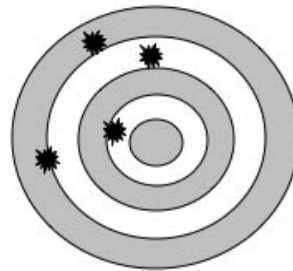


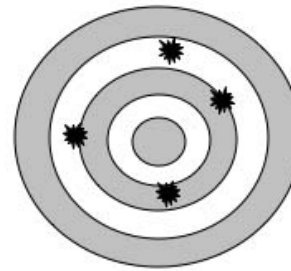
# Bioimage Informatics

Lecture 2, Spring 2012

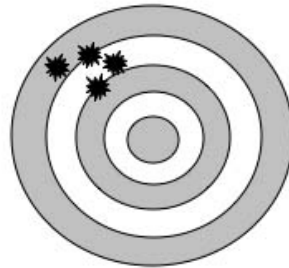
## Fundamentals of Light Microscopy



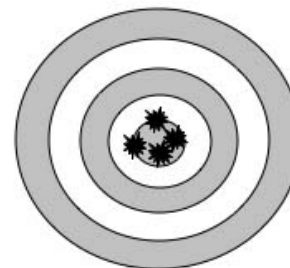
**Not Accurate  
Not Precise**



**Accurate  
Not Precise**



**Not Accurate  
Precise**



**Accurate  
Precise**

[http://celebrating200years.noaa.gov/magazine/tct/tct\\_side1.html](http://celebrating200years.noaa.gov/magazine/tct/tct_side1.html)

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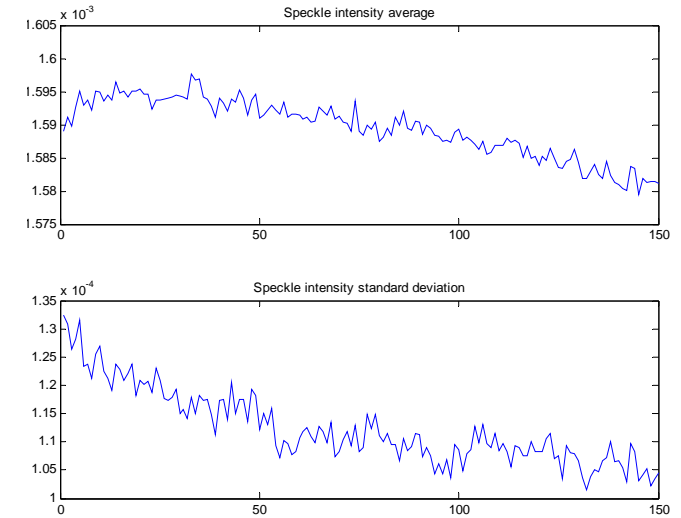
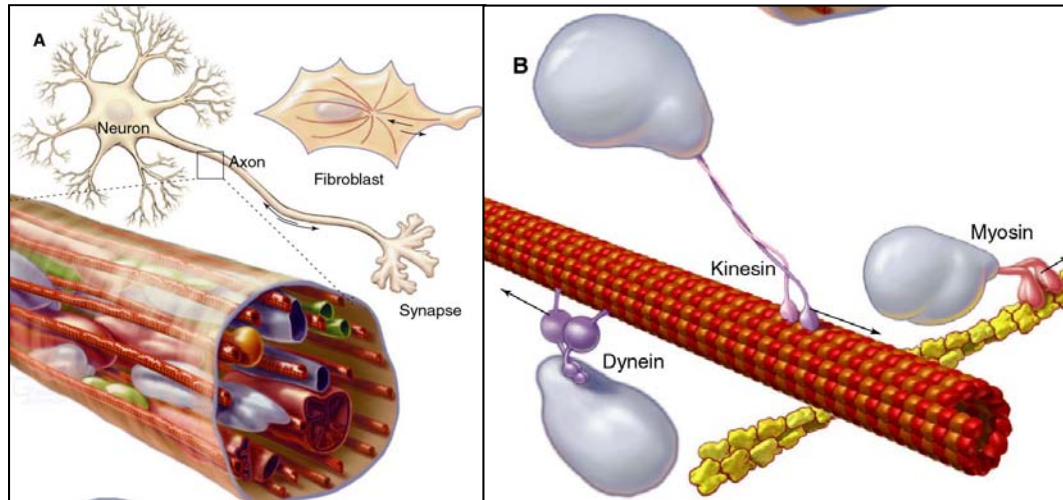
# Outline

---

- Importance of understanding and optimizing image formation
- Some basic optics facts
- Light microscope structure
- Contrast generation in microscopy
- Practical considerations of microscopy

- 
- Importance of understanding and optimizing image formation
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# Understanding Image Formation



axonal transport of APP vesicles

<http://micro.magnet.fsu.edu/primer/java/fluorescence/photobleaching/index.html>

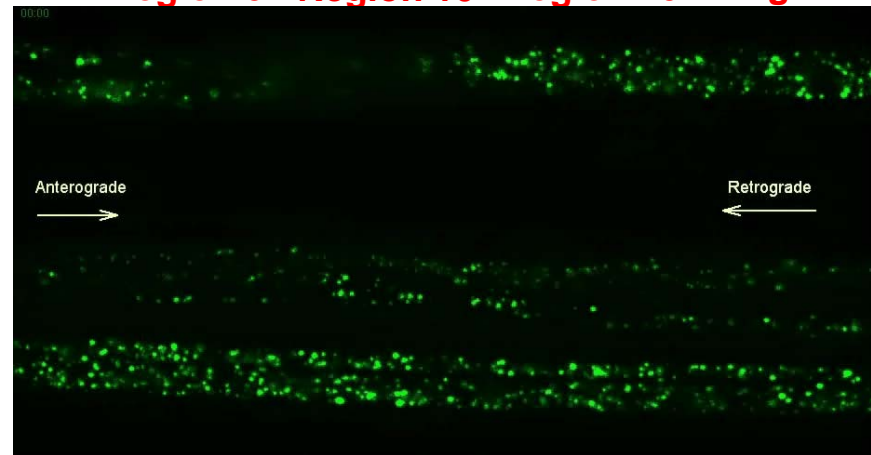
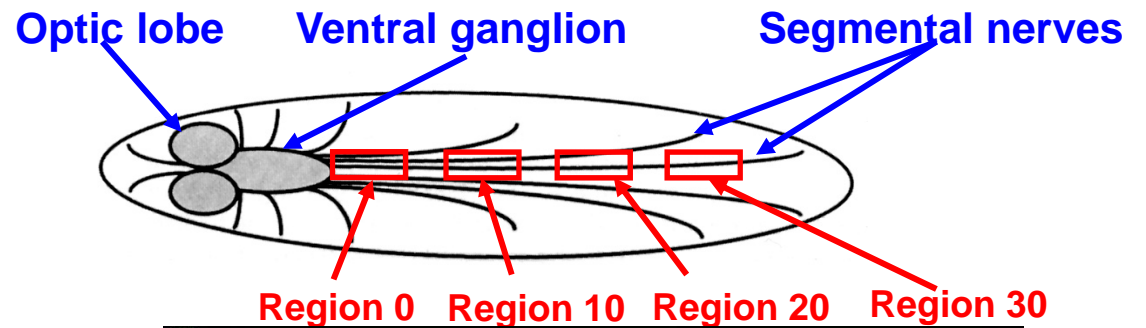
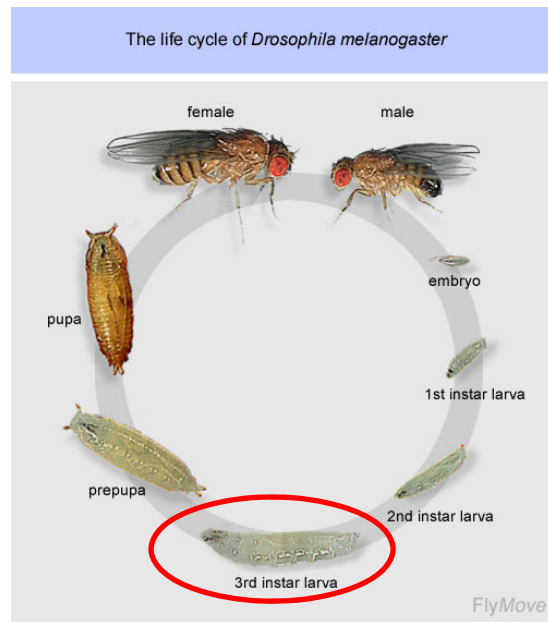
- Photobleaching results in gradual decreasing of image signal intensities over time under repetitive exposure.

# Optimizing Image Formation

---

- Bioimage data collection and data analysis must be collaborative processes.
  - Images that are improperly collected cannot be analyzed.  
E.g. violation of Nyquist sampling
  - Optimization of image collection can significantly simplify data analysis.  
This requires concurrent design of imaging and data analysis.

