

BME 42-620 Engineering Molecular Cell Biology

Lecture 10:

The Cytoskeleton (II): Microtubule & Intermediate Filament

The Cytoskeleton (III): Molecular Motors

# Outline

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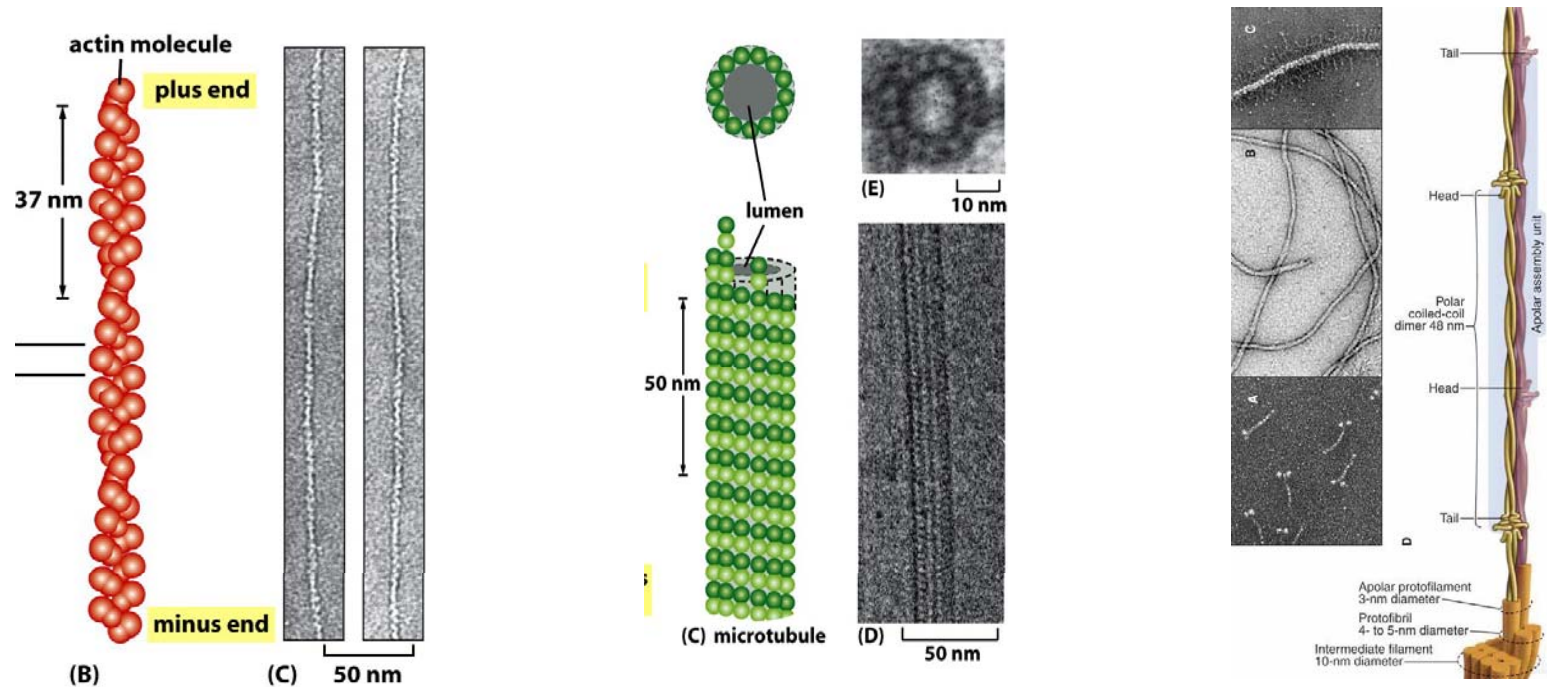
- Summary: actin and its associated proteins
- Microtubule and its associated proteins; Centrosome
- Intermediate filament and its associated proteins
- An overview of molecular motors

# Outline

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- Summary: actin and its associated proteins
- Microtubule and its associated proteins; Centrosome
- Intermediate filament and its associated proteins
- An overview of molecular motors

