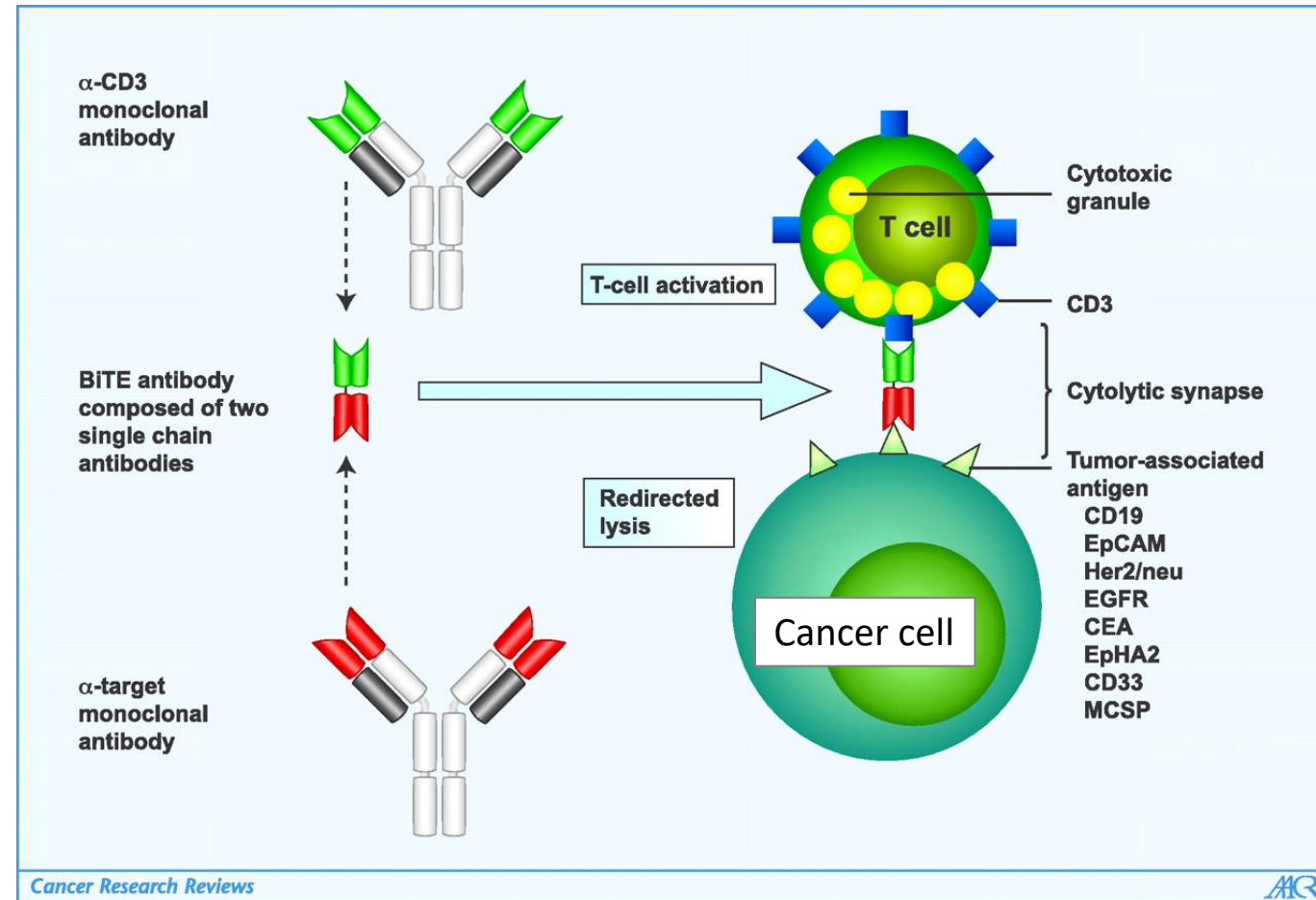


# Cancer Treatment with Antibodies - 1. Cancer Evasion - Loss of MHC I on Tumor Cell

Loss of MHC I expression means that  $T_{CTL}$  cells can no longer recognize and kill cancer cells because T-cell activation requires recognition of the MHC-peptide complex.



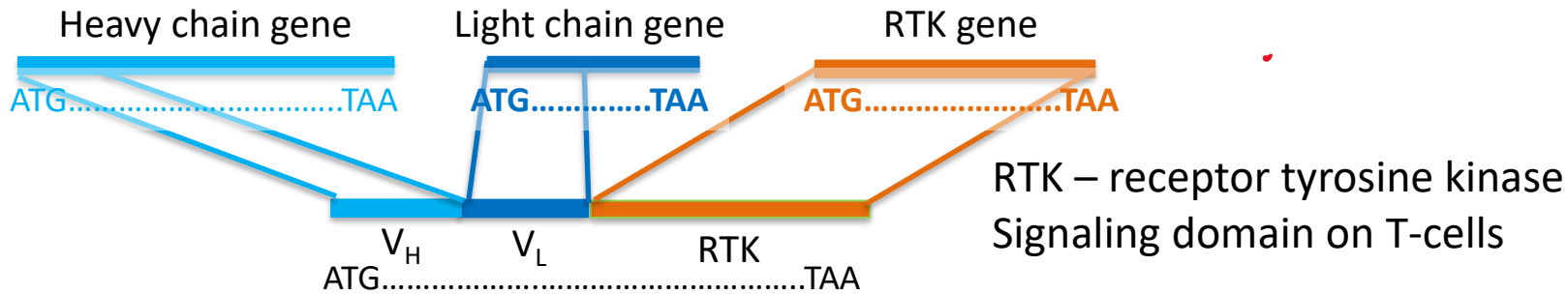
*How to re-establish  $T_C$  contact with tumor cell and activation of the T-cell so that the cancer cell is killed?*



- Bispecific antibodies are generated from two separate antibodies:
  - One recognizes CD3, which is part of the T-cell receptor (TCR)
  - Other recognizes a tumor antigen.
- The two variable regions from each antibody are linked into a single polypeptide chain.
- The dual binding event mimics the original MHC-I TCR interaction.,

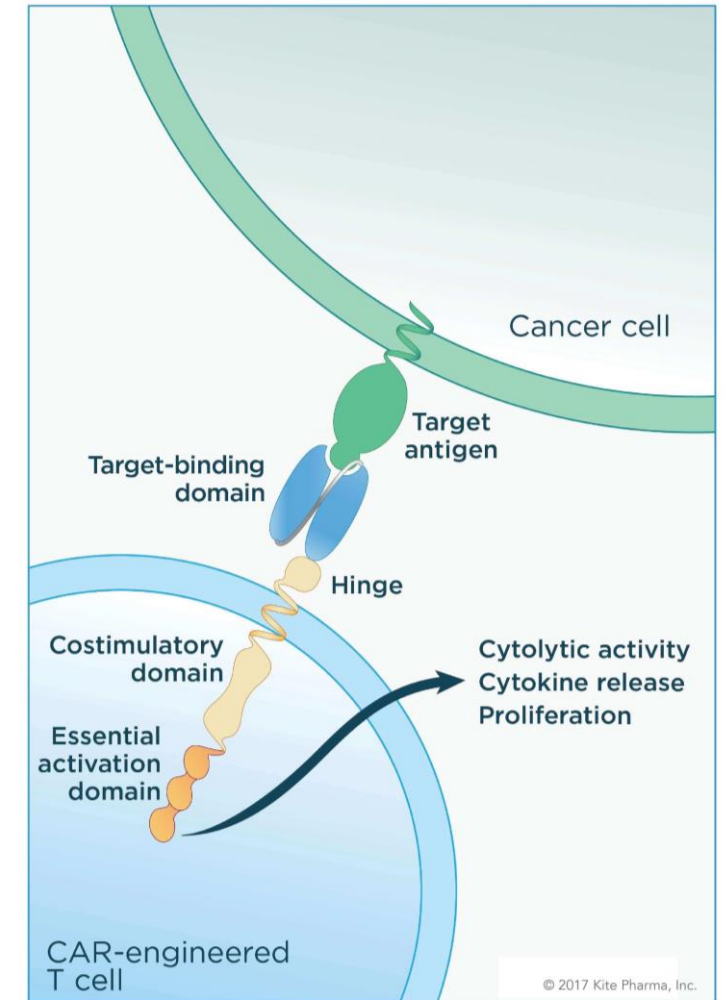
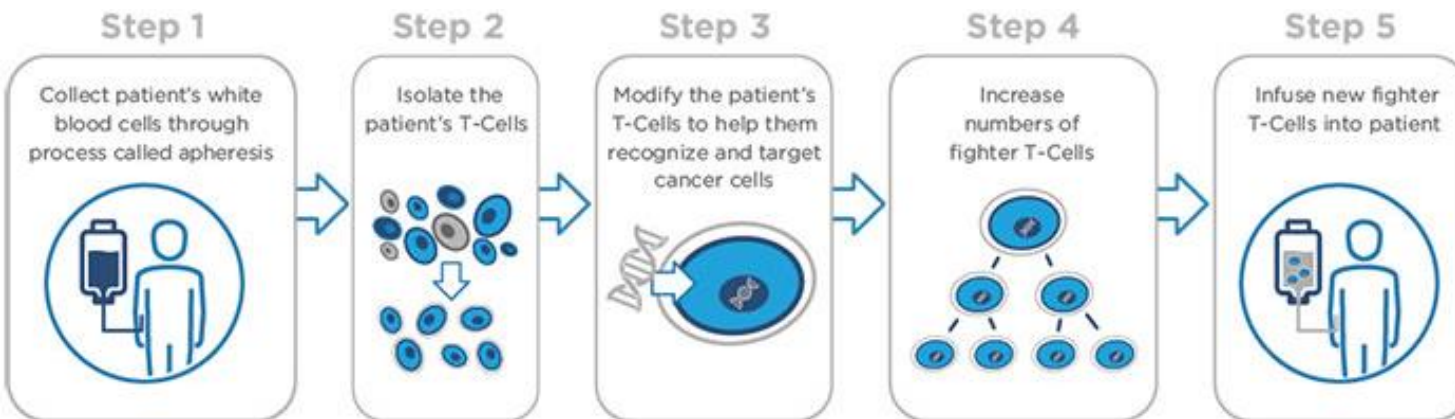
# Chimeric Antigen Receptor T-cells = CAR T-Cells

- A. Obtain antibodies against cancer antigen, isolate genes that code for light and heavy chains for those antibodies.
- B. Fuse coding region for variable light and heavy domains to coding region for RTK on T-cells = single CAR-T gene.



- C. Introduce gene for CAR-T cell into Patient
  - 1. Obtain white blood cells from patient
  - 2. Isolate T-cells
  - 3. Introduce DNA into T-cells
  - 4. Obtain large amounts of T-cells by cell culture
  - 5. Inject CAR-T cells into cancer patient.