# Example: Imaging Axonal Cargo Transport



Saxton lab, UC Santa Cruz



axonal transport of human APP-YFP vesicles; 0.1 sec/frame

10  $\mu$ m

# Relation between Image Collection and Image Analysis

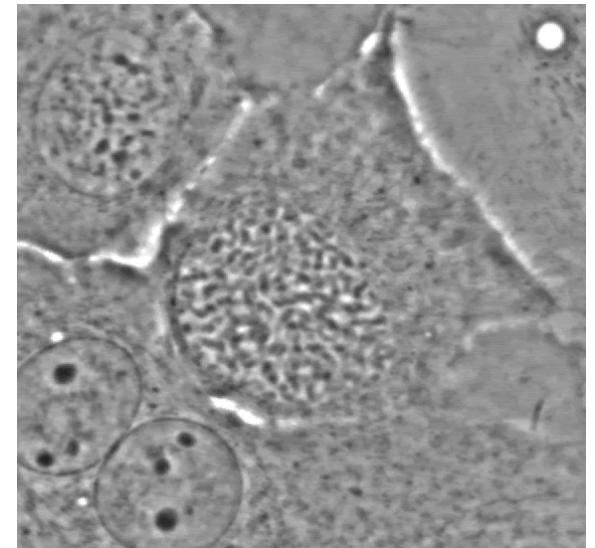
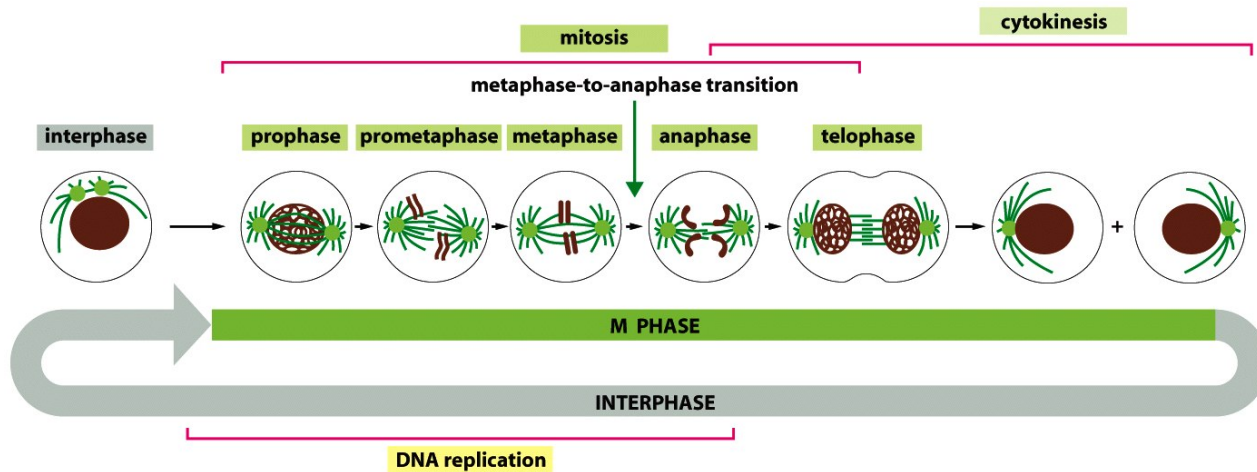
---

- Bioimage data collection and data analysis must be collaborative processes.
  - Image processing and computer vision can significantly reduce challenges in image collection.  
This is one of the purposes of this class.



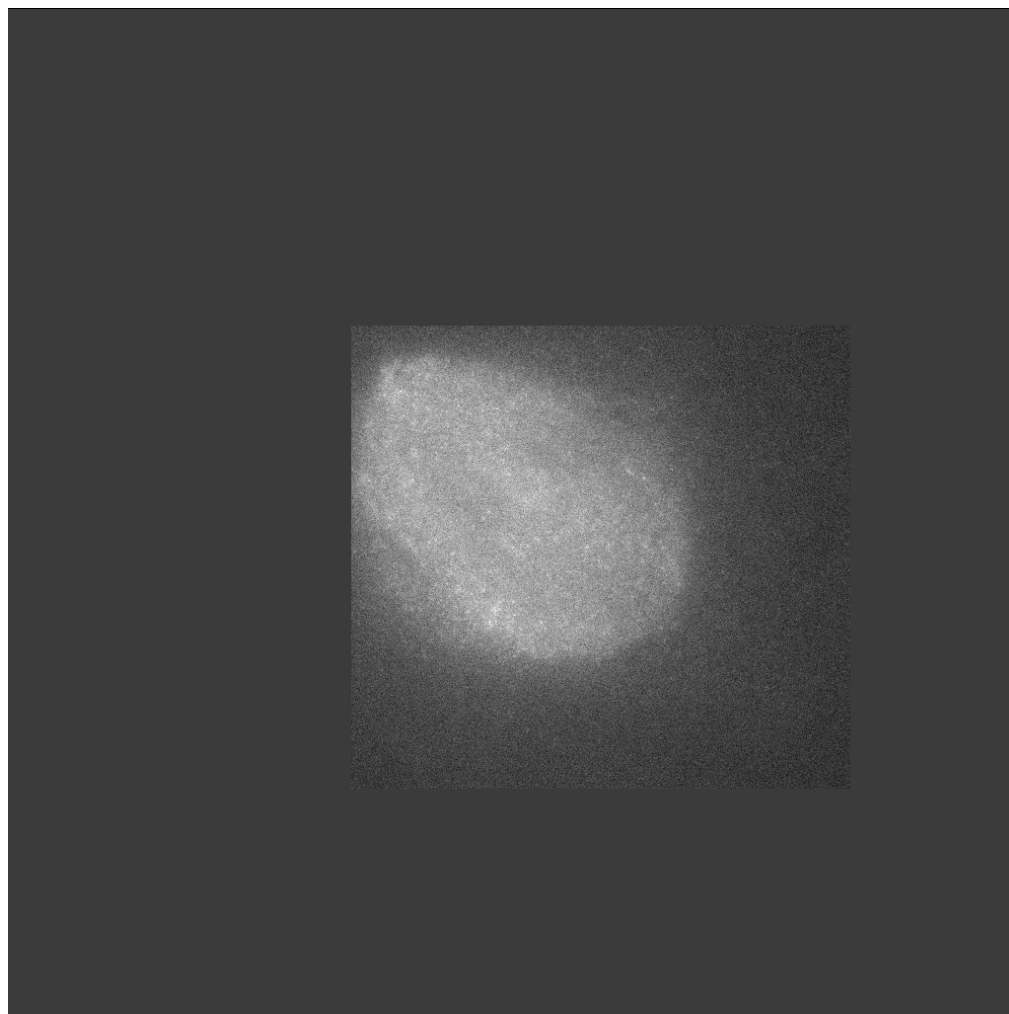
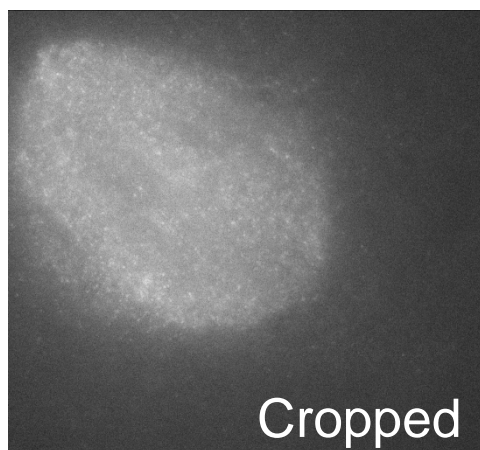
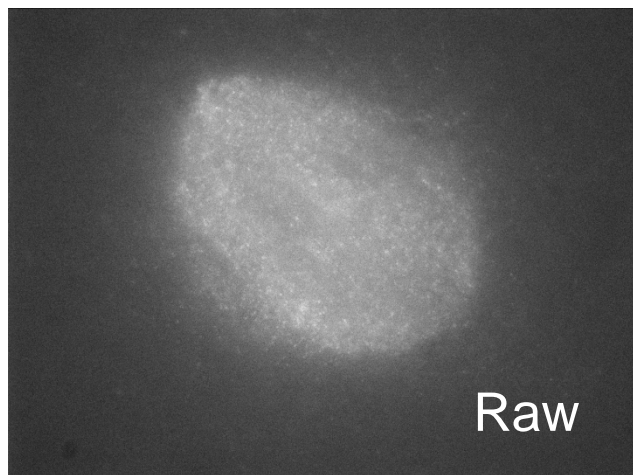
# Example: Image Alignment/Registration

- Alignment of mitotic spindle images
- Introduction to the mitotic spindle



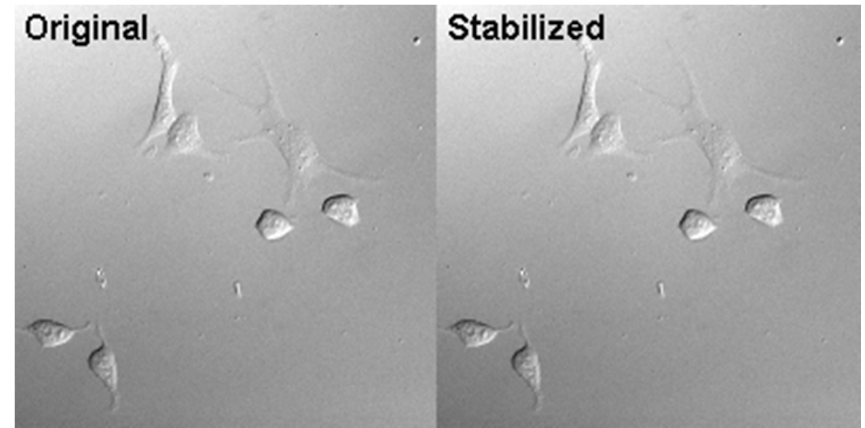
# Demo I: Image Alignment

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# Demo II: Image Alignment

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[http://www.cs.cmu.edu/~kangli/code/Image\\_Stabilizer.html](http://www.cs.cmu.edu/~kangli/code/Image_Stabilizer.html)

# Summary

---

- Image collection and image analysis should be collaborative processes.
- Correct collection of image data is essential to subsequent data analysis.
- Computational image analysis can help overcome some of the challenges in image collection.
- Understanding image formation is essential to image analysis.
- Proper design of image collection can significantly simplify subsequence.

- 
- Importance of understanding and optimizing image formation
  - **Some basic optics facts**
  - Light microscope structure
  - Contrast generation in microscopy
  - Practical considerations of microscopy