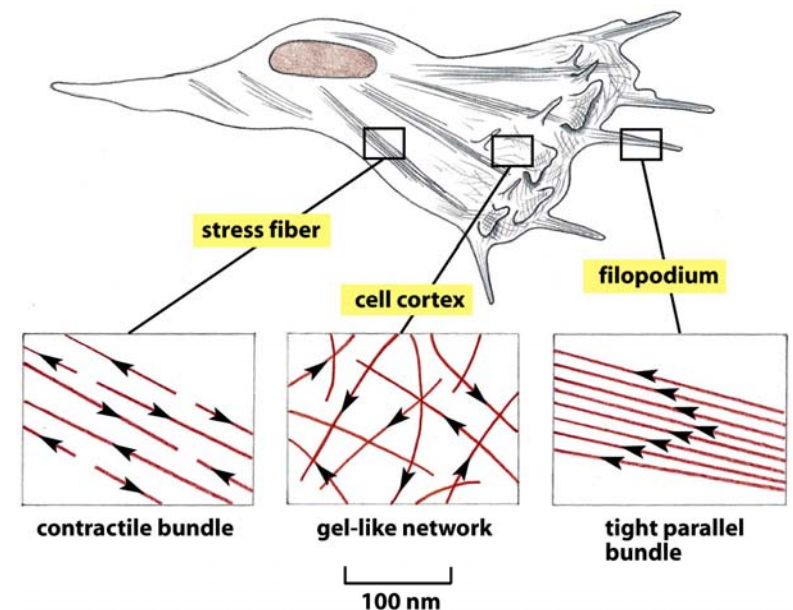
# Overview of Cytoskeletal Filaments



|                       | Shape | Diameter | Subunits            | Polarized |
|-----------------------|-------|----------|---------------------|-----------|
| actin                 | cable | ~6 nm    | actin monomer       | yes       |
| microtubule           | tube  | ~25nm    | tubulin heterodimer | yes       |
| intermediate filament | rope  | ~10nm    | Various dimers      | no        |

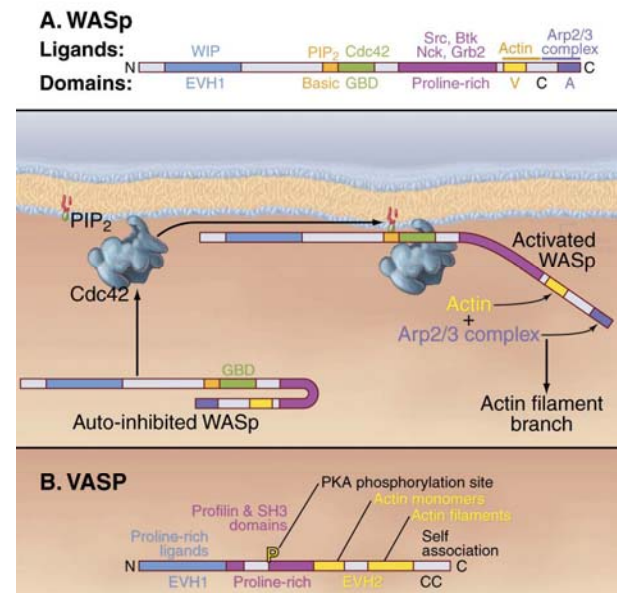
# Summary: Actin and its Associated Proteins (I)

- Actin is relatively soft (we will study related quantification in later lectures).
- Actin often form bundles; their mechanical strength comes mostly from bundling and crosslinking.
- Actin function mostly to withstand tension rather than compression.
- Actin is relatively stable and easy to work with biochemically.



# Summary: Actin and its Associated Proteins (II)

- Different actin associated proteins serve a broad range of functions.
- These proteins generally have multiple functional domains serving multiple functions.
- Some but not all of them are essential.
- Most of the proteins have functional overlap.
- Mathematical models are required to understand complex interactions between these proteins.



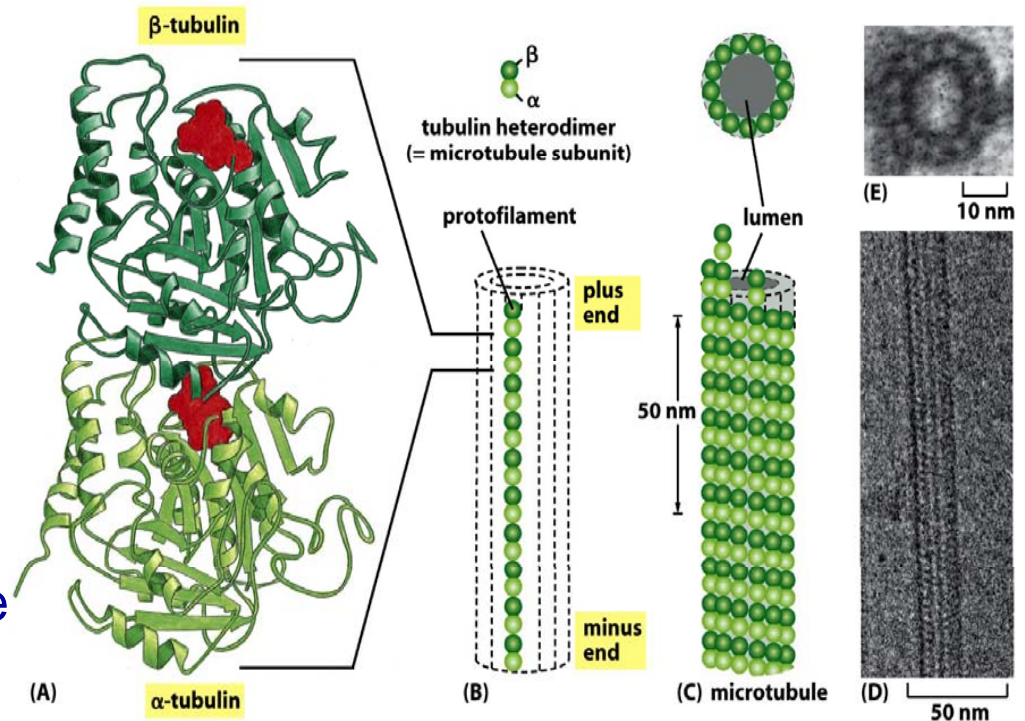
# Outline

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- Summary: actin and its associated proteins
- **Microtubule and its associated proteins; Centrosome**
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# Overview of Microtubule Structure