D. What happens when Antigen is encountered?





# Cancer Evasion Mechanism II – Downregulation/killing of Tc cells.



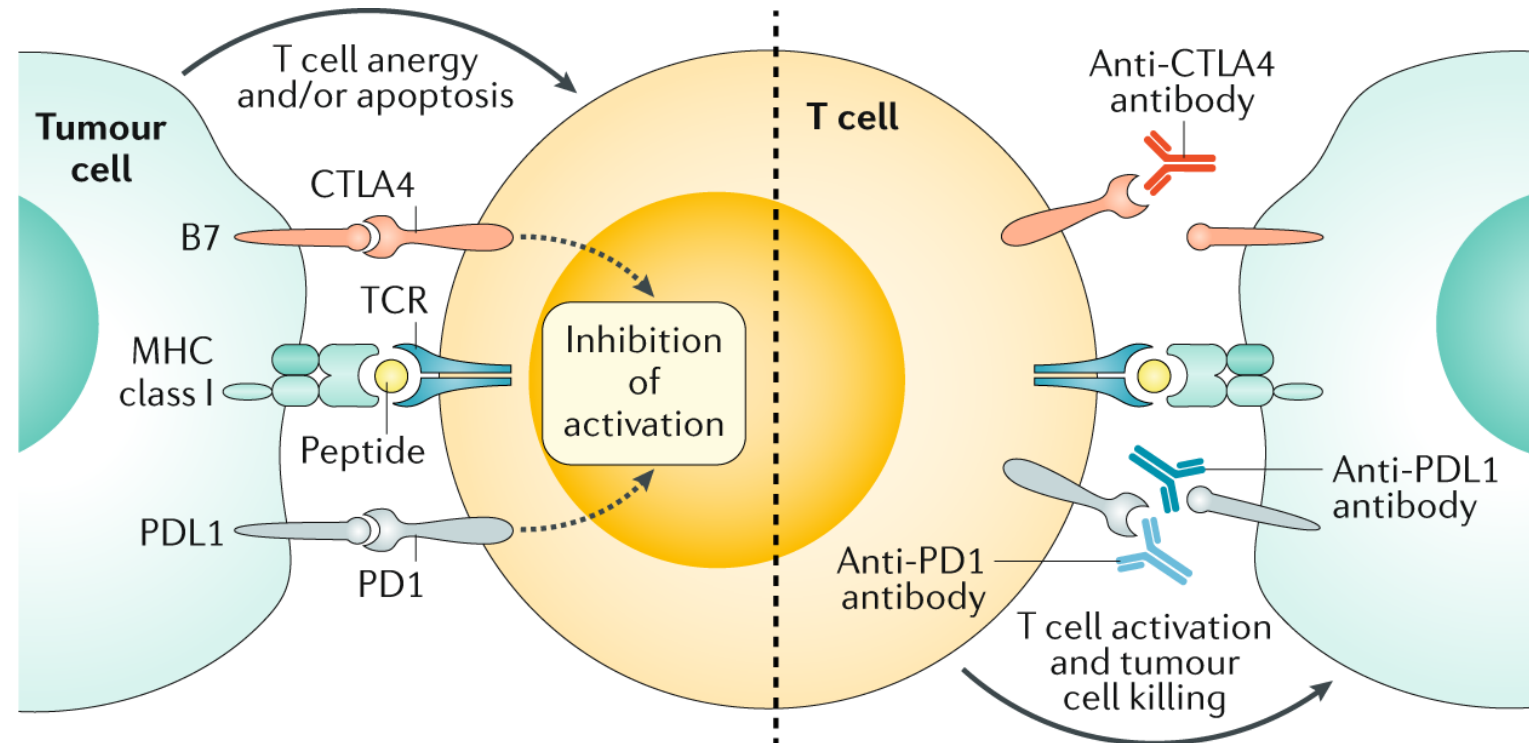
James P. Allison  
Prize share: 1/2

Tasuku Honjo  
Prize share: 1/2

The Nobel Prize in Physiology or Medicine 2018 was awarded jointly to James P. Allison and Tasuku Honjo "for their discovery of cancer therapy by inhibition of negative immune regulation."

- Cancer cells overproduce b7.**  
Binding of b7 to CTLA4 receptor on the surface of the Tc cell deactivates the Tc cell - immunosuppressive reaction called anergy.
- Cancer cells overproduce PDL1.**  
PDL1 binds to PD1 on T-cells. Activation of signaling causes Tc cell to enter apoptosis (programmed cell death).

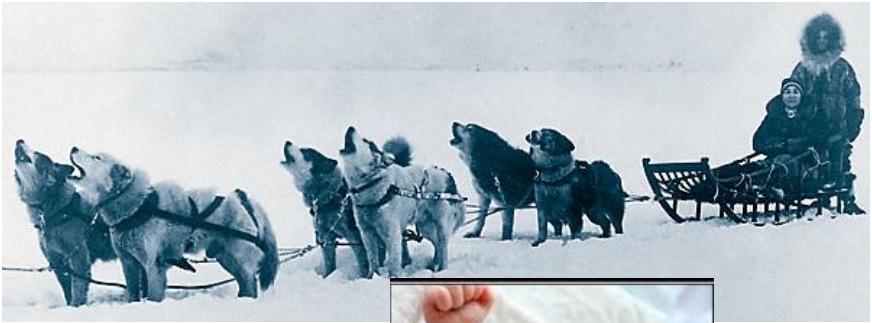
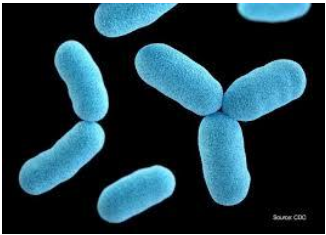
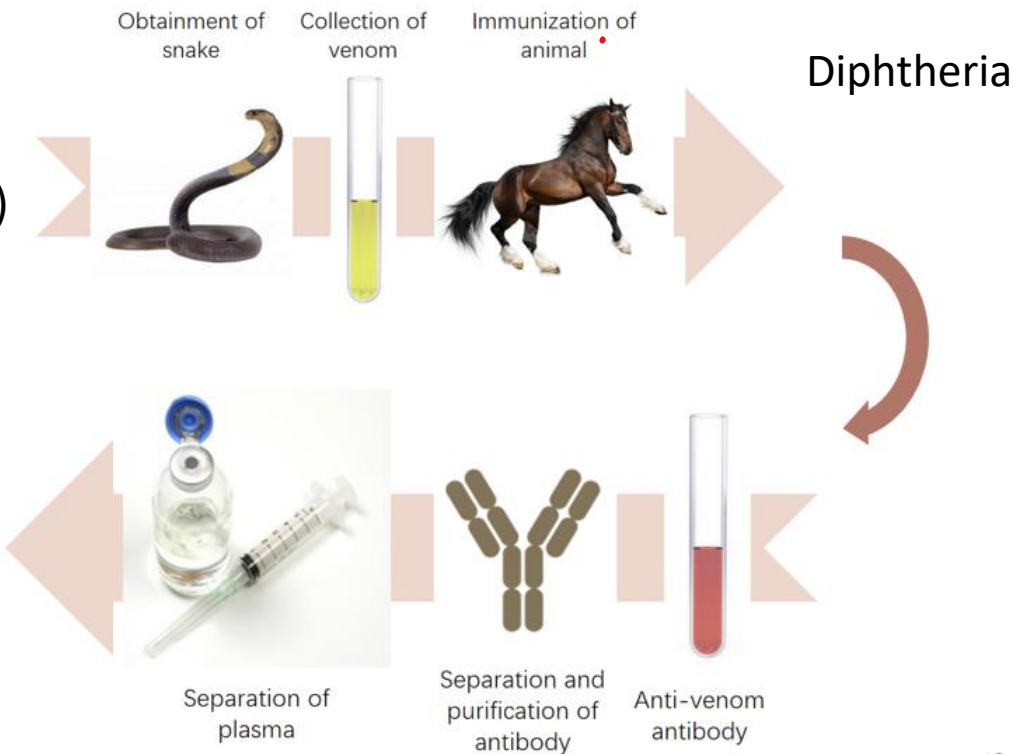
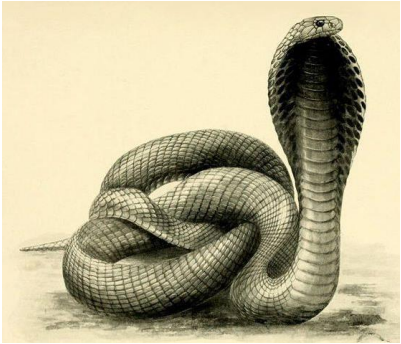
*How to block this signaling?*



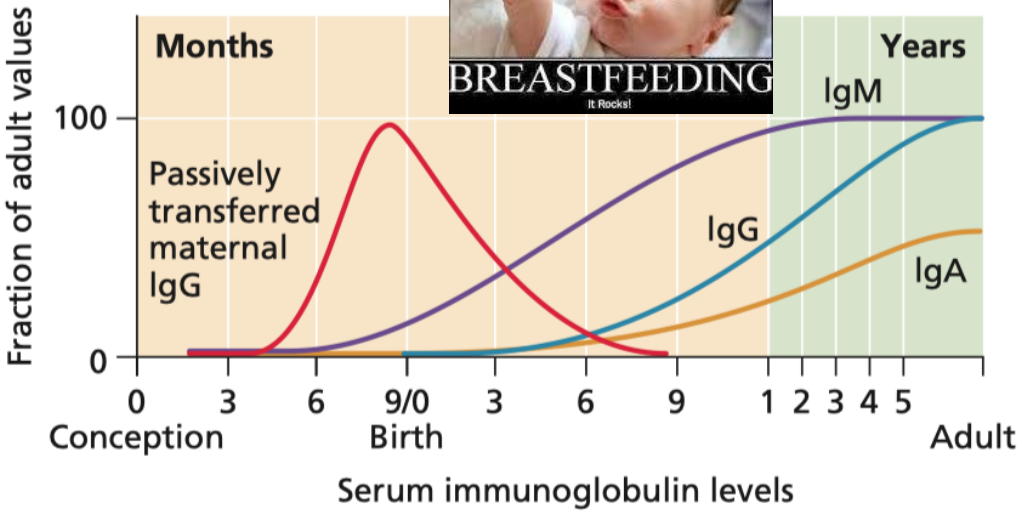
# Vaccination

## Types of vaccines:

1. Passive (Ab injected)

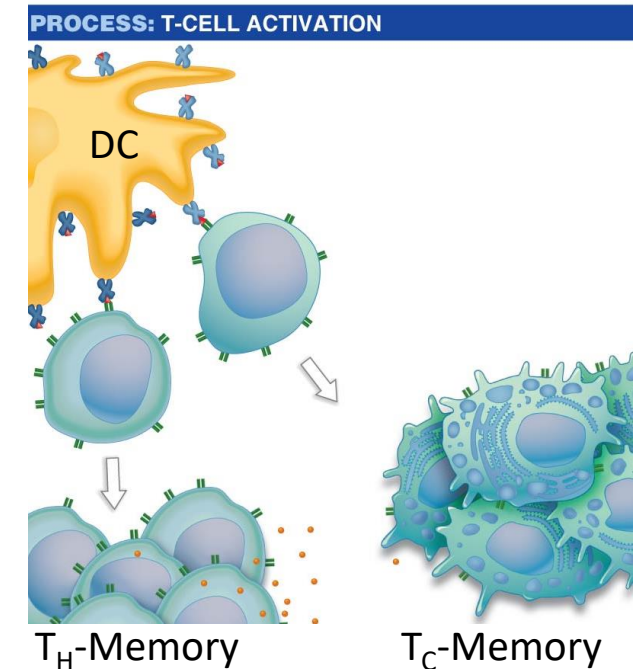
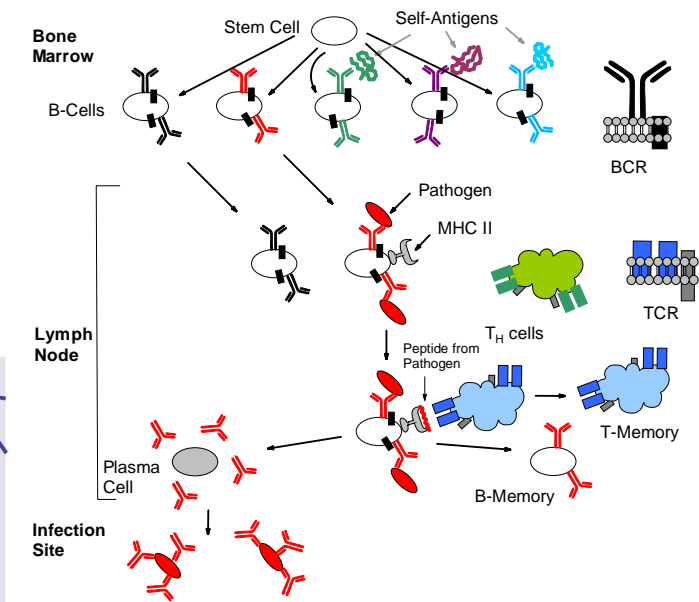
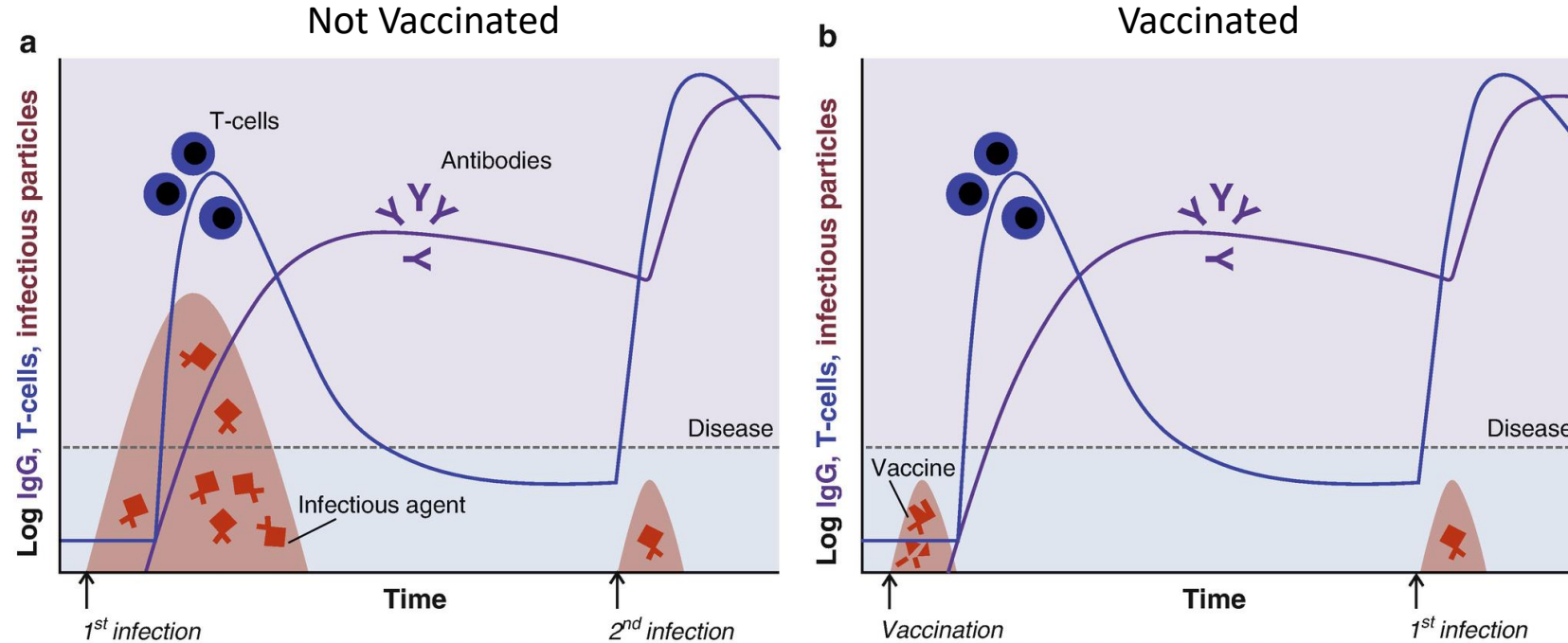