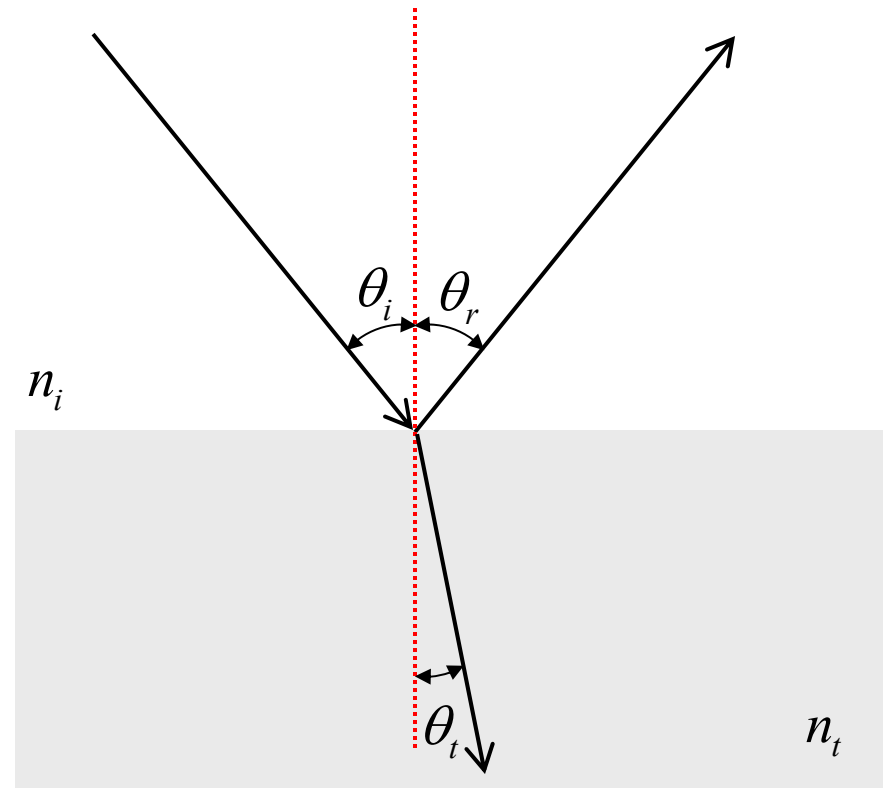
# Reflection & Refraction

- Law of reflection

$$\theta_i = \theta_r$$

- Law of refraction  
(Snell's law)

$$n_i \sin \theta_i = n_t \sin \theta_t$$



# Refractive Index

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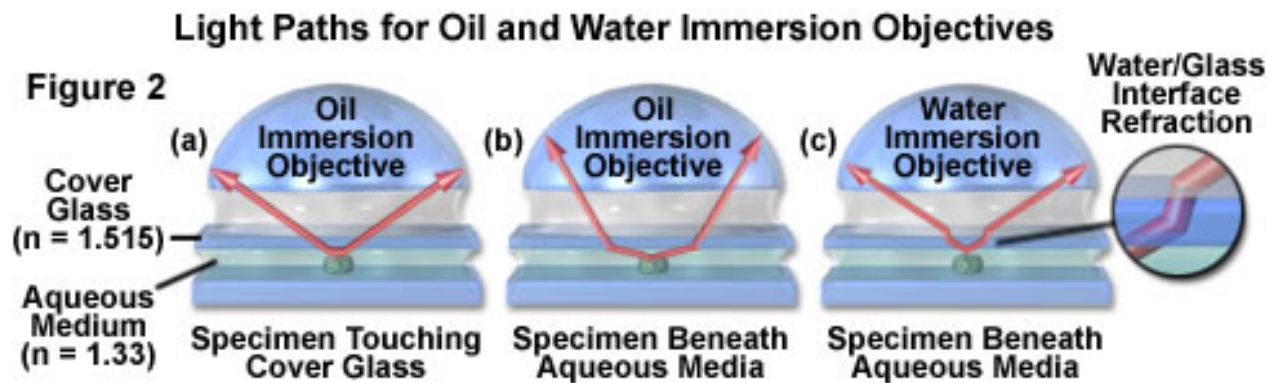
- Absolute refractive index of a material

$$\frac{\text{velocity of electromagnetic wave in vacuum}}{\text{velocity of electromagnetic wave in the material}}$$

- Air 1.000293
- Refractive index (also called relative refractive index)
  - Water 1.333
  - Glass ~1.50
  - Immersion oil 1.51

$$n_{12} = \frac{n_2}{n_1}$$

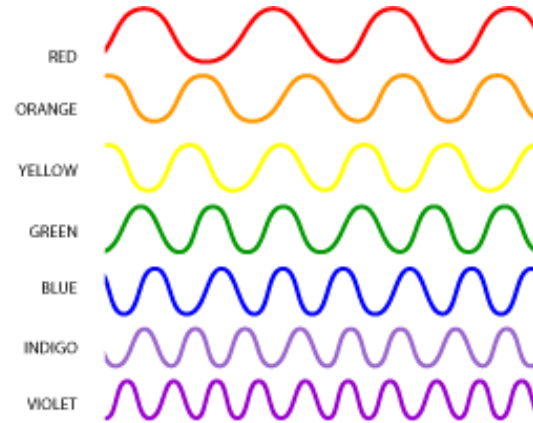
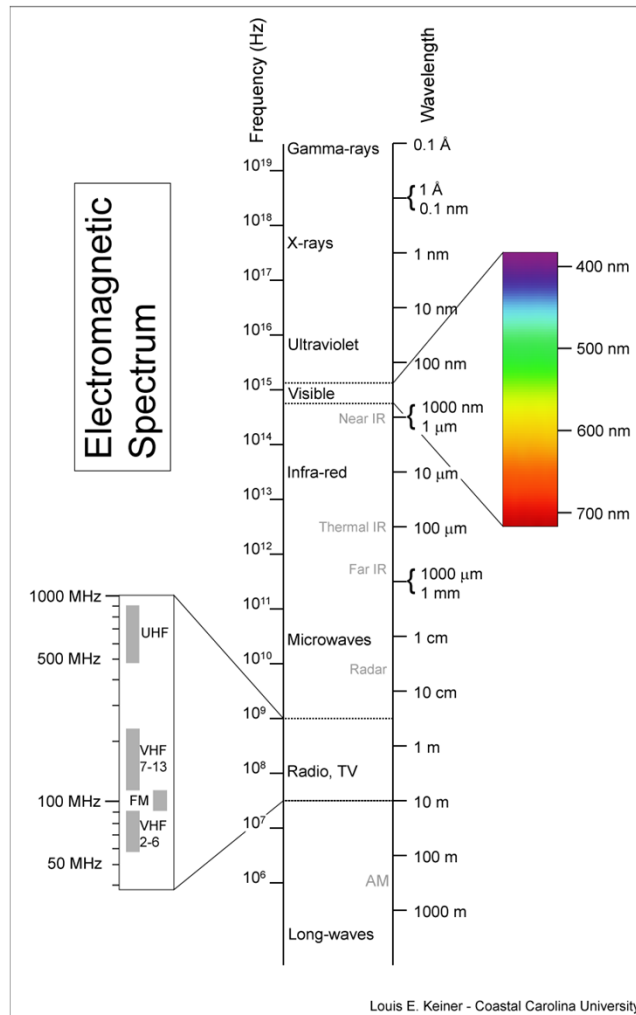
# Minimizing Distortion by Matching Refractive Indices



<http://www.microscopyu.com/articles/optics/waterimmersionobjectives.html>



# Spectrum of Visible Light



<u>Color</u>	<u>Wavelength</u>
<u>violet</u>	380–450 nm
<u>blue</u>	450–495 nm
<u>green</u>	495–570 nm
<u>yellow</u>	570–590 nm
<u>orange</u>	590–620 nm
<u>red</u>	620–750 nm

[http://en.wikipedia.org/wiki/Visible\\_spectrum](http://en.wikipedia.org/wiki/Visible_spectrum)

<http://science.hq.nasa.gov/kids/imagers/ems/visible.html>

# Photon Energy

- Energy of a photon: Planck's law

$$E = h\nu = h \frac{c}{\lambda}$$

$h$ : Planck's constant;  $6.626 \times 10^{-34} \text{ J}\cdot\text{s}$

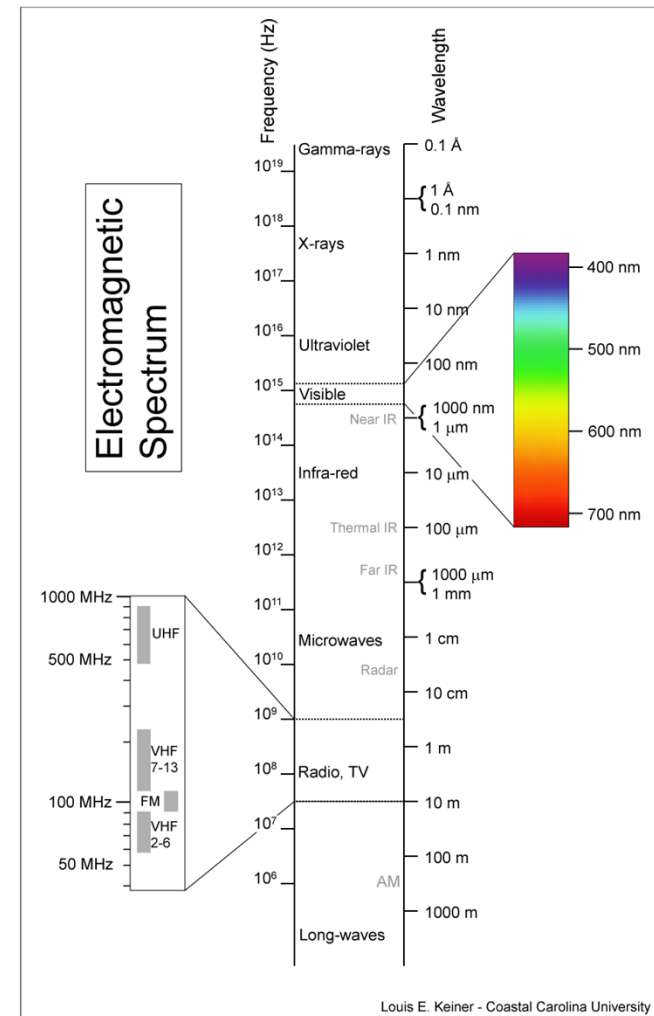
$\nu$ : frequency of light;

$\lambda$ : wavelength of light

$c$ : speed of light

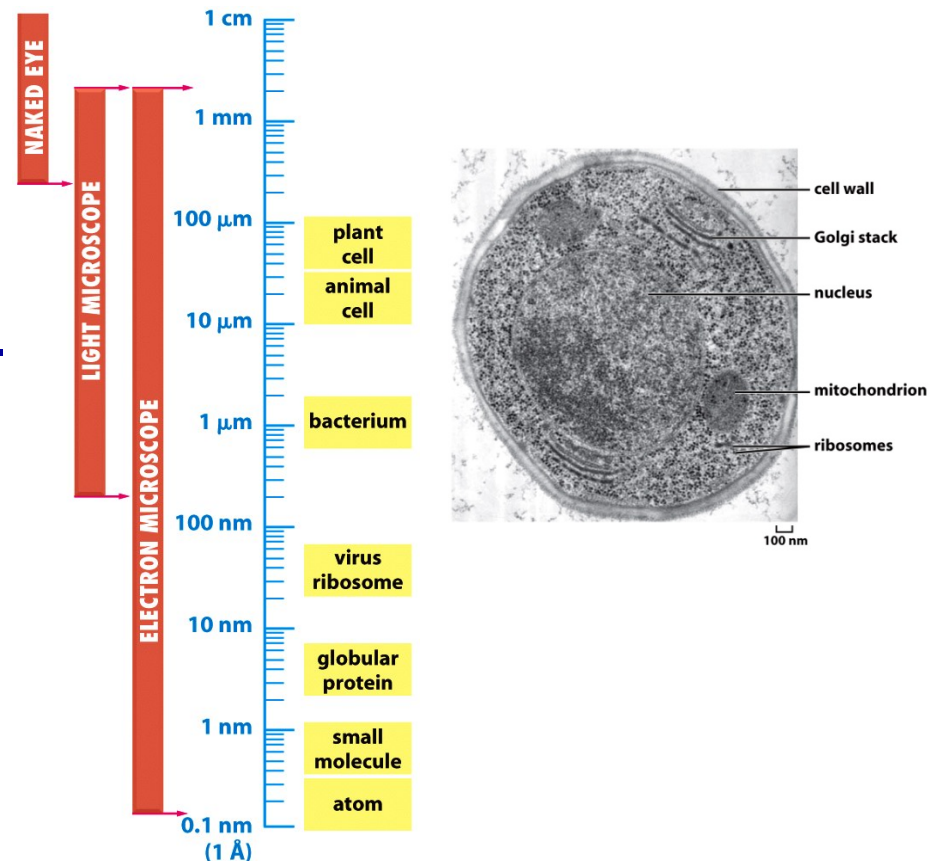
energy of a photon =  
 $3.973 \times 10^{-19} \text{ J}$  at 500nm

- Shorter waves have higher energy.