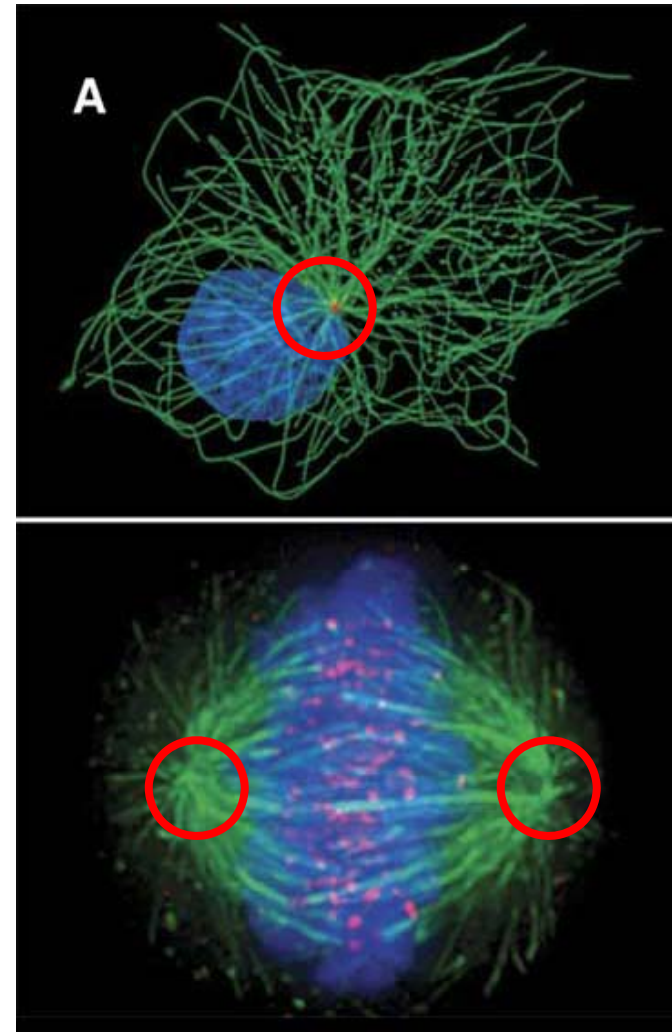
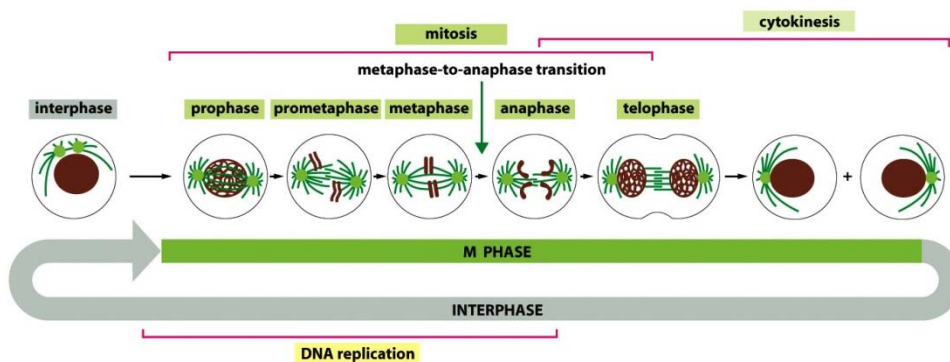
- Microtubule is polarized.
  - plus ( $\beta$ -tubulin) end
  - minus ( $\alpha$ -tubulin) end
- The outer diameter of a microtubule is ~25 nm.
- A microtubule typically has 13 protofilaments; Some may have 11, 15, or even 16.
- The GTP bound to the  $\alpha$ -tubulin monomer does not hydrolyze.





# Microtubule Organization at Different Stages

- Microtubule in interphase
  - Organized into a radial pattern centered at the centrosome
- Microtubule in metaphase
  - Organized into a bipolar architecture