2. Active (Antigen Provided)





**Vaccine:** a vehicle containing a **form of an antigen** that is administered to induce memory B and T cells specific for that antigen.



Jiskoot W., Kersten G.F.A., Mastrobattista E., Slütter B. (2019) Vaccines. In: Crommelin D., Sindelar R., Meibohm B. (eds) Pharmaceutical Biotechnology. Springer, Cham. [https://doi.org/10.1007/978-3-030-00710-2\\_14](https://doi.org/10.1007/978-3-030-00710-2_14)

# Vaccine History

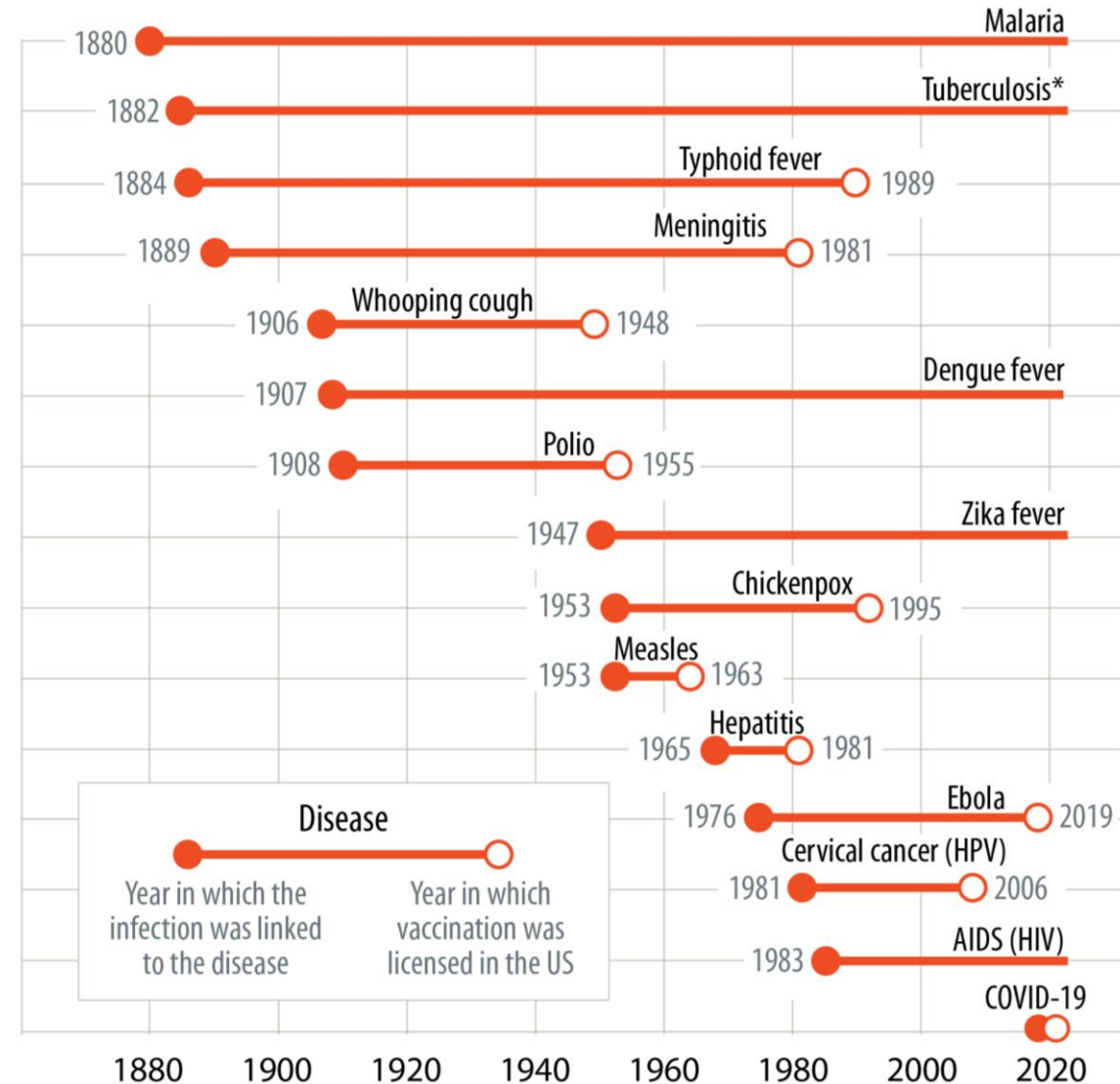
- Some diseases still do not have vaccines
- Other diseases have been eliminated by vaccination

<https://www.imf.org/en/Publications/fandd/issues/2021/12/Journey-covid-19-vaccine-Stanley>

8/26/2023

## From lab to jab

COVID-19 vaccines were developed at a speed never seen before in history.



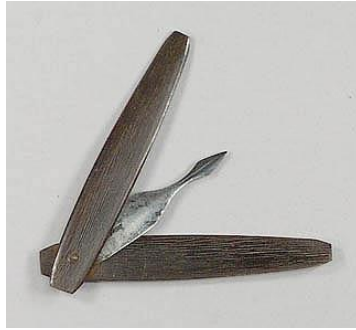
**Sources:** Our World in Data; and IMF staff analysis.

# Smallpox - A Success Story for Vaccination

*Vaccination – to introduce immunity prior to infection by pathogen*



Smallpox - 30% lethality

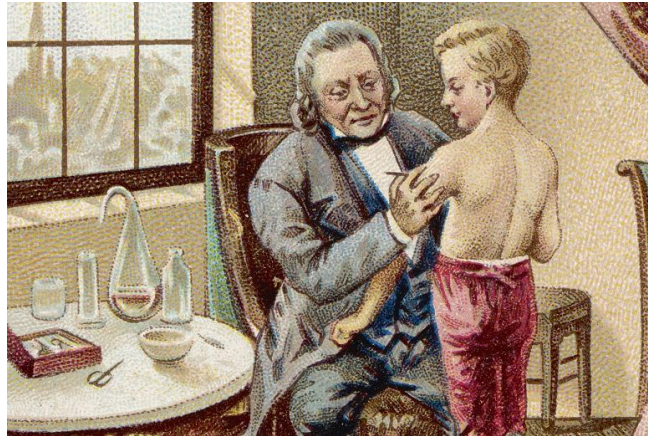
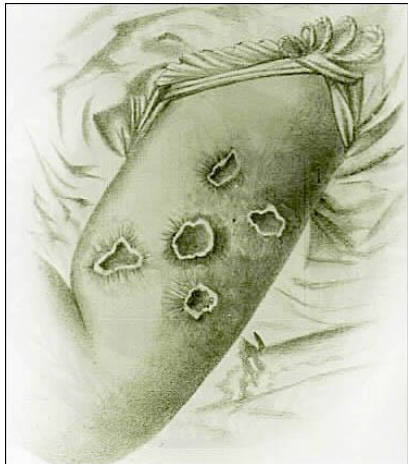


Variolation (1800) provided protection by exposing people to small amounts of smallpox virus (obtained from blisters on infected people).

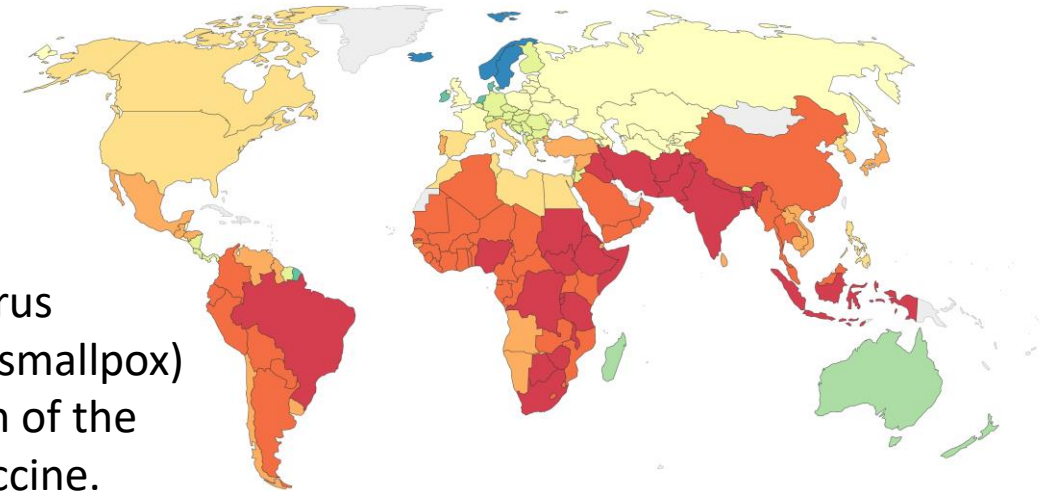
Risky because smallpox was used to vaccinate

Decade in which smallpox ceased to be endemic by country  
The decade in which smallpox was eliminated by country. Smallpox was globally eradicated in 1977.

Our W  
in D



Vaccinia virus  
(similar to smallpox)  
is one form of the  
current vaccine.



Cowpox virus causes production of antibodies against smallpox  
Jenner was the first to use cowpox to vaccinate against smallpox



# Types of Vaccines

## A. Subunit Vaccine:

A protein from the pathogen is used to induce memory cells, e.g. spike protein from the virus.


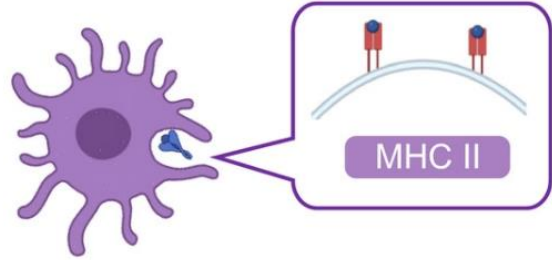

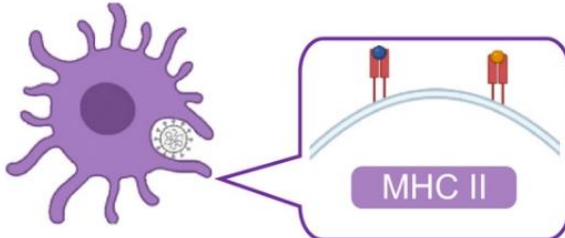
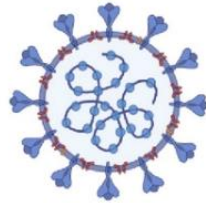
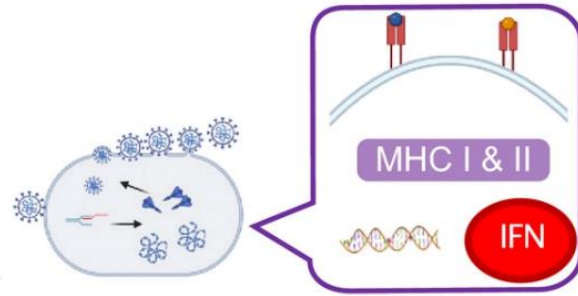
## B. Inactivated Virus

The virus is chemically inactivated before administration.