# Why Use Light Microscopy?

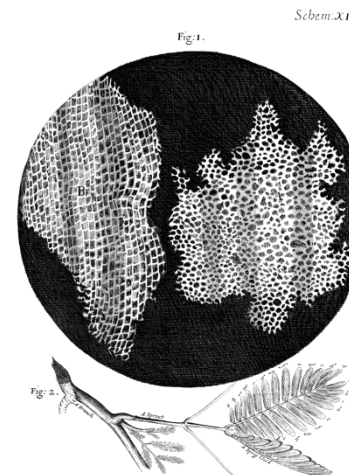
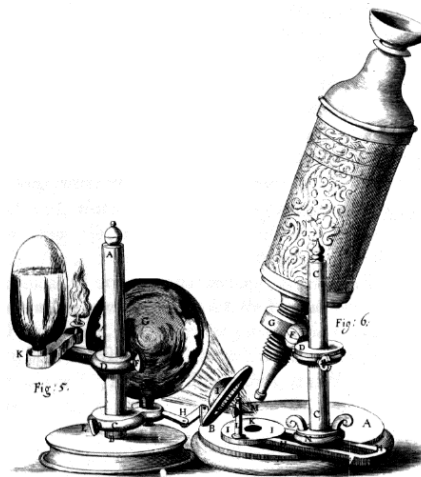
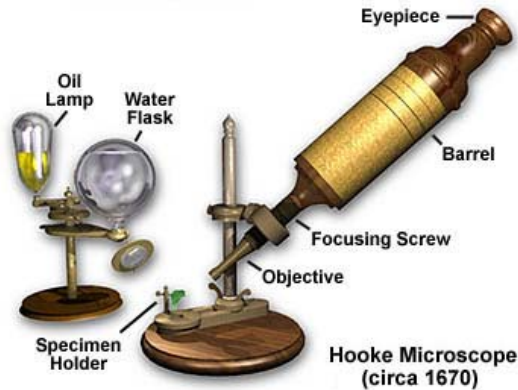
- Microscopy makes it possible to visualize cell structure and dynamics.
- Light microscopy permits live imaging of cellular processes.
- Electron microscopy provides higher resolution but requires samples to be fixed.



- 
- Importance of understanding and optimizing image formation
  - Some basic optics facts
  - **Light microscope structure**
  - Contrast generation in microscopy
  - Practical considerations of microscopy

# Origination of Light Microscopes

- Light microscope was invented more than three hundred years ago.  
(*Micrographia*, Robert Hooke, 1665)



<http://micro.magnet.fsu.edu/index.html>  
Molecular expressions: microscopy world

# Two Microscope Configurations

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Upright



Inverted

[http://www.olympusamerica.com/seg\\_section/seg\\_home.asp](http://www.olympusamerica.com/seg_section/seg_home.asp)

- Modern microscopes are computer-controlled .
- Modern microscopes can be configured to be highly automated.

# Some Reference Information

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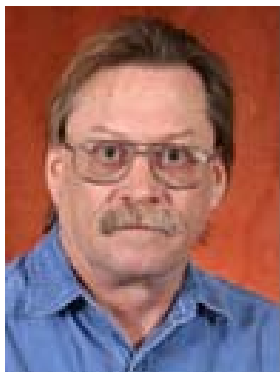
- Major microscope manufacturers

The Olympus logo consists of the word "OLYMPUS" in a bold, white, sans-serif font, centered within a dark blue rectangular background.

- Basic microscope structures and performance from different suppliers are very similar.

# Some Reference Information

---



Michael W. Davidson  
Florida State University



<http://micro.magnet.fsu.edu/index.html>  
Molecular expressions: microscopy world



<http://www.olympusmicro.com/>



<http://www.microscopyu.com/>



<http://zeiss-campus.magnet.fsu.edu/index.html>



# External Structure of Modern Microscopes

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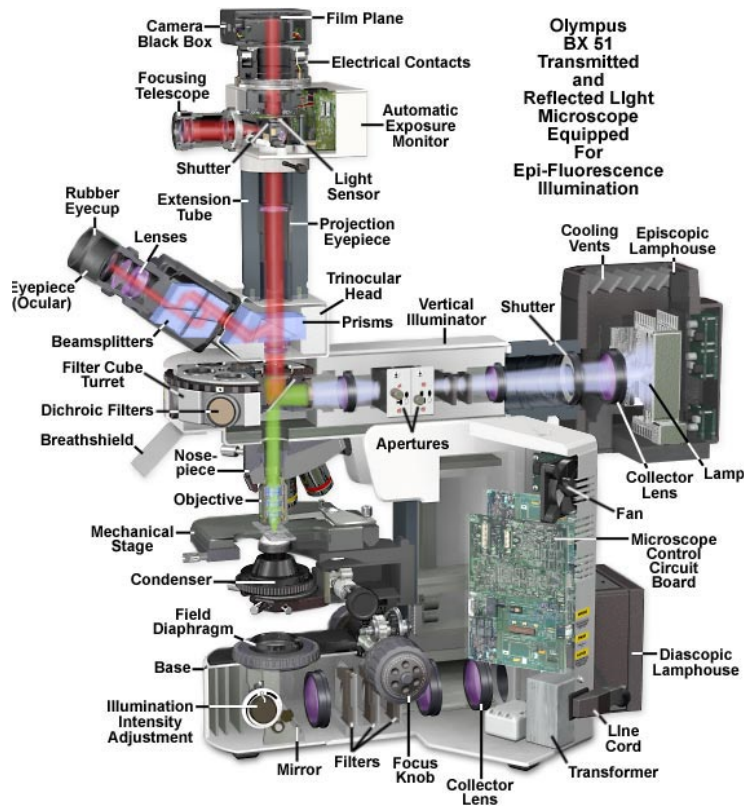