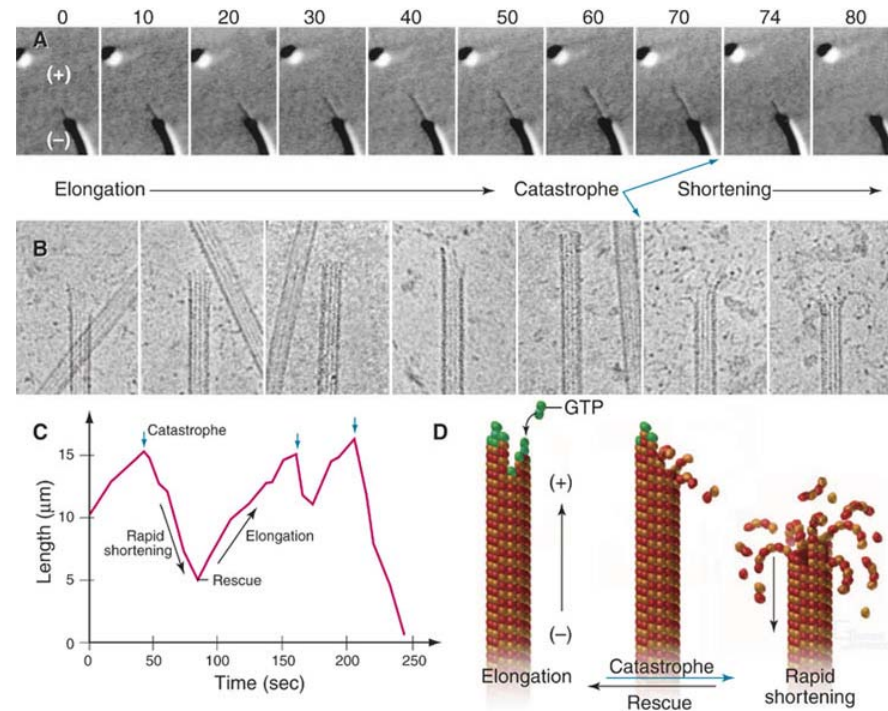


# Microtubule Dynamic Instability (I)

## Video 1 (Figure 1A)

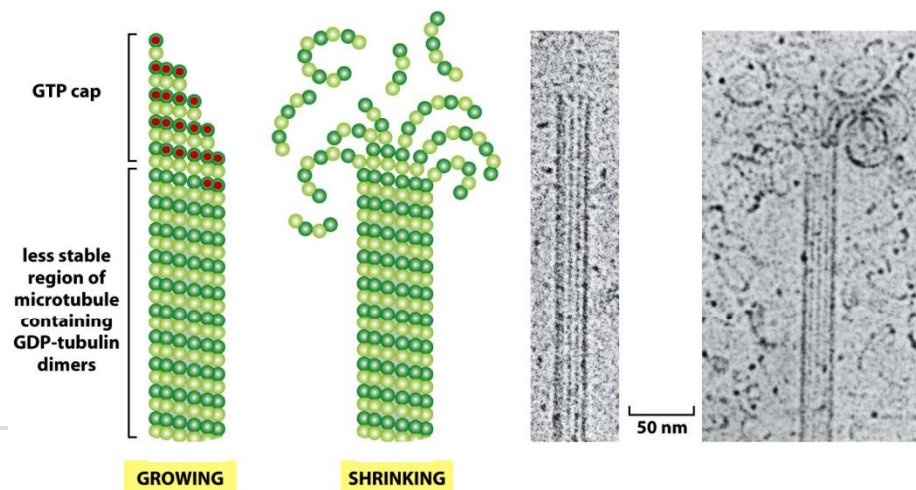
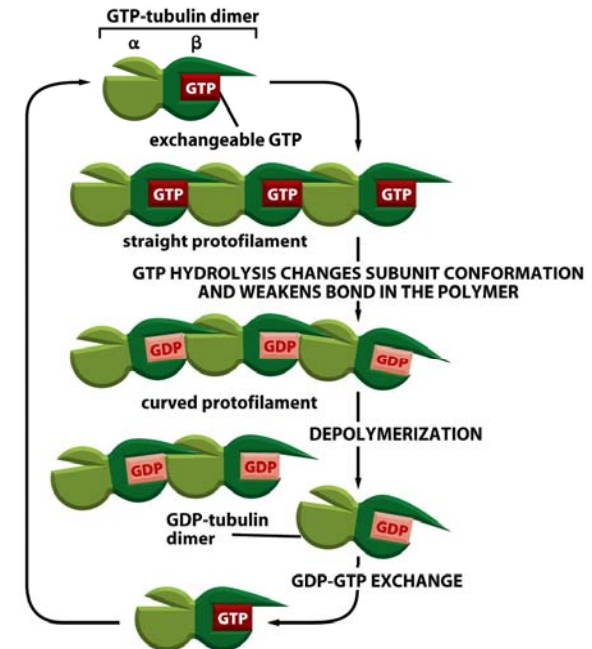
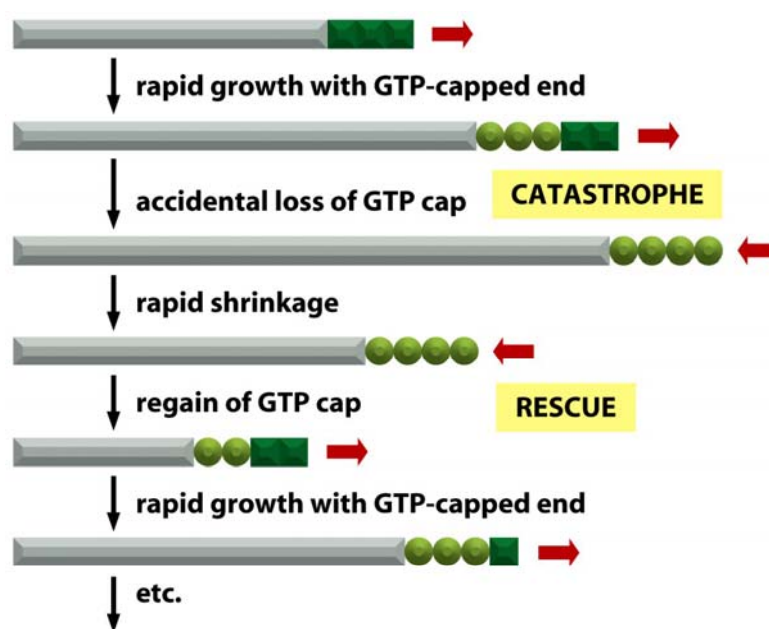
Microtubules in a PtK1 cell at the edge of an epithelial cell island.  
Few microtubules rapidly grow into nascent protrusions.