## C. Live Attenuated

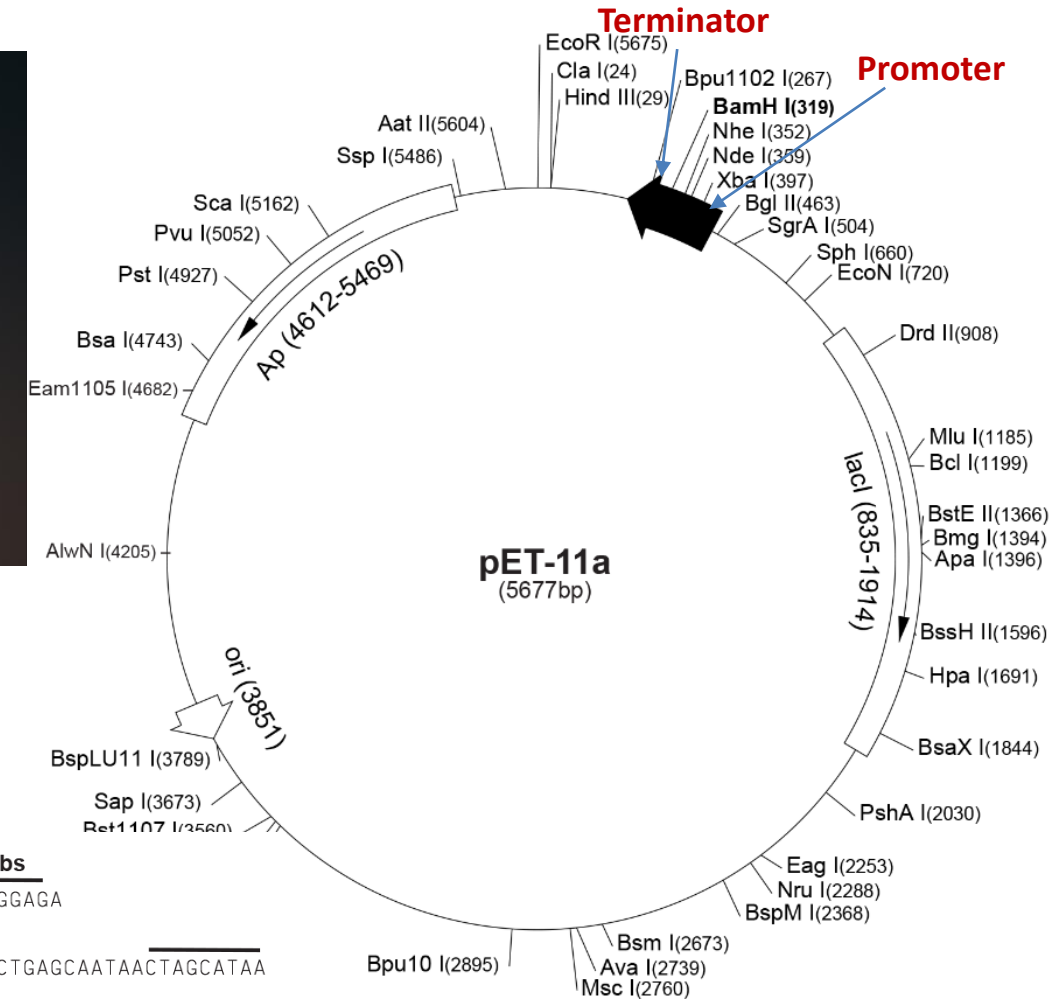
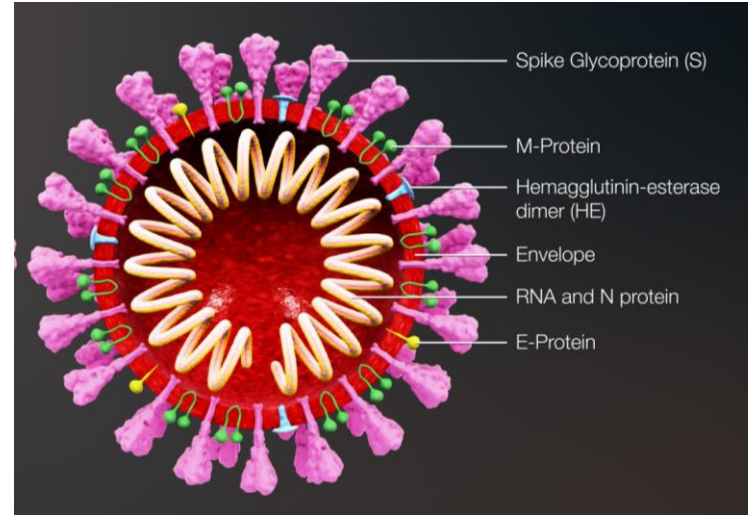
The virus is grown under conditions that select for mutant viruses that:

- i) Induce memory cells in humans
- ii) Do not cause disease symptoms

Type of vaccine	Mechanism	Advantages & disadvantages
<b>A</b> Subunit 		<div> <div>✓ Do not cause disease</div> <div>✓ Very stable</div> <div>✗ Needs booster strategy</div> <div>✗ Short memory</div> </div>
<b>B</b> Inactivated 		<div> <div>✓ Do not cause disease</div> <div>✓ Very stable</div> <div>✗ Needs booster strategy</div> <div>✗ Short memory</div> </div>
<b>C</b> Live attenuated 		<div> <div>✓ Mimic natural infection</div> <div>✓ Creates cross reactivity</div> <div>✗ Can revert and cause disease</div> <div>✗ Might be harmful in immunocompromised</div> </div>

# A. Subunit Vaccine - Producing Covid-19 Spike Protein in Bacteria

- We will incorporate the gene for the spike protein into a plasmid, which is replicated by the bacteria.
- The plasmid will also provide a promoter and mRNA termination sites so that the bacteria will make mRNA and then the mRNA will be translated into the spike protein.

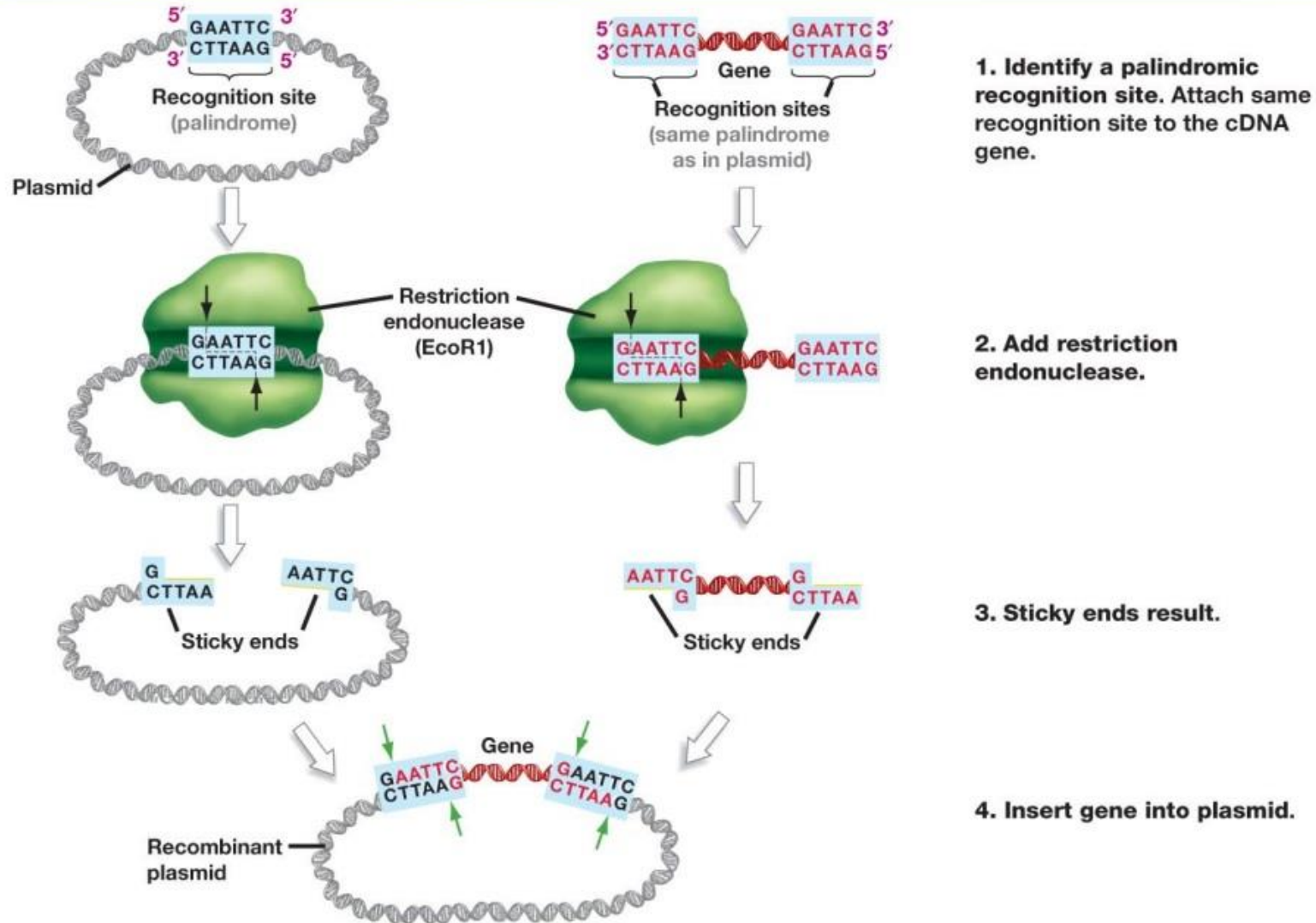


**T7 promoter** → **lac operator** **rbs**  
 AGATCTCGATCCCGCGAAATTAATACGACTCACTATAGGGGAATTGTGAGCGGATAACAATTCCCTCTAGAAATAATTTTGTTTAACTTTAAGAAGGAGA  
Nde I Nhe I BamH I  
 TATACATATGGCTAGCATGACTGGTGGAGAGCAAAATGGGTGCGCGATCCGGCTGCTAACAAAGCCCGAAAGGAAGCTGAGTTGGCTGCTGCCACCGCTGAGCAATAACTAGCATAA  
 MetAlaSerMetThrGlyGlyGlnGlnMetGlyArgGlySerGlyCysEnd  
**T7 terminator**  
 CCCCTTGGGGCCTCTAAACGGGTCTTGAGGGGTTTTTGTG

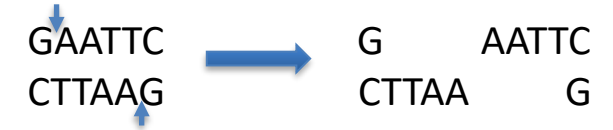
*A synthetic gene the encodes the entire spike protein will be inserted downstream from the promoter using restriction enzymes.*

# How to insert a DNA fragment into a Plasmid – Restriction enzymes

## PROCESS: INSERTING GENES INTO PLASMIDS



EcoR1



NdeI



BamHI



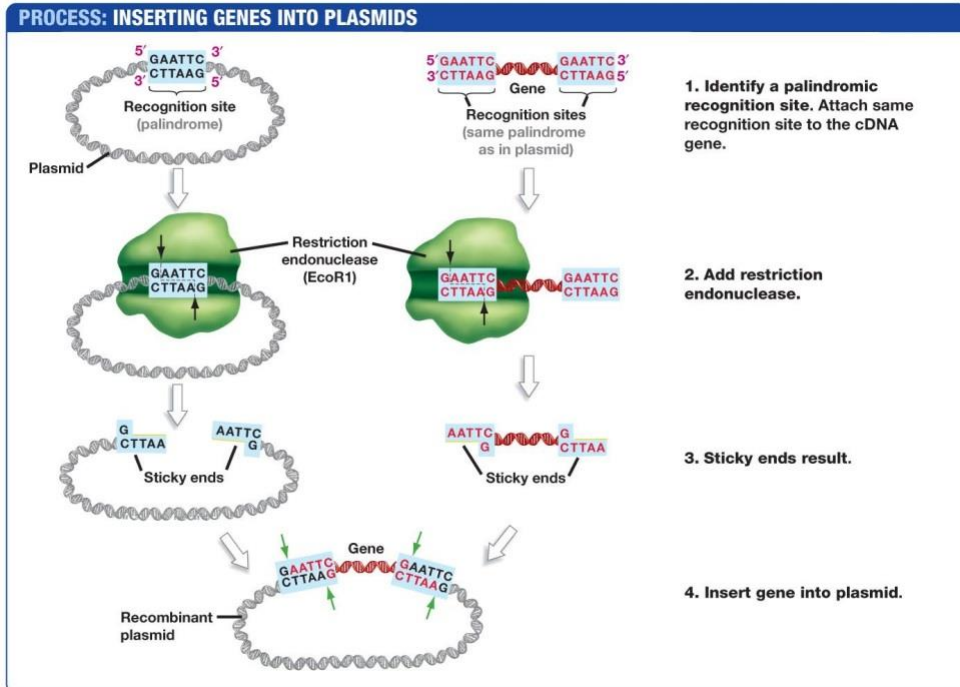
Cuts both strands, generating single-stranded DNA (sticky ends).