Upright



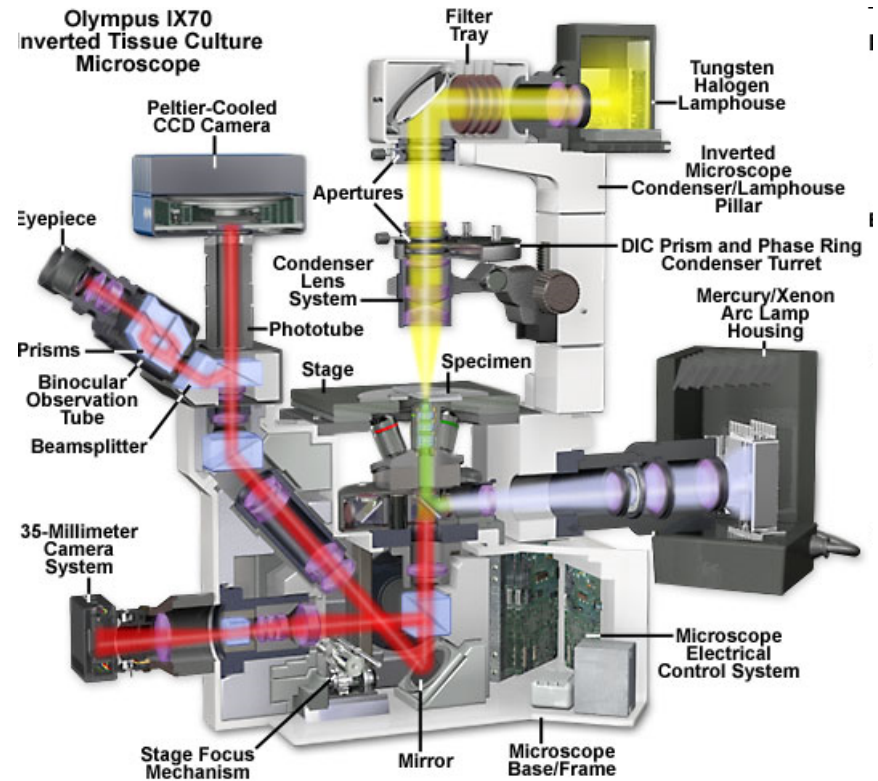
Inverted

# Internal Structure of Modern Microscopes



Upright

<http://www.olympusmicro.com/>

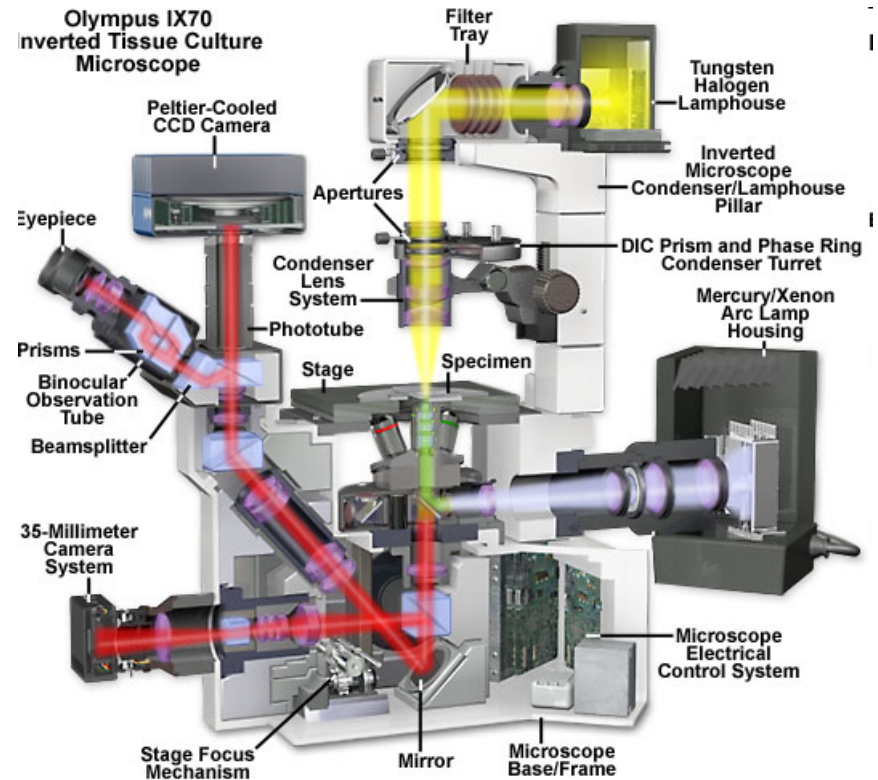
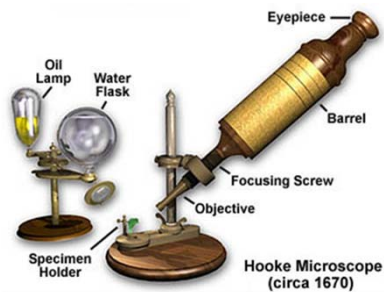


Inverted

Episcopic: reflected; Diascopic: transmitted

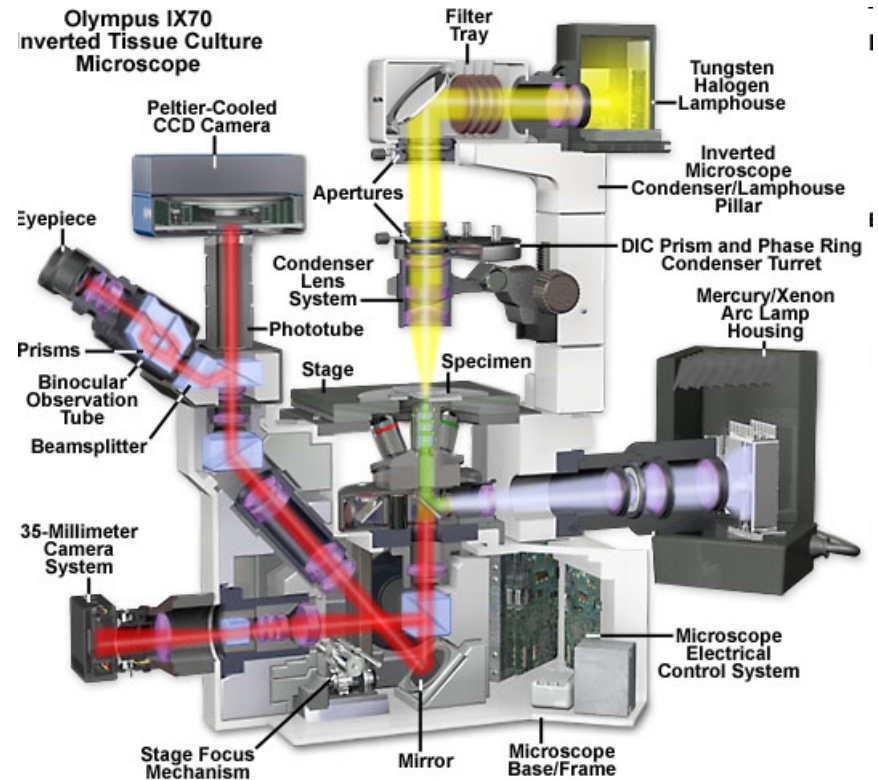
# Light Path Components (I)

- Illumination source
- Illumination filters
- Condenser
- Specimen stage



# Light Path Components (II)

- Objective lens
- Image filter
- Image sensor
- Eyepiece



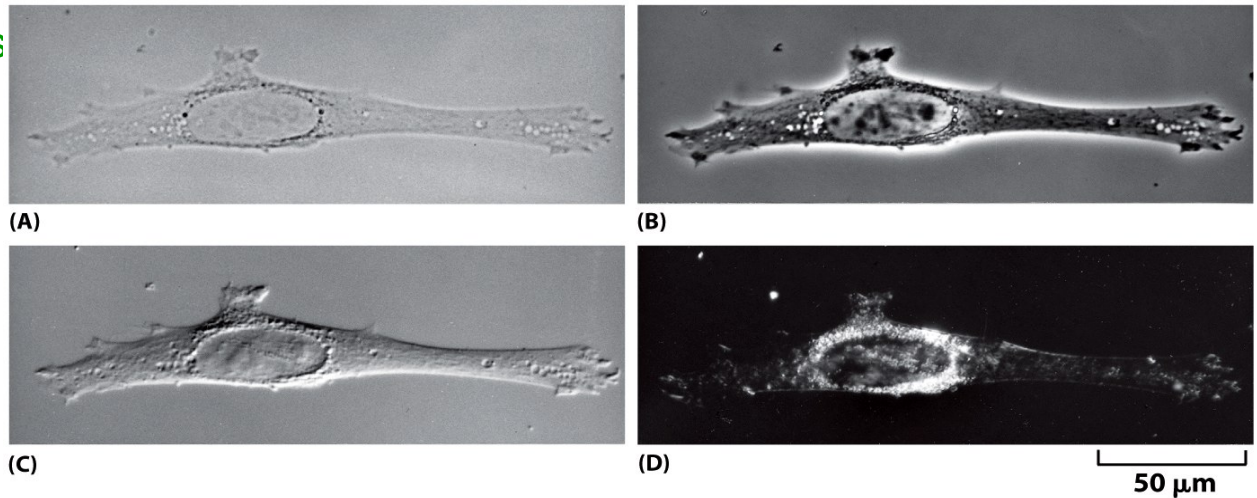
- 
- Importance of understanding and optimizing image formation
  - Some basic optics facts
  - Light microscope structure
  - **Contrast generation in microscopy**
  - Practical considerations of microscopy



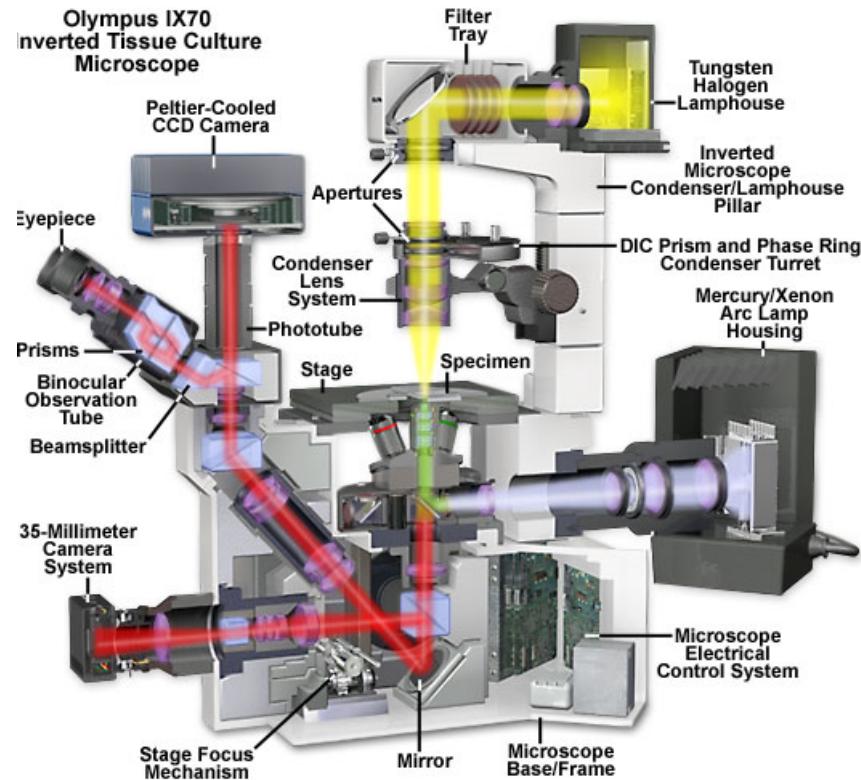
# Contrast Generation in Light Microscopy

- Two fundamental roles of any microscope
  - To provide adequate contrast
  - To provide adequate resolution.
- Contrast generation
  - Transmitted light illumination vs reflected light illumination
  - Bright-field vs dark-field
  - Phase contrast
  - Fluorescent micros

(A) Bright-field  
(B) Phase  
(C) DIC  
(D) Dark-field



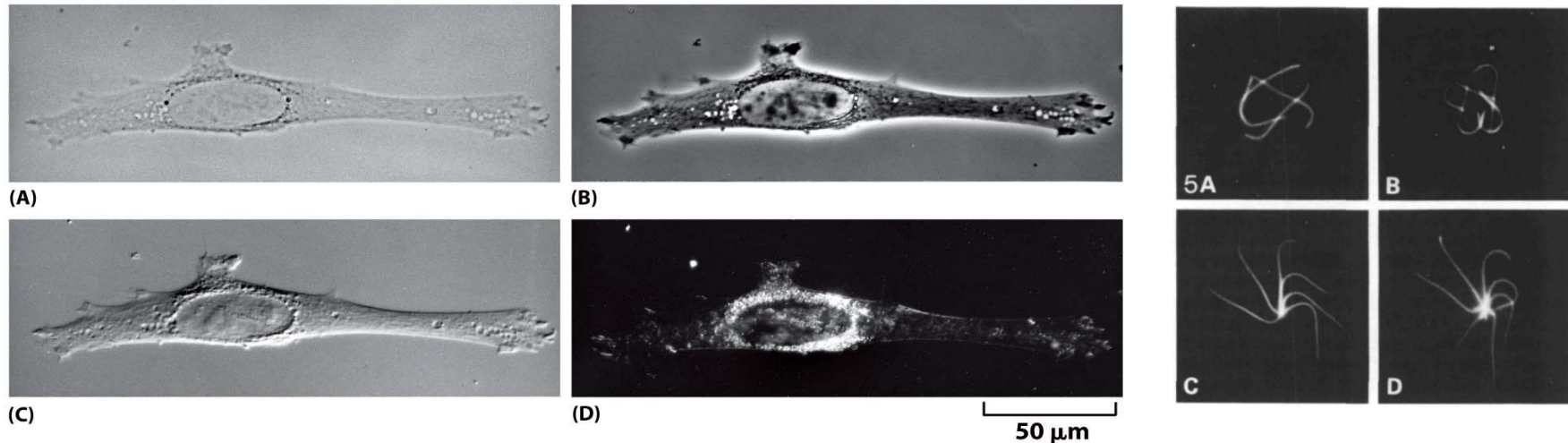
# Reflected Light vs Transmitted Light



<http://www.olympusmicro.com/primer/java/lightpaths/ix70fluorescence/ix70.html>

# Bright-field vs Dark-field (I)

- Under bright-field contrast, the specimen appears dark against a bright background.
- Dark-field contrast is particularly useful when imaging thin filaments or small particles.