Elapsed time: 9 min 05 sec

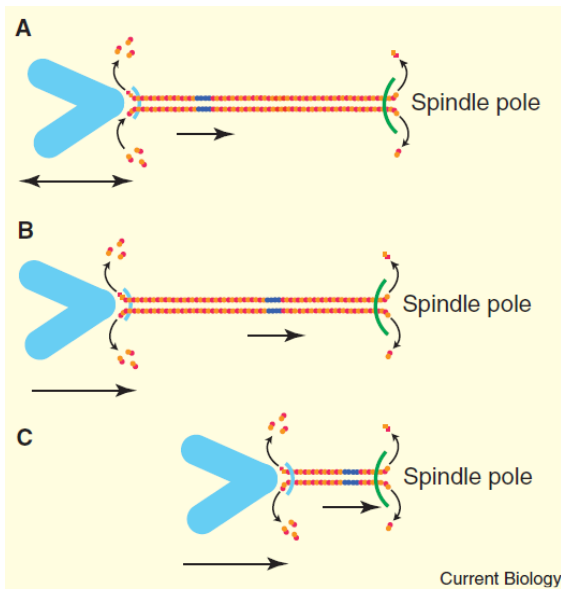


T. Wittmann et al, *J. Cell Biol.*, 161:845, 2003.

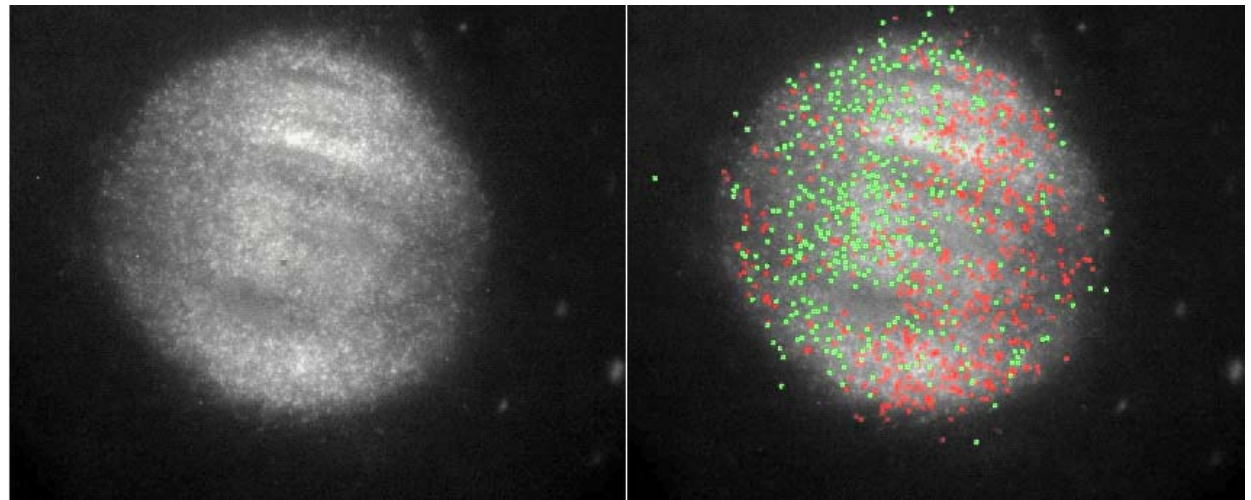
# Microtubule Dynamic Instability (II)



# Microtubule Treadmilling vs Dynamic Instability



Khodjakov & Kapoor, *Current Biology*,  
15:R966-R968, 2005.