Complementary sticky ends can bind to each other.

DNA can be joined by DNA ligase.



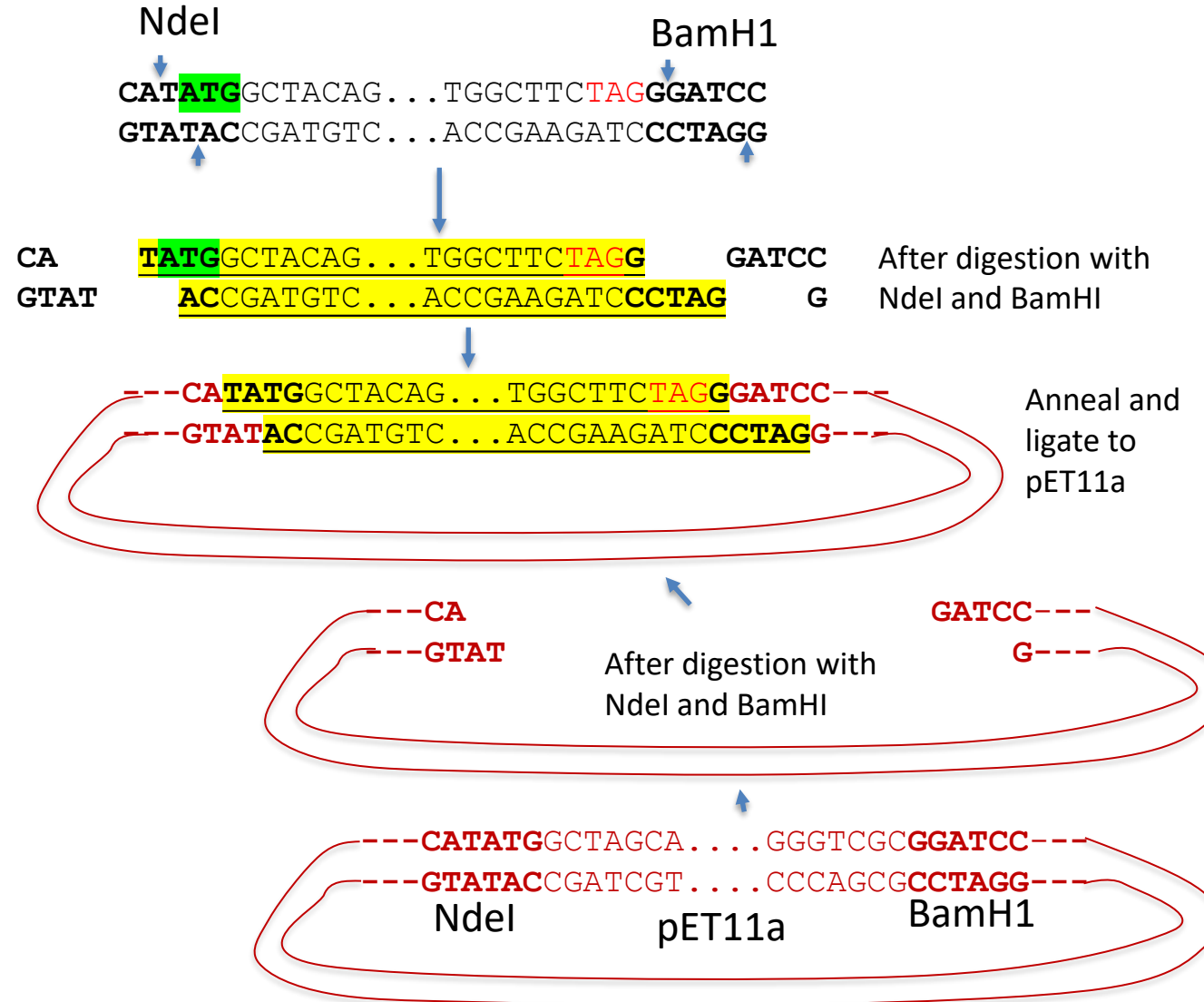
# How to insert a DNA fragment into a Plasmid



**T7 promoter** → **lac operator**  
 AGATCTCGATCCGCGAAATTAATACGACTCACTATAGGGGAATTGTGAGCGGATAACAATTCCCCT  
Nde I Nhe I BamH I  
 TATACATATGGCTAGCATGACTGGTGGACAGCAAAATGGGTCGCGGATCCGGCTGCTAACAAGCCCC  
 MetAlaSerMetThrGlyGlyGlnGlnMetGlyArgGlySerGlyCysEnd  
**T7 terminator**  
 CCCCTTGGGGCCTCTAAACGGGTCTTGAGGGGTTTTTTTG

- Cut plasmid and synthetic gene with both NdeI and BamHI enzymes to make sticky ends.
- Cool to allow sticky ends to anneal TA from NdeI will anneal, GATC from BamHI will anneal.
- DNA ligase to join fragments.

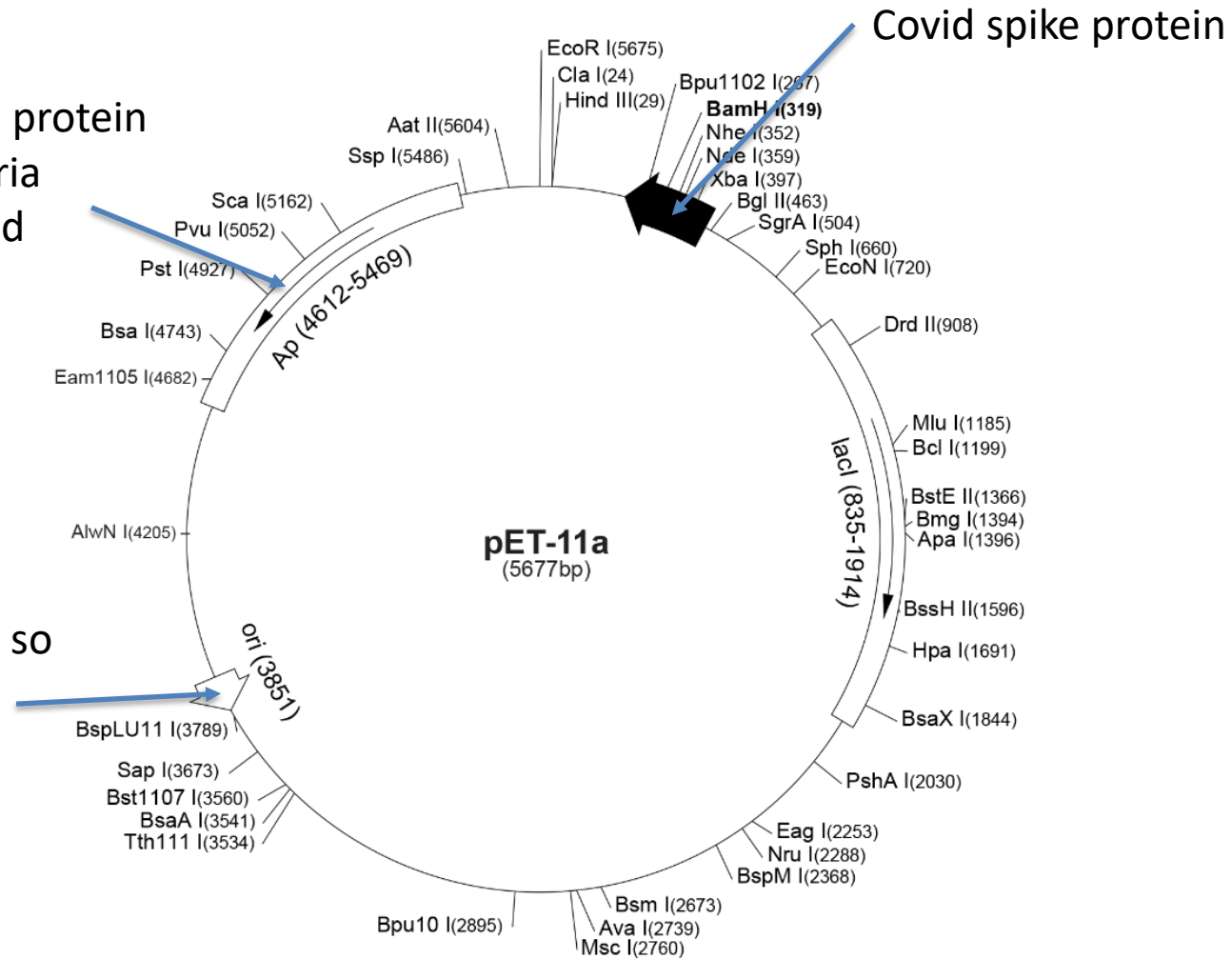
## Synthetic spike protein gene



# Final Product – Covid-19 Spike protein codons in plasmid

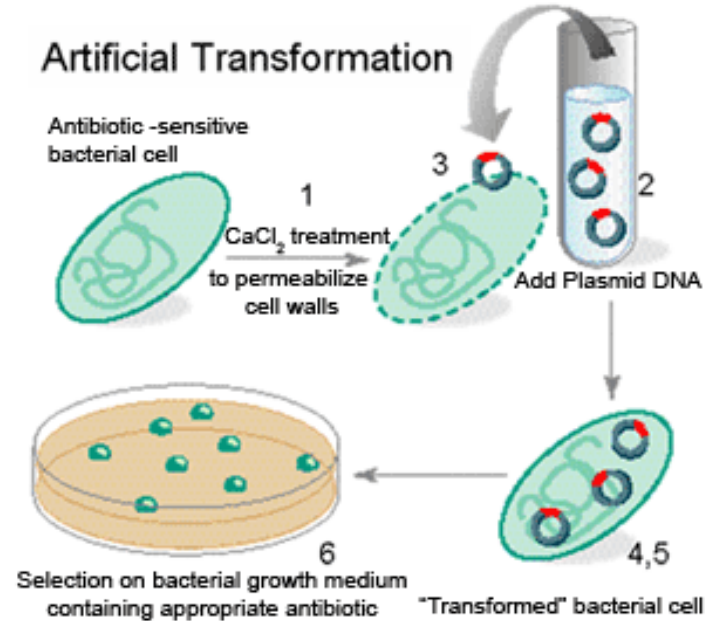
Gene that codes for a protein that makes the bacteria containing the plasmid resistant to penicillin.

Origin of replication so that the plasmid is copied when the bacteria divide.