# Bright-field vs Dark-field (II)

- Under dark-field contrast, by using a special condenser, only the light scattered by the specimen can enter the objective lens.

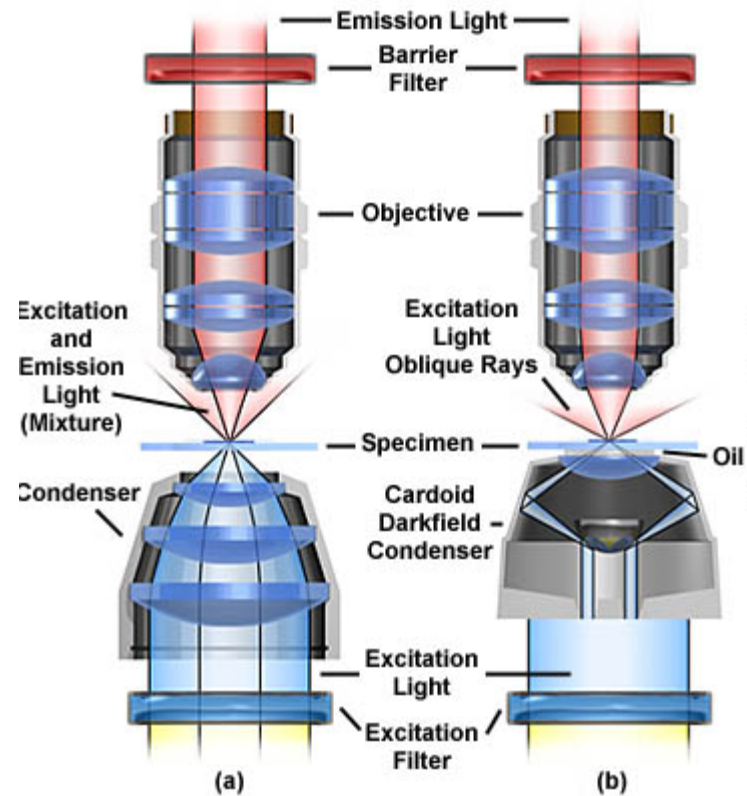
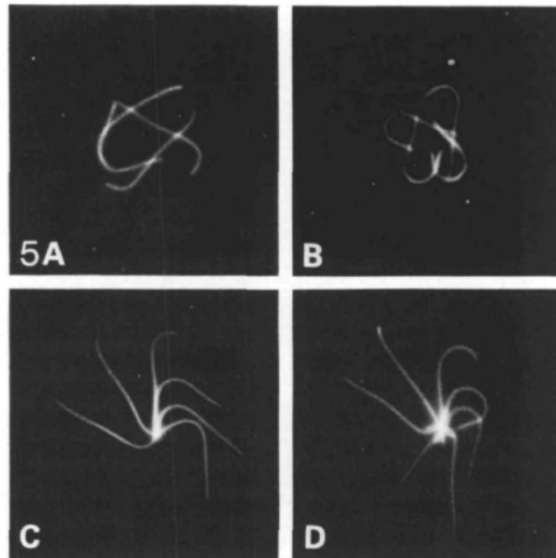
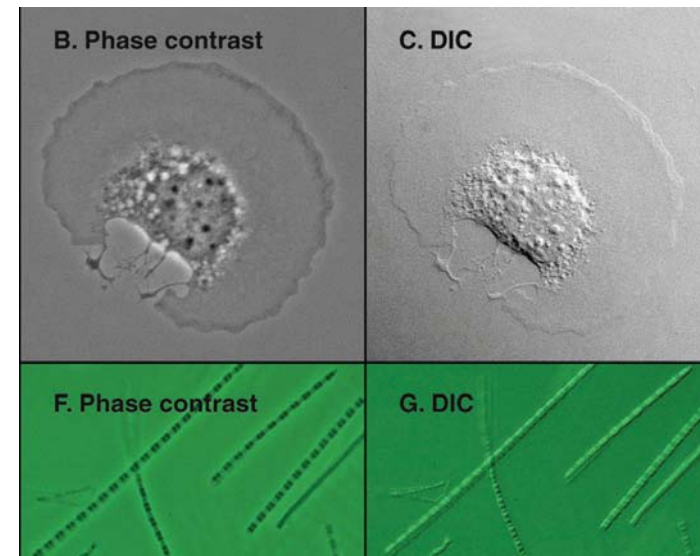
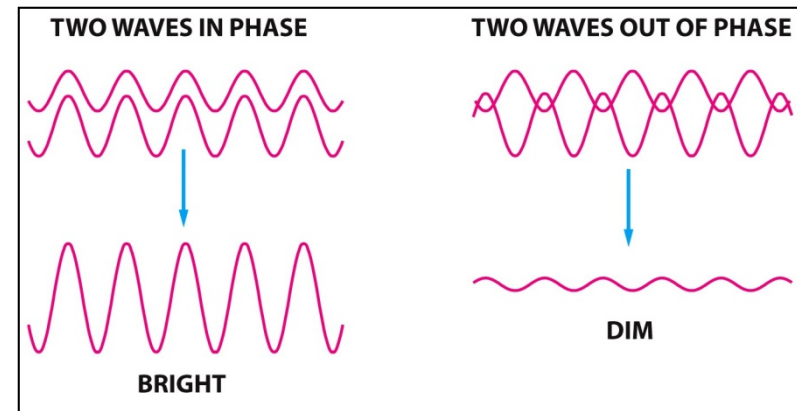


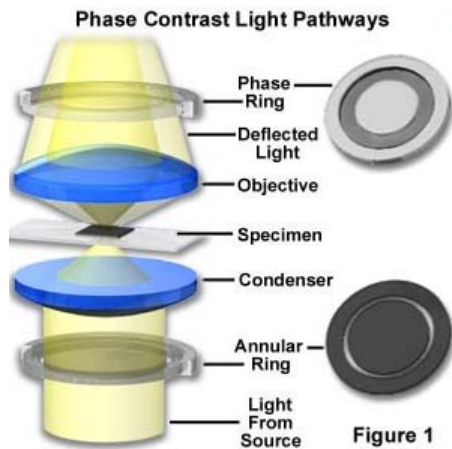
Figure 1

# Phase Contrast & DIC (I)

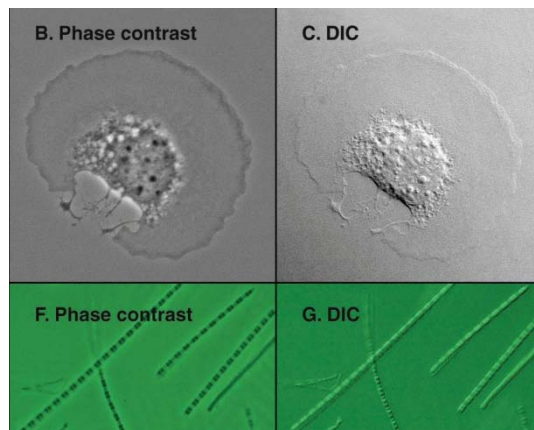
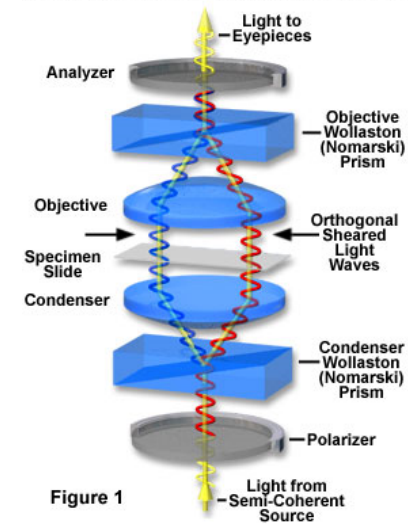
- Phase contrast is very useful in imaging transparent specimens, which do not change light magnitude.
- Contrast is generated due to the different refractive indices of the sample and the background.
- Phase contrast can generate artifacts.
  - Halos by boundary
  - Artificial shadows
- DIC significantly reduces halos and shadows.



# Phase Contrast & DIC (II)



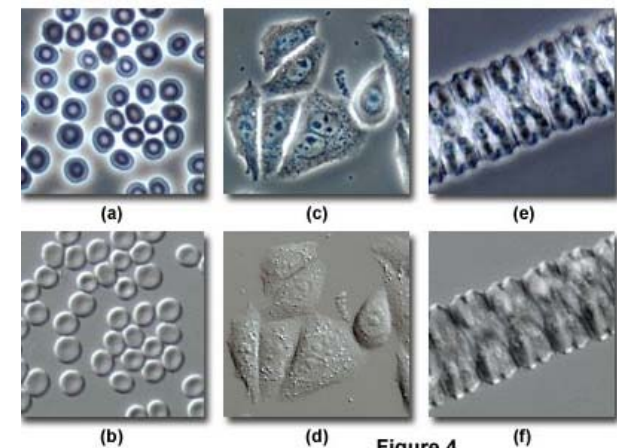
**Differential Interference Contrast Schematic**



Phase

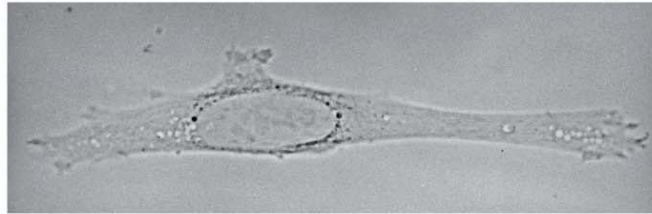
DIC

**Halos in Phase Contrast and DIC Microscopy**

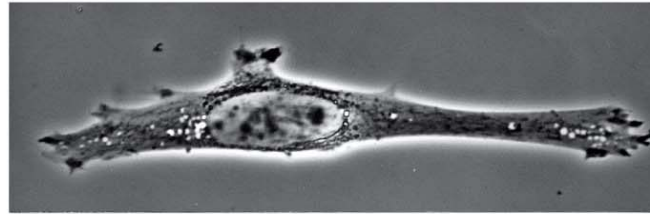


**Figure 4**

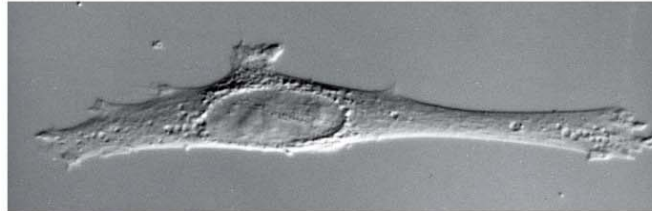
# What is wrong with ALL the cell images?