# Representative Microtubule Dynamic Instability Parameters

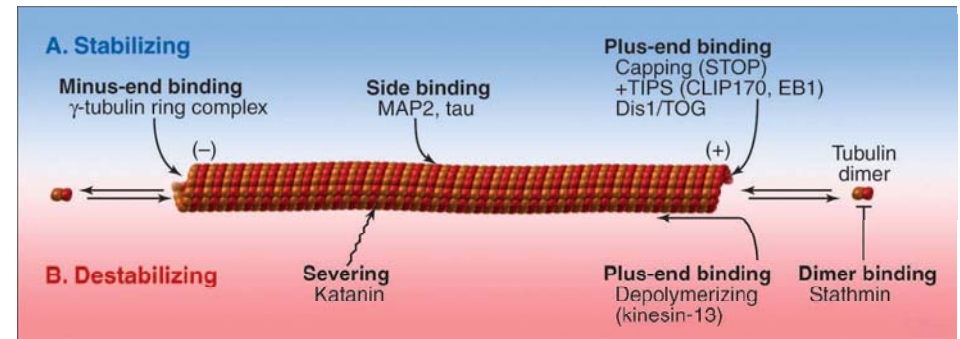
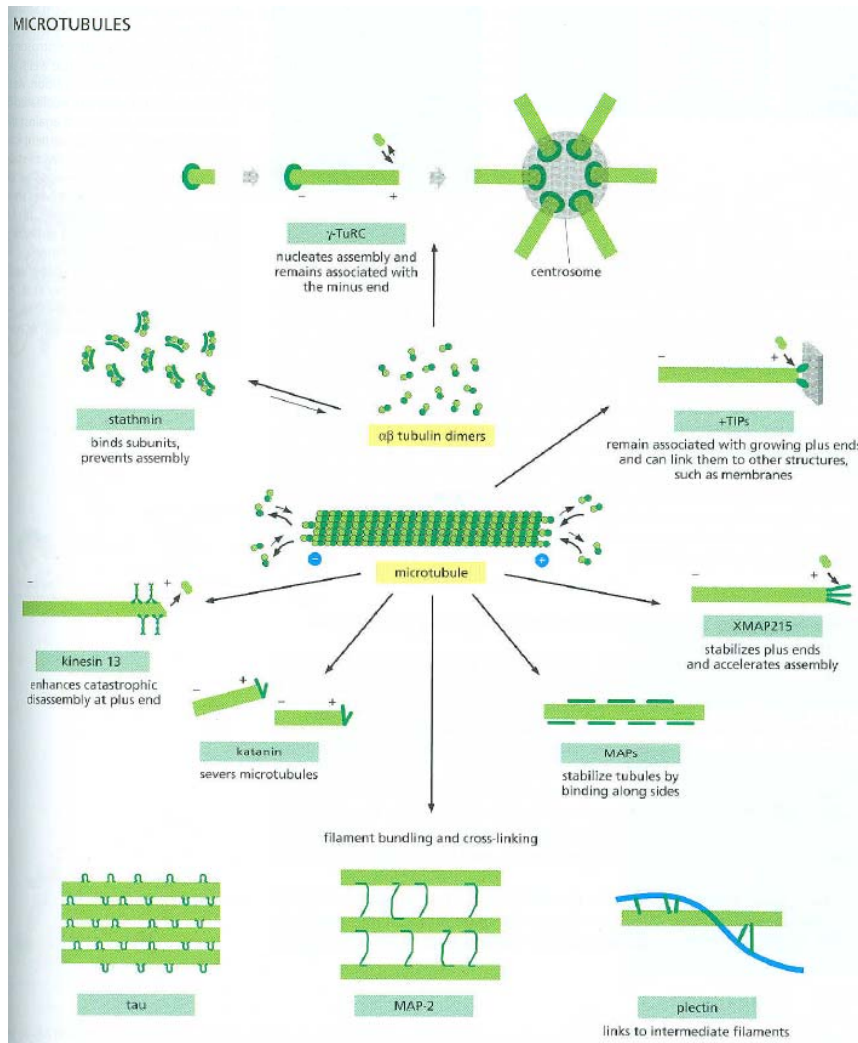
**Table 1.** Comparison of the parameters of microtubule dynamics in vitro and in vivo. Dashes indicate that no microtubules were formed. NA, not applicable.

| Parameter  | In vitro<br>(25 $\mu$ M tubulin) |                         |                        |  | In vivo                        |          |                     |                             |         |
|--|----------------------------------|-------------------------|------------------------|--|--------------------------------|----------|---------------------|-----------------------------|---------|
|  |                                  |                         |                        |  | <i>Xenopus</i><br>egg extracts |          | Newt<br>lung cells‡ | Mammalian<br>LLCPK-1 cells§ |         |
|  | Tubulin<br>alone                 | +0.8 $\mu$ M<br>XMAP215 | + 0.2 $\mu$ M<br>XKCM1 | + 0.8 $\mu$ M<br>XMAP215<br>+ 0.2 $\mu$ M<br>XKCM1 | Interphase*                    | Mitotic† |                     | Interphase                  | Mitotic |
| Growth ( $V_g$ )<br>( $\mu$ m/min)   | 2.56<br>( $\pm 0.75$ )           | 6.76<br>( $\pm 1.76$ )  | —                      | 8.73<br>( $\pm 3.72$ )                             | 7.10                           | 11.40    | 7.20                | 11.50                       | 12.80   |
| Shrinkage ( $V_s$ )<br>( $\mu$ m/min)  | NA                               | NA                      | —                      | 19.94<br>( $\pm 5.20$ )                            | 9.40                           | 13.50    | 17.30               | 13.10                       | 14.10   |
| Catastrophe ( $F_{cat}$ )<br>(events/min)  | 0.04                             | 0                       | —                      | 1.06   | 0.69                           | 2.44     | 0.84                | 1.56                        | 3.48    |
| Rescue ( $F_{res}$ )<br>(events/min)   | NA                               | NA                      | —                      | 1.30   | 1.08                           | 0.70     | 2.64                | 10.50                       | 2.70    |
| *Verde <i>et al.</i> (9).   †Tournebise <i>et al.</i> (10).   ‡Cassimeris <i>et al.</i> (5).   §Rusan <i>et al.</i> (7). |                                  |                         |                        |  |                                |          |                     |                             |         |