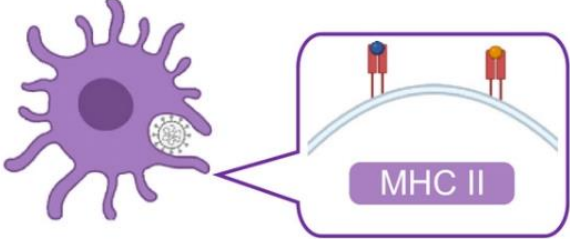




# Inserting the Plasmid into Bacterial Cells

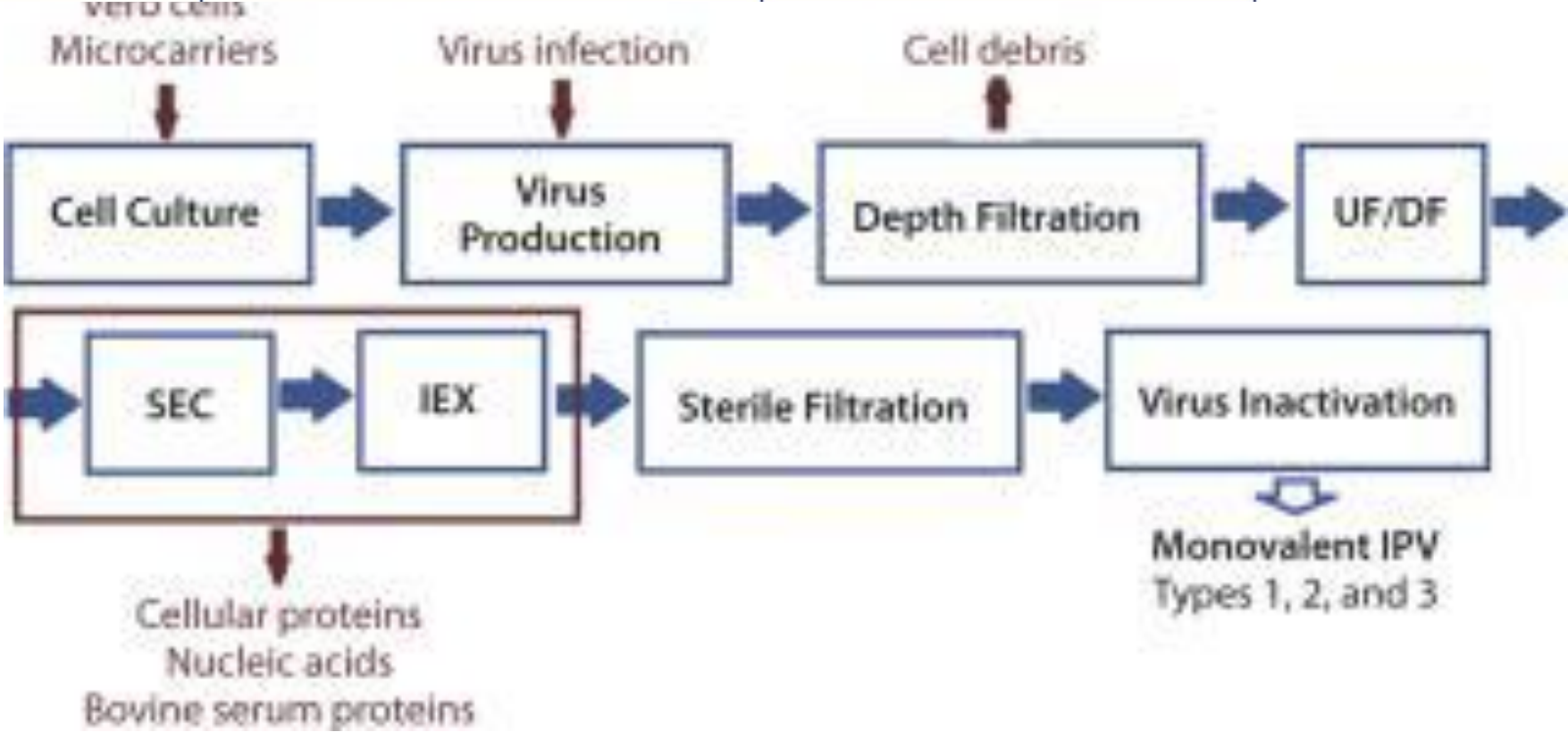
- Cells can take up DNA from the environment and incorporate it into their genomes in a process called transformation.
- To transform bacterial cells with a plasmid, researchers increase the permeability of the cell's membrane using a chemical treatment (calcium chloride).
- Cells that contain the plasmid are selected by growth on ampicillin.
- Only those cells with plasmid can grow.



- The plasmid with the cDNA produces the spike protein in bacteria.
- Large amounts of E.coli that produce the vaccine can be grown in a fermenter.
- The spike protein can be used as a vaccine

# B. Inactivated Viruses

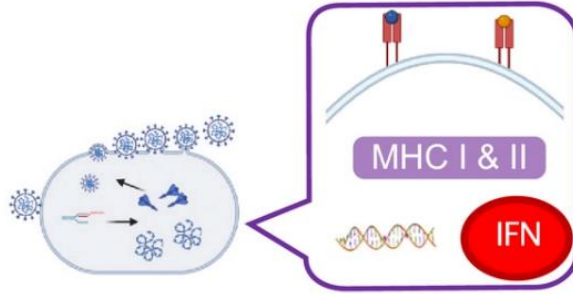
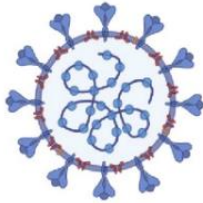
<p>Inactivated</p> 		<div> <div>  <p>Do not cause disease</p> </div> <div>  <p>Very stable</p> </div> </div> <div> <div>  <p>Needs booster strategy</p> </div> <div>  <p>Short memory</p> </div> </div>
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# C. Attenuated Viruses

Live attenuated

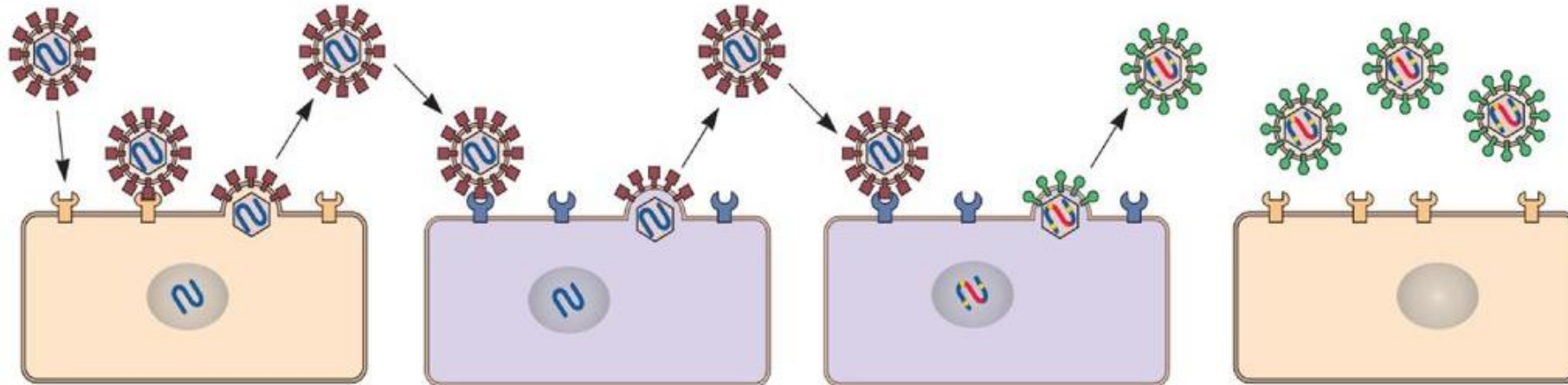
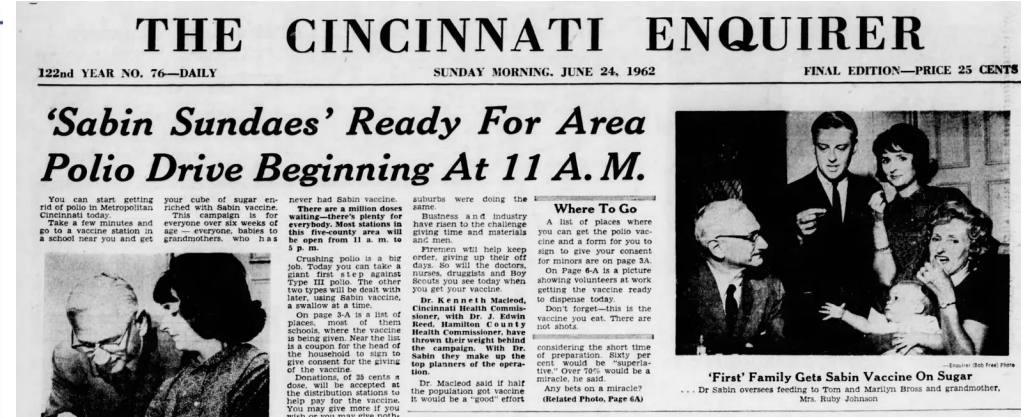


Mimic natural infection

Creates cross reactivity

Can revert and cause disease

Might be harmful in immunocompromised



Pathogenic virus is isolated from a patient and grown in human cultured cells

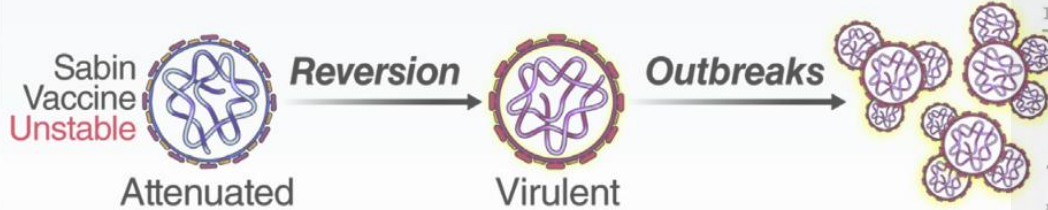
The cultured virus is used to infect monkey cells

The virus acquires many mutations that allow it to grow well in monkey cells

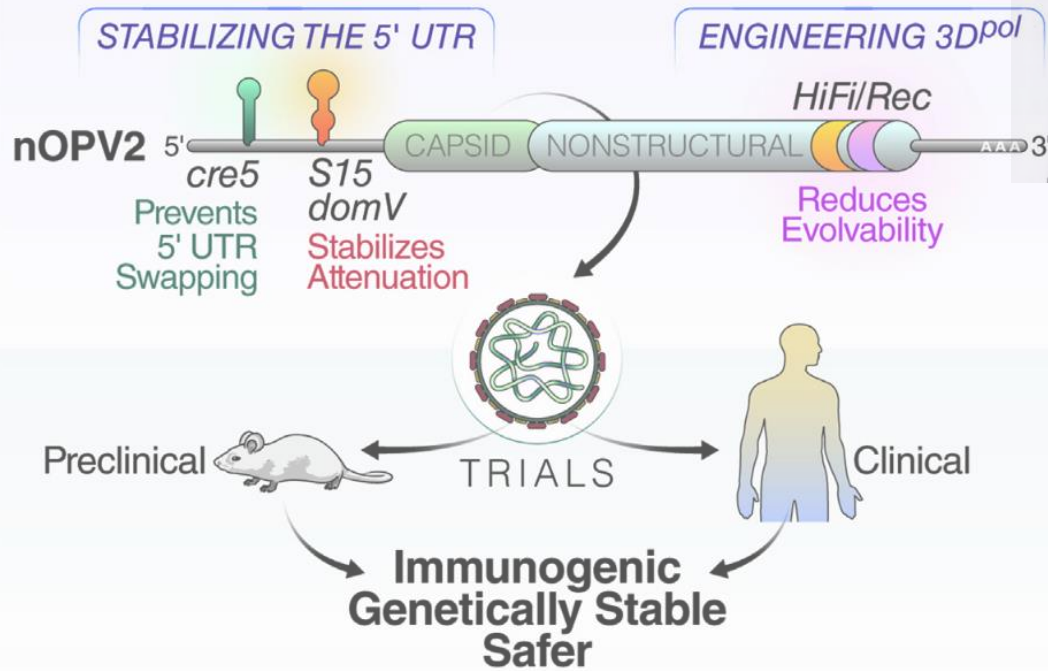
The virus no longer grows well in human cells and may be a candidate for a vaccine

# C. Attenuated Viruses

## CURRENT LIVE-ATTENUATED POLIO VACCINE



## POLIO VACCINE REDESIGN



## THE CINCINNATI ENQUIRER

122nd YEAR NO. 76—DAILY

SUNDAY MORNING, JUNE 24, 1962

FINAL EDITION—PRICE 25 CENTS

### 'Sabin Sundaes' Ready For Area Polio Drive Beginning At 11 A. M.

You can start getting rid of polio in Metropolitan Cincinnati today. This campaign is for everyone over six weeks of age — everyone, babies to grandmothers, who has

never had Sabin vaccine. There are a million doses waiting—there's plenty for everybody. Most stations in this five-county area will be open from 11 a. m. to 5 p. m.

Crushing polio is a big job. Today you can take a giant first step against Type III polio. The other two types will be dealt with later, using Sabin vaccine, a swallow at a time.

On page 3-A is a list of places, most of them schools, where the vaccine is being given. Near the list is a coupon for the head of the household to sign to give consent for the giving of the vaccine.

Donations, of 25 cents a dose, will be accepted at the distribution stations to help pay for the vaccine. You may give more if you wish on any other other matter.

suburbs were doing the same.