(A)



(B)

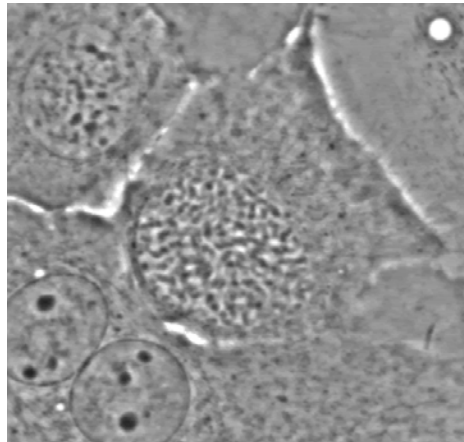


(C)



(D)

50  $\mu\text{m}$



The Nobel Prize in Physics 1953

"for his demonstration of the phase contrast method, especially for his invention of the phase contrast microscope"



Frits (Frederik)  
Zernike

the Netherlands

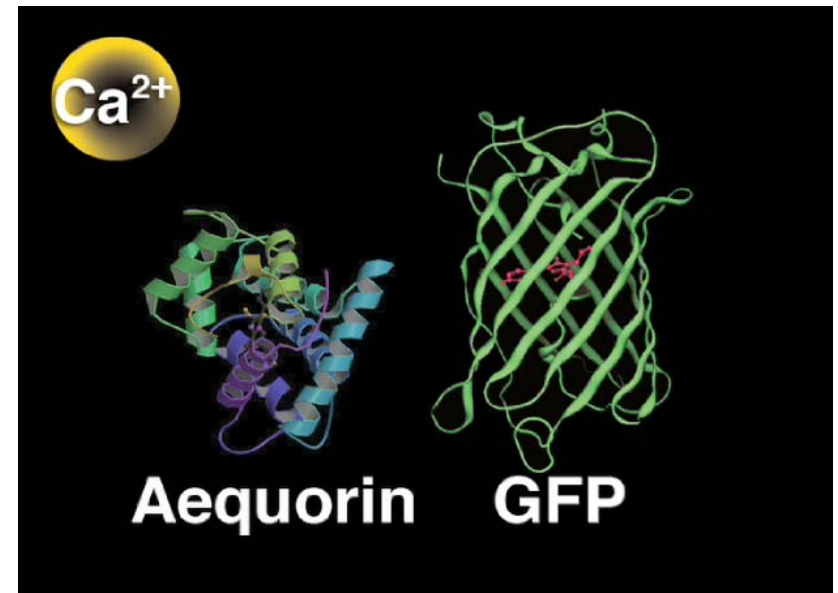
Groningen University  
Groningen, the  
Netherlands

b. 1888  
d. 1966

# Green Fluorescence Protein



Jellyfish: *Aequorea victoria*

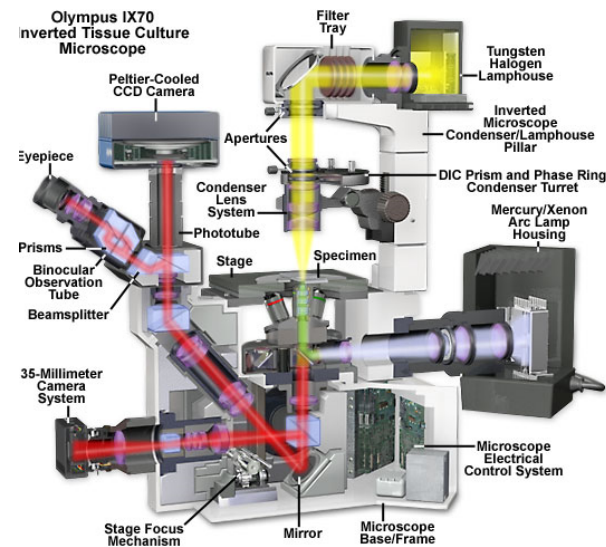
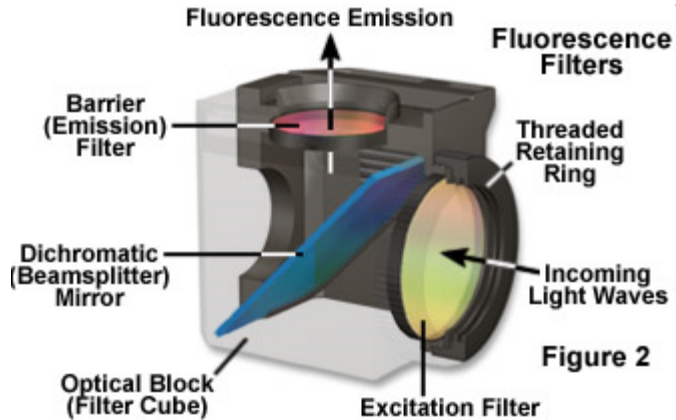
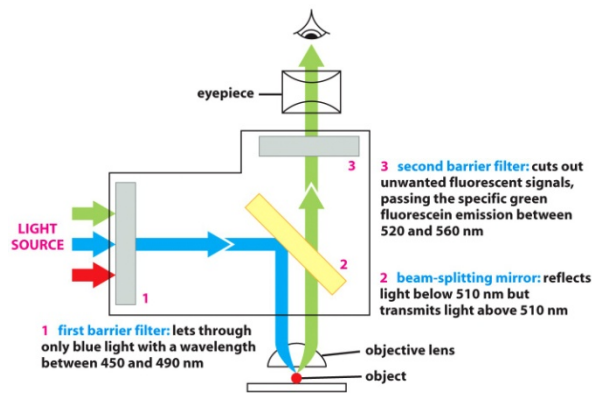
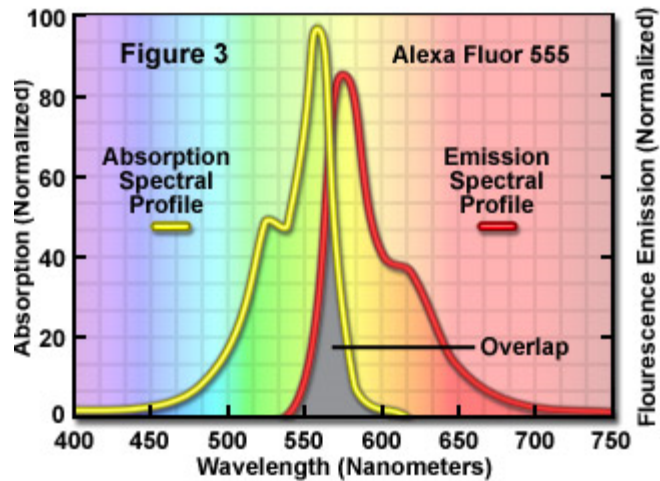


<http://gfp.conncoll.edu/GFP-1.htm>



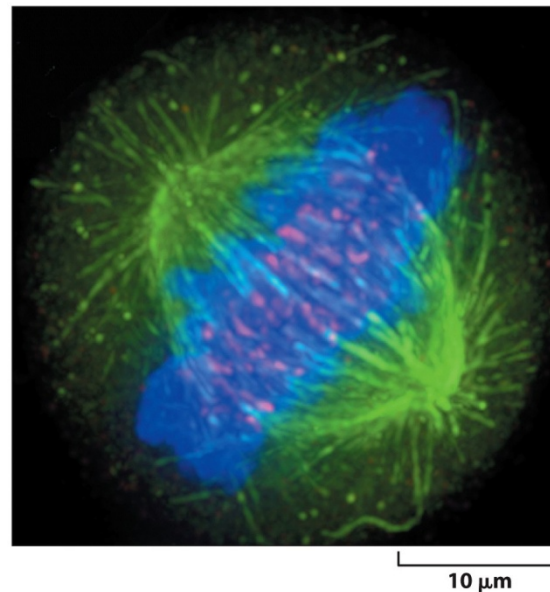
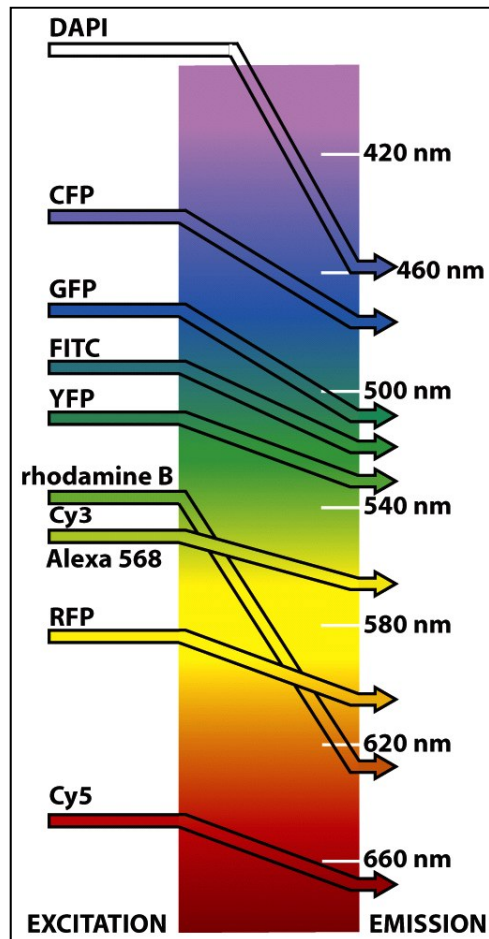
# Fluorescence Microscopy (I)