Kinoshita *et al.*, *Science*, 294:1340, 2001



# Overview of Microtubule Associated Proteins



# Microtubule Associated Proteins (I)

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- **Stabilizing MAPs**

Tau Binds side and stabilizes microtubules

MAP2, MAP4 Binds side and stabilizes microtubules

- **Destabilizing MAPs**

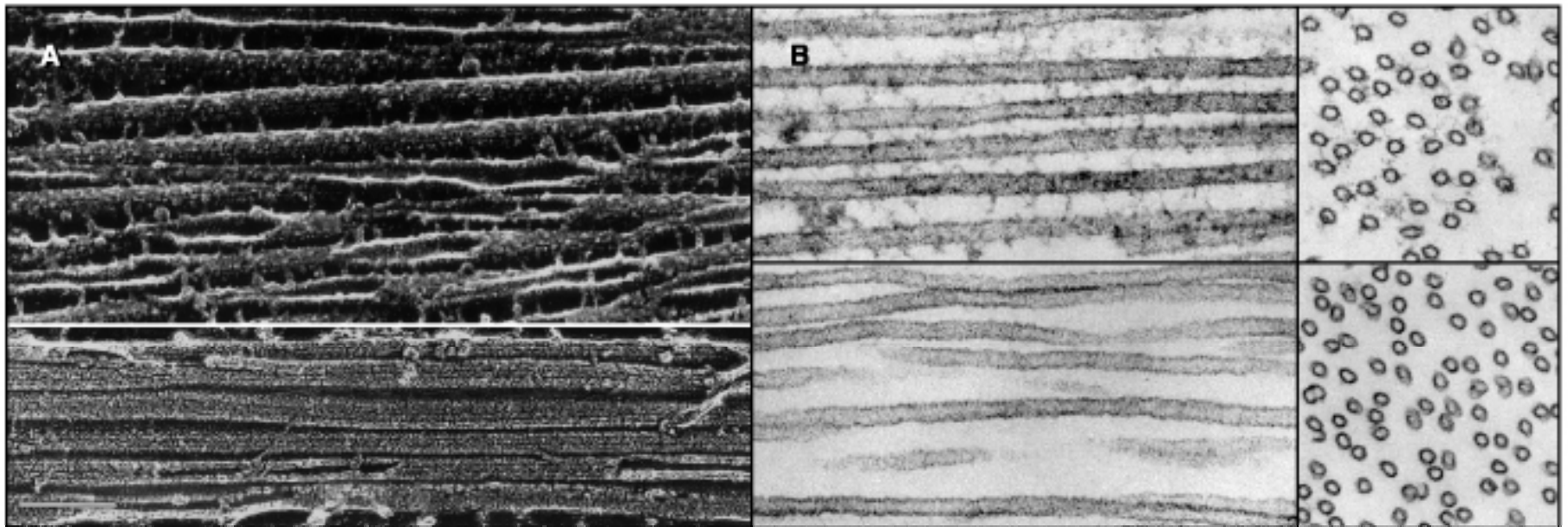
Op18/stathmin Binds tubulin dimers & destabilizes MTs

Katanin AAA ATPase that severs microtubules

XKCM/MCAK Kinesin-related; destabilizes plus ends

# Microtubule Associated Proteins (II)

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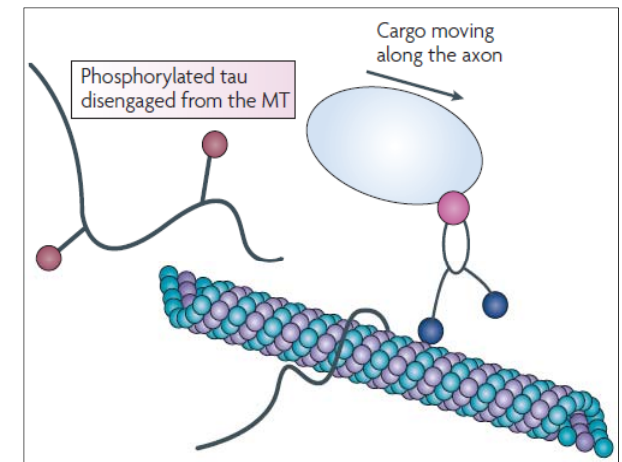
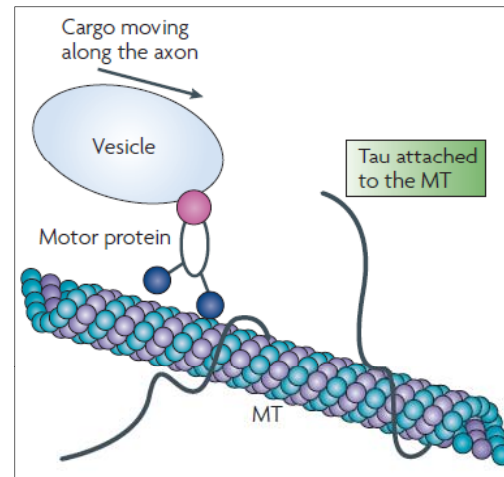
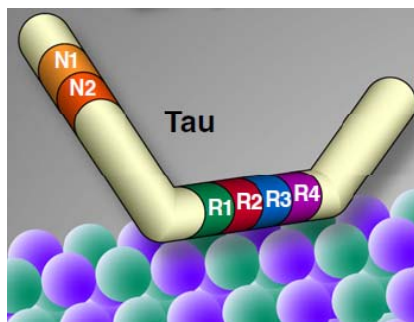
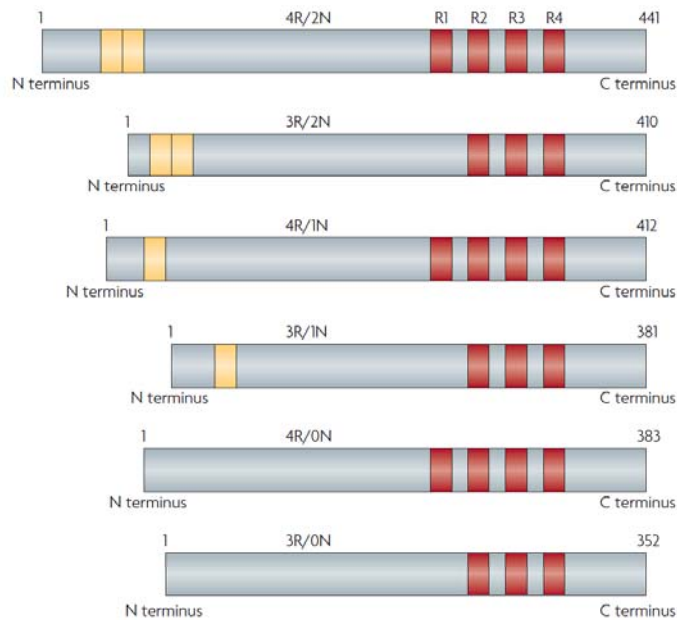
Frozen deep-etched, shadowed microtubules with (upper) and without (lower) tau