Business and industry have risen to the challenge giving time and materials and men.

Firemen will help keep order, giving up their off days. So will the doctors, nurses, druggists and Boy Scouts you see today when you get your vaccine.

Dr. Kenneth Macleod, Cincinnati Health Commissioner, with Dr. J. Edwin Reed, Hamilton County Health Commissioner, have thrown their weight behind the campaign. With Dr. Sabin they make up the top planners of the operation.

Dr. Macleod said if half the population got vaccine it would be a "good" effort

#### Where To Go

A list of places where you can get the polio vaccine and a form for you to sign to give your consent for minors are on page 3A.

On Page 6-A is a picture showing volunteers at work getting the vaccine ready to dispense today.

Don't forget—this is the vaccine you eat. There are not shots.

considering the short time of preparation. Sixty per cent would be "superlative." Over 70% would be a miracle, he said.

Any bets on a miracle? (Related Photo, Page 6A)



'First' Family Gets Sabin Vaccine On Sugar  
... Dr. Sabin oversees feeding to Tom and Marilyn Bross and grandmother, Mrs. Ruby Johnson

## Cell Host & Microbe

Volume 27, Issue 5, 13 May 2020, Pages 736-751.e8

Article

### Engineering the Live-Attenuated Polio Vaccine to Prevent Reversion to Virulence

Ming Te Yeh<sup>1</sup>, Erika Bujaki<sup>2</sup>, Patrick T. Dolan<sup>1</sup>, Matthew Smith<sup>2</sup>, Rahnuma Wahid<sup>3</sup>, John Konz<sup>3</sup>, Amy J. Weiner<sup>4</sup>, Ananda S. Bandyopadhyay<sup>4</sup>, Pierre Van Damme<sup>5</sup>, Ilse De Coster<sup>5</sup>, Hilde Revets<sup>5</sup>, Andrew Macadam<sup>2</sup>, Raul Andino<sup>1,6</sup>



## D – Virus Like Particles:

Proteins isolated from the virus form virus like particles, *without* the genetic material of the virus

## E. Recombinant Virus:

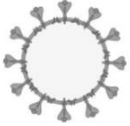
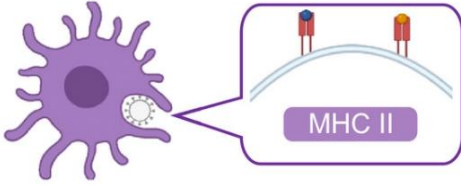
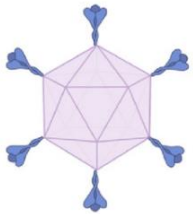
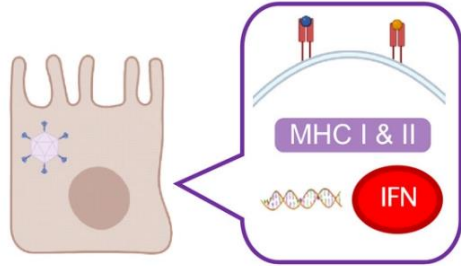

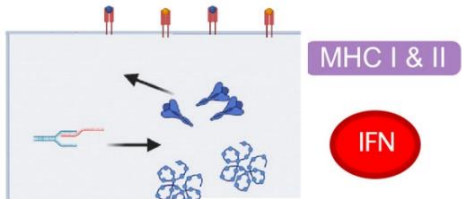
A “safe virus” is used (e.g. cold virus)  
Gene for a protein from a pathogen is inserted into the DNA of the virus.

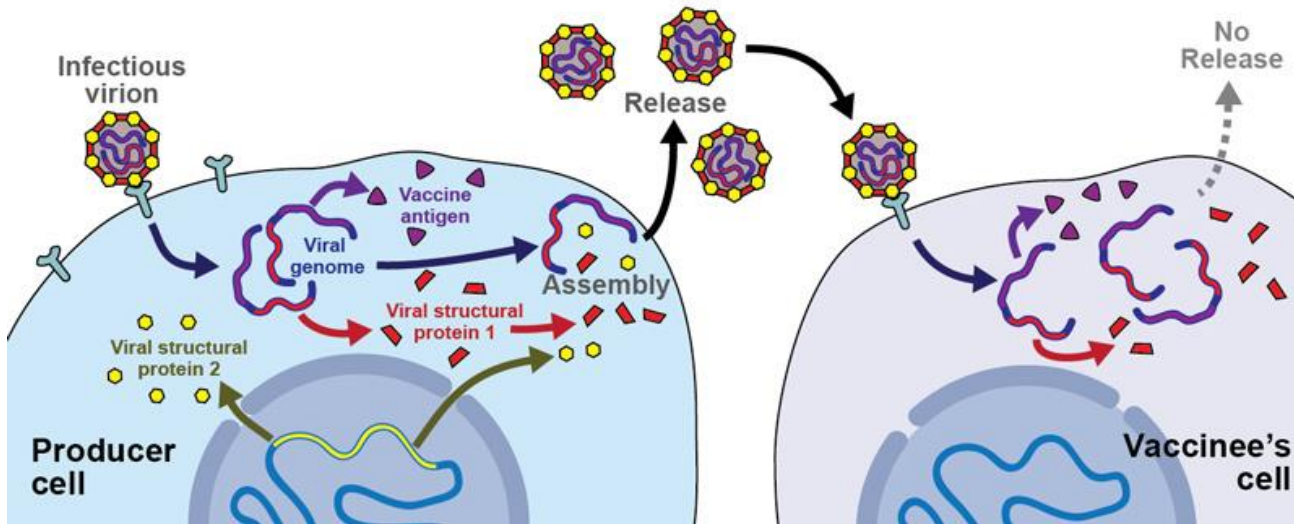
- When virus grows it produces the protein from the pathogen generating immunity.

Also includes vaccines that are a mixture of genetic material from human and animal viruses (reassortment viruses)

## F. RNA Vaccines (Pfizer Covid Vaccines)

RNA coding for a viral protein is introduced into cells. The RNA is used by the cell to make viral proteins, inducing an immune response.

Type of vaccine	Mechanism	Advantages & disadvantages
<b>D</b> Virus like particles 		<div> <div>✓ Increased uptake by lymph node</div> <div>✓ Do not cause disease</div> <div>✗ Dependant on efficient expression platform</div> <div>✗ Difficult to make VLP stable in long term</div> </div>
<b>E</b> Recombinant viruses 		<div> <div>✓ Mimics natural infection</div> <div>✓ Strong memory</div> <div>✓ Cannot revert to natural disease</div> <div>✗ Pre-existent memory against vector lowers efficacy</div> <div>✗ Recombination with other viruses</div> </div>
<b>F</b> RNA vaccines 		<div> <div>✓ Easy to modify</div> <div>✓ Do not cause disease</div> <div>✗ Short immune memory if not stable</div> <div>✗ Low immune priming if efficacy of delivery is low</div> </div>

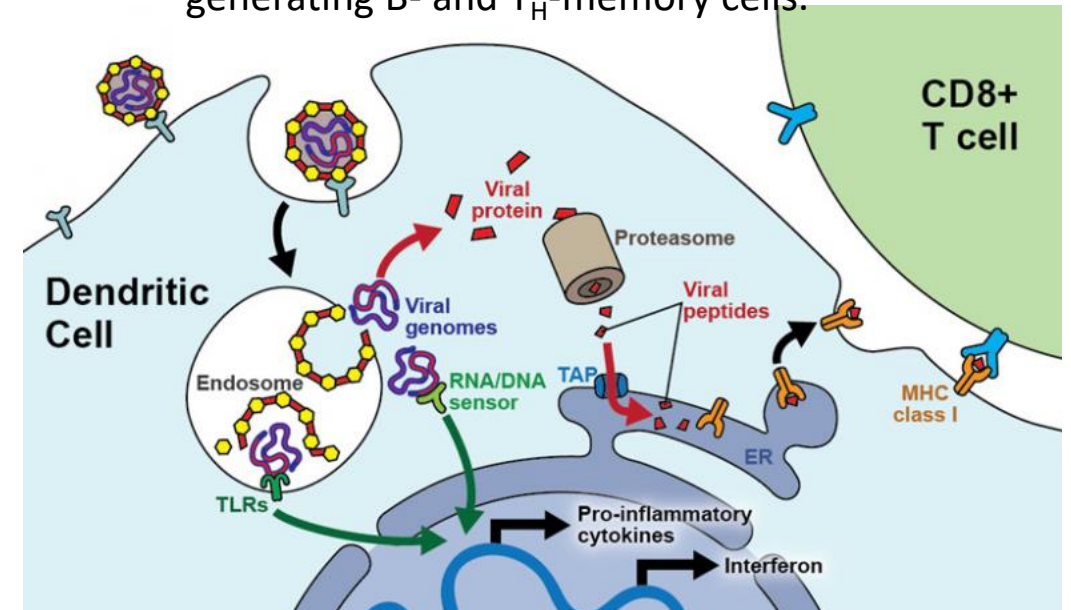


## 1. Production of vaccine

- I. Genes from the pathogenic virus are added to an adenovirus (common cold)
- II. Adenovirus is defective and cannot replicate without key structural proteins that are provided in the producer cell.
- III. No viral particles are released in the vaccinee's cells.

## 2. Action of vaccine

- a) Virus infects host cell in vaccinated person.
- b) Viral genome is used to make viral proteins, including proteins from the pathogen.
- c) Activate  $T_C$  cells to become  $T_C$  – memory cells (can be re-activated by MHC I + Peptide).
- d) B-cell response can occur due to antigens that are sent to the surface of the cell, generating B- and  $T_H$ -memory cells.




# Herd Immunity:


- Vaccinated individuals prevent disease from spreading from sick to unvaccinated.
- At sufficient levels, the “herd” is immune because the virus cannot spread.


**High risk  
Can't be  
vaccinated  
(too young,  
immune -  
compromised)**

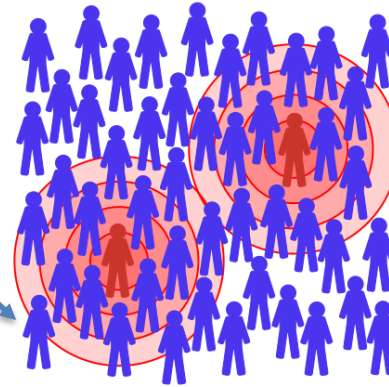
**Below  
herd  
immunity**

**At herd  
immunity**

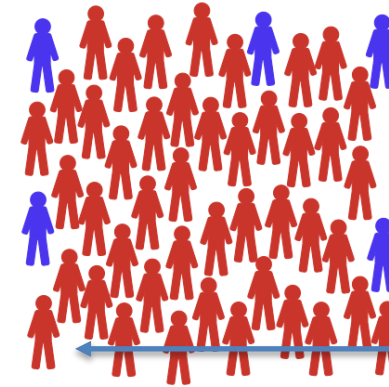
 = not immunized, but still healthy

 = immunized and healthy

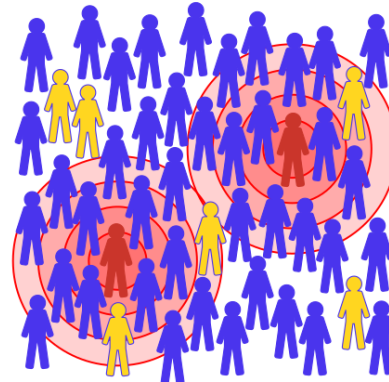
 = not immunized, sick, and contagious



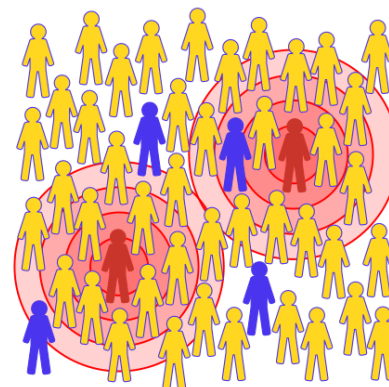
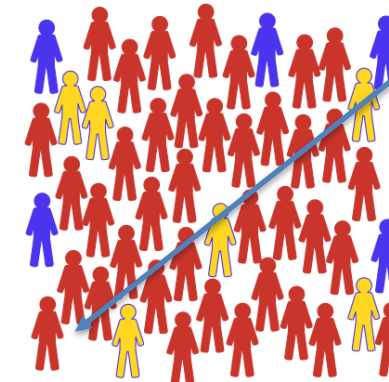
No one is immunized.  
Contagious disease spreads through the population.