Fluorophore Absorption and Emission Profiles



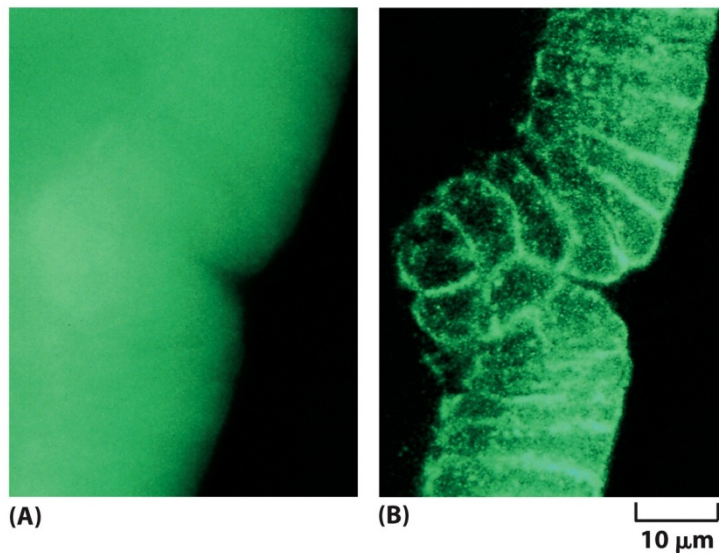
# Fluorescence Microscopy (II)

Fluorophores are available at many different colors



Blue: chromosome  
Green: microtubules  
Red: kinetochores

# Widefield vs Confocal Microscopy



Confocal and Widefield Fluorescence Microscopy

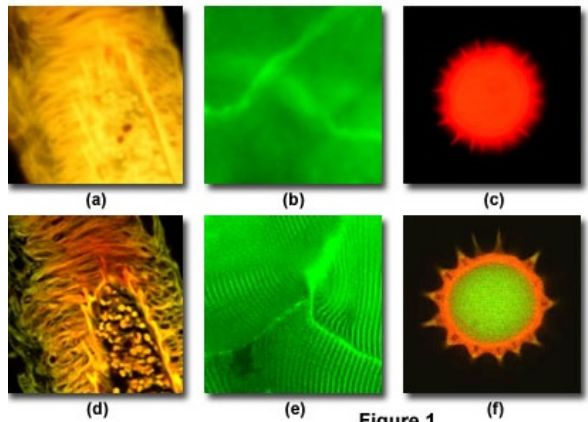


Figure 1

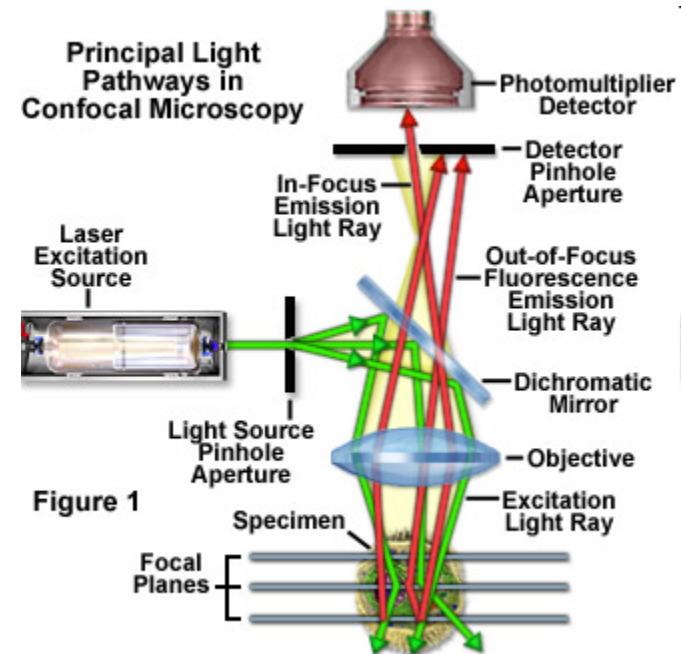


Figure 1

Widefield versus Confocal Point Scanning of Specimens

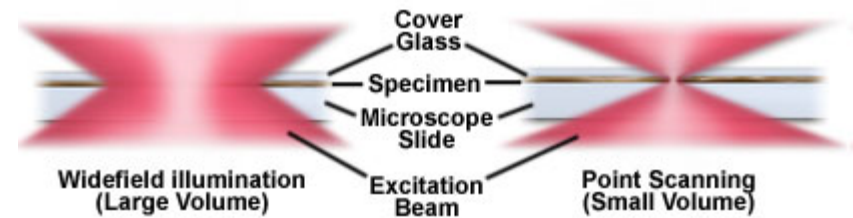


Figure 4

<http://www.olympusfluoview.com/theory/confocalintro.html>



# Total Internal Reflection Microscopy (I)

- Typical thickness of the evanescent layer is less than 200nm
- Often used for imaging
  - membrane related cellular processes
  - single molecules

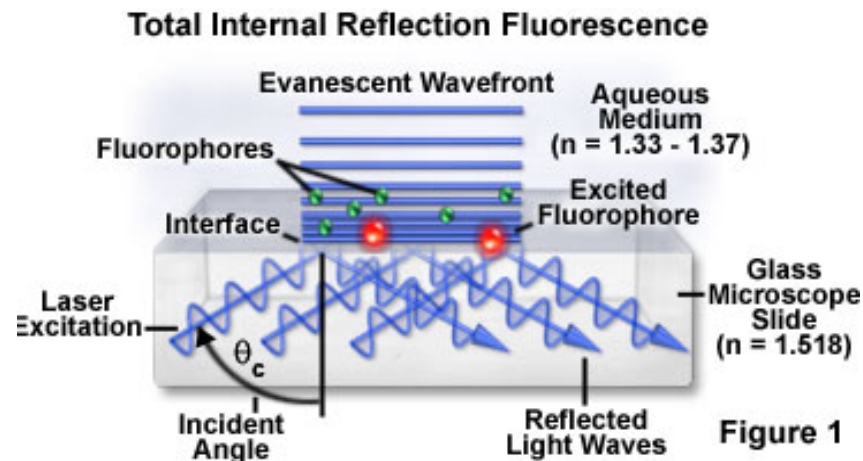
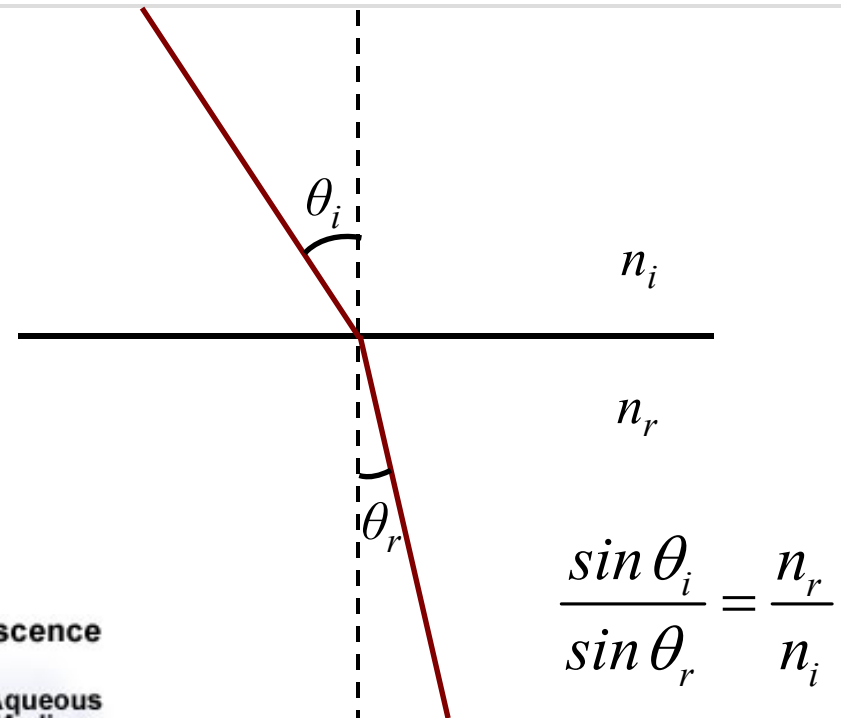
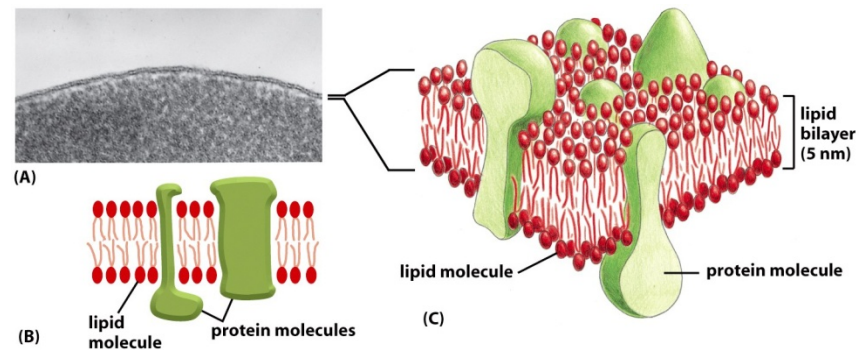
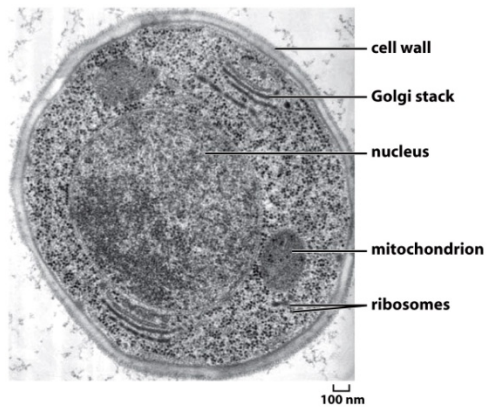


Figure 1

# Total Internal Reflection Microscopy (II)

- TIRF is often used for imaging
  - membrane related cellular processes
  - single molecules



# Fluorescence Microscopy Summary

---

- High specificity:
  - Chemical fluorophores
  - Fluorescent proteins
- High sensitivity: up to single molecules.

- 
- Importance of understanding and optimizing image formation
  - Some basic optics facts
  - Light microscope structure
  - Contrast generation in microscopy
  - **Practical considerations of microscopy**

# Practical Considerations

---

- Photobleaching

- Fluorophores gradually lose their ability of light emission.
- This results in a sustained decrease in image intensity.

- Phototoxicity

- Constant illumination generates free radicals that cause cell death.
- This places a fundamental limit on how many frame of images can be collected.

---

**Questions?**