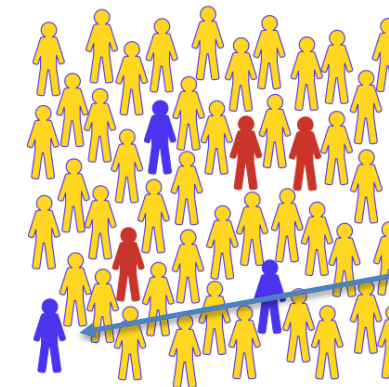
**Gets infected**



Some of the population gets immunized.  
Contagious disease spreads through some of the population



Most of the population gets immunized.  
Spread of contagious disease is contained.



**Protected**

# Herd Immunity

How Many People need to be vaccinated to achieve herd immunity?

10% ?

20% ?

50% ? *It depends on the how infectious the virus is*

90% ?

100% ?

## Our Experimental Viruses:

Ebola: Low infectivity

Polio: Moderate infectivity

Measles: High infectivity



# Simulation to Determine Infectivity Versus Vaccination Level

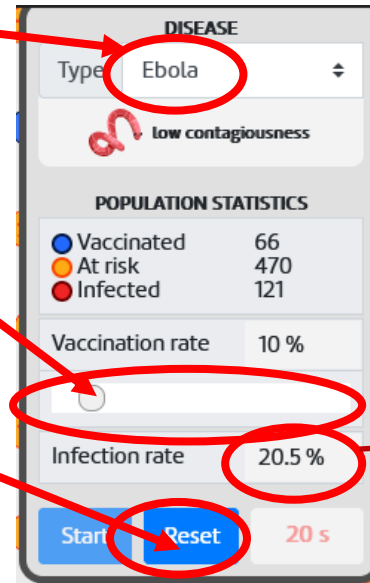
1. Go to the following web site and open **both** links: <http://www.andrew.cmu.edu/~rule/stayin-alive>

2. On the **Infection Simulator** link, scroll down (2/3 page) to the image of the plane, and click on it.



3. Select the virus for your group

4. Use the slider to select the different vaccination levels. For each of the vaccination levels do **three** simulations. Enter the value for the **% Infection rate at 20s** into the appropriate cell of the google sheet. Your data will be automatically averaged and plotted.



Group Number	Virus	% Vaccination
1	Ebola	10, 20, 40, 50
2	Ebola	70, 80, 90
2	Polio	10, 20, 40, 50
3	Polio	70, 80, 90
4	Measles	10, 20, 40, 50
5	Measles	70, 80, 90

**3 Simulation  
Runs at 10%**

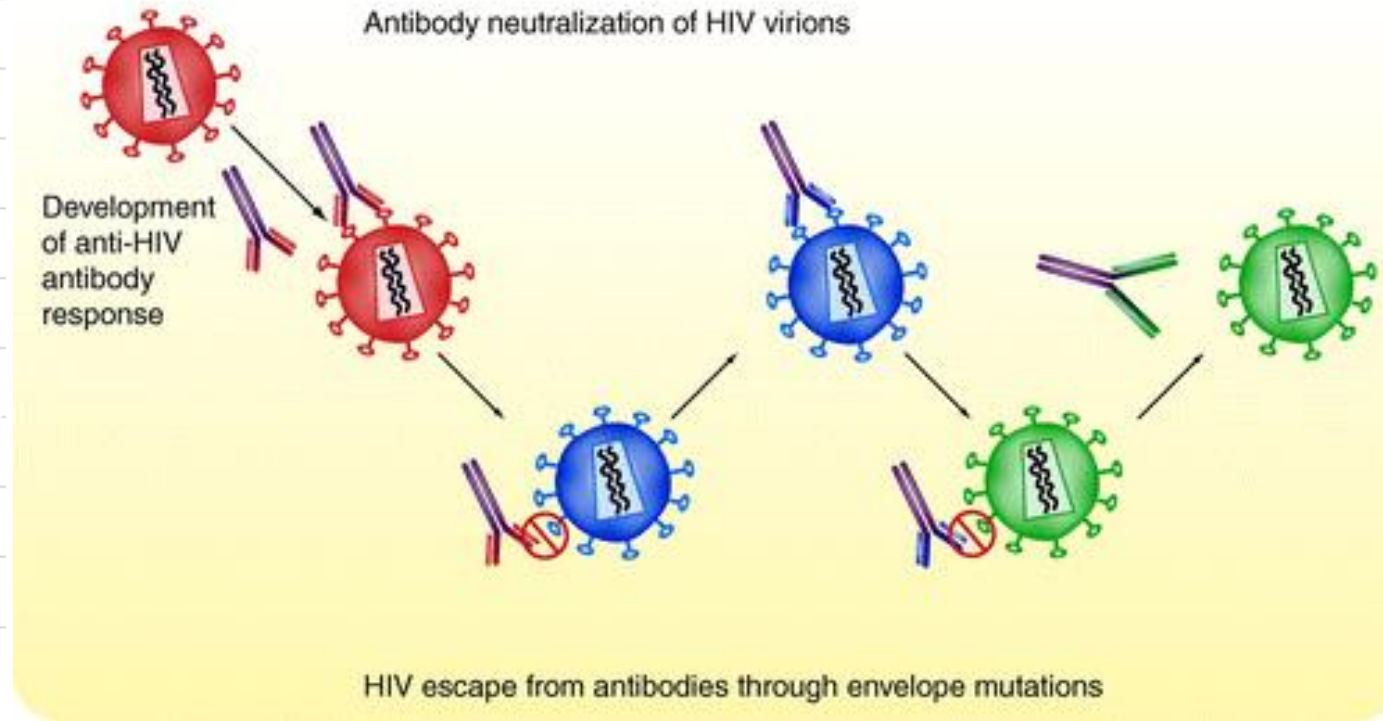
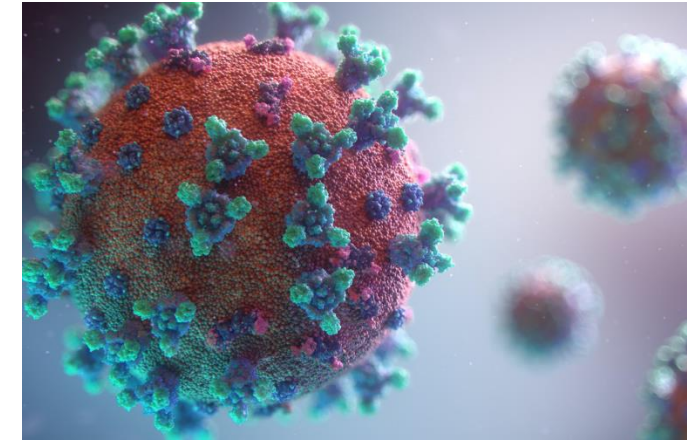
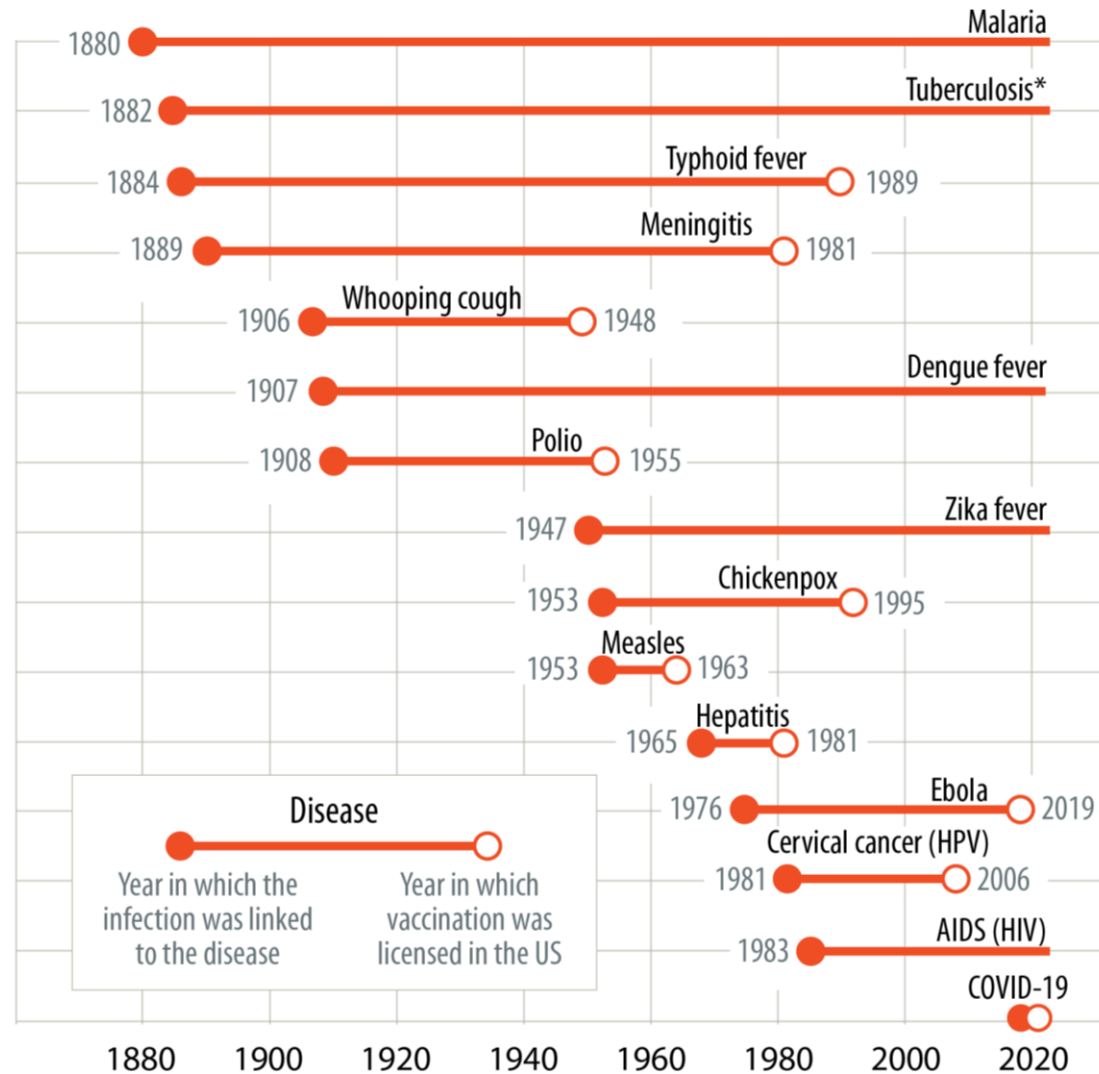
A	B	C	D	E	F	G	H	I	J	K	L	M
%Vaccinated	Ebola (Ave)	Polio (Ave)	Measles (Ave)	#1	#2	#3	#1	#2	#3	#1	#2	#3
10	#DIV/0!	#DIV/0!	#DIV/0!									
20	#DIV/0!	#DIV/0!	#DIV/0!									
40	#DIV/0!	#DIV/0!	#DIV/0!									
50	#DIV/0!	#DIV/0!	#DIV/0!									
70	#DIV/0!	#DIV/0!	#DIV/0!									
80	#DIV/0!	#DIV/0!	#DIV/0!									
90	#DIV/0!	#DIV/0!	#DIV/0!									



# Why Are There No Vaccines for HIV?

## From lab to job

COVID-19 vaccines were developed at a speed never seen before in history.



# Summary Questions for Immunology:

1. What are the two major branches of the immune system? Why are both important?
2. What are the roles of different cell types in each system, e.g. what would happen if  $T_H$ -cells disappeared?
3. What is the quaternary structure of an antibody? Can you sketch an antibody and indicate where the antigen binds?
4. What defines the specificity of antibodies?
5. What are the steps in the production of antibody genes, at the molecular level:
  - a) How do DNA rearrangements produce functional heavy and light chain genes
  - b) How are the mature mRNA generated in B-cells and Plasma cells.
  - c) What is the difference between the heavy chain export process for B-cells and plasma cells.
6. Can you describe how antibodies kill/inactivate pathogens
7. How are virally infected cells and tumor cells recognized by  $T_c$  cells?
8. How does the  $T_c$  cell kill those cells?
9. What evasion mechanisms are used by cancer cells and how have these been addressed by antibody therapy?
10. What was the origin of the idea for vaccination?
11. What was one of the first “safe” vaccines? What disease has now been eradicated due to this vaccine?
12. Why is it important to be vaccinated (Herd immunity)
13. Can you describe one way to generate a vaccine for a pathogen? Do you know the pros and cons for that method?