Fixed, embedded sections of microtubules with (upper) and without (lower) MAP2

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# Microtubule Associated Proteins (III): Tau

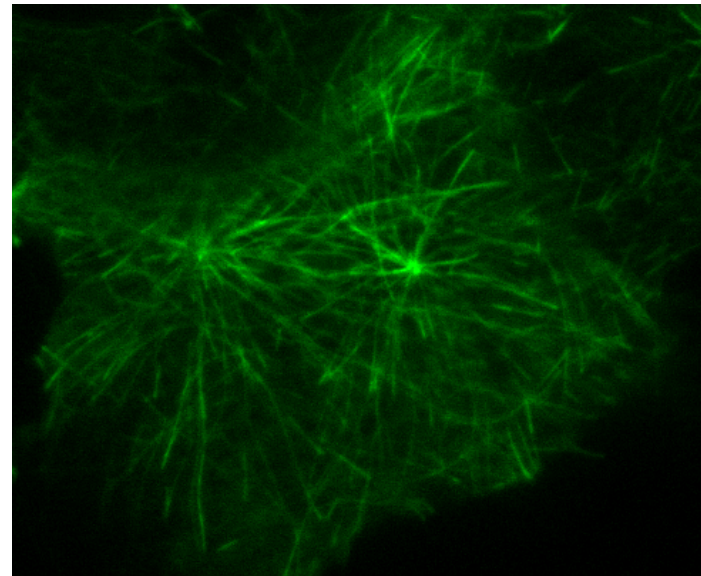
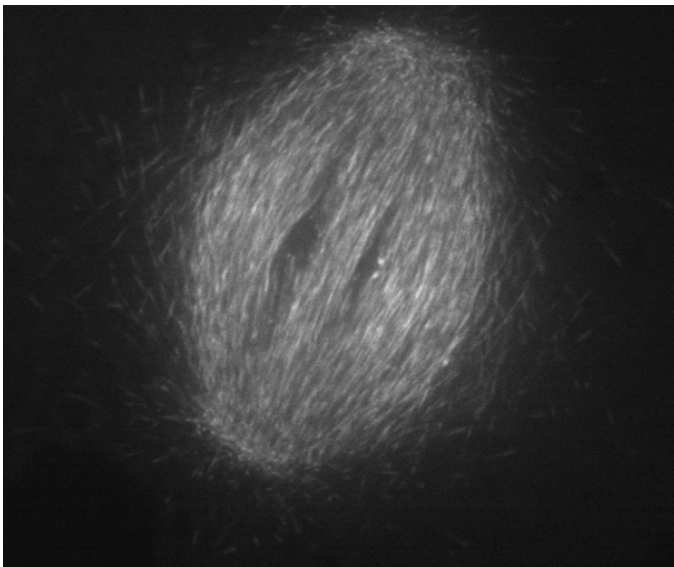


Ballatore et al, *Nat. Rev. Neurosci.* 2007  
Morris et al, *Neuron*, 2011

# Microtubule Associated Proteins (IV)

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- CLIP170 is the first microtubule plus end tracking protein identified.
  - Links membranes to growing plus ends.
  - Binds the microtubule plus end to reduce catastrophe.
- EB1 is another microtubule plus end tracking protein identified.
  - Binds the microtubule plus end to reduce catastrophe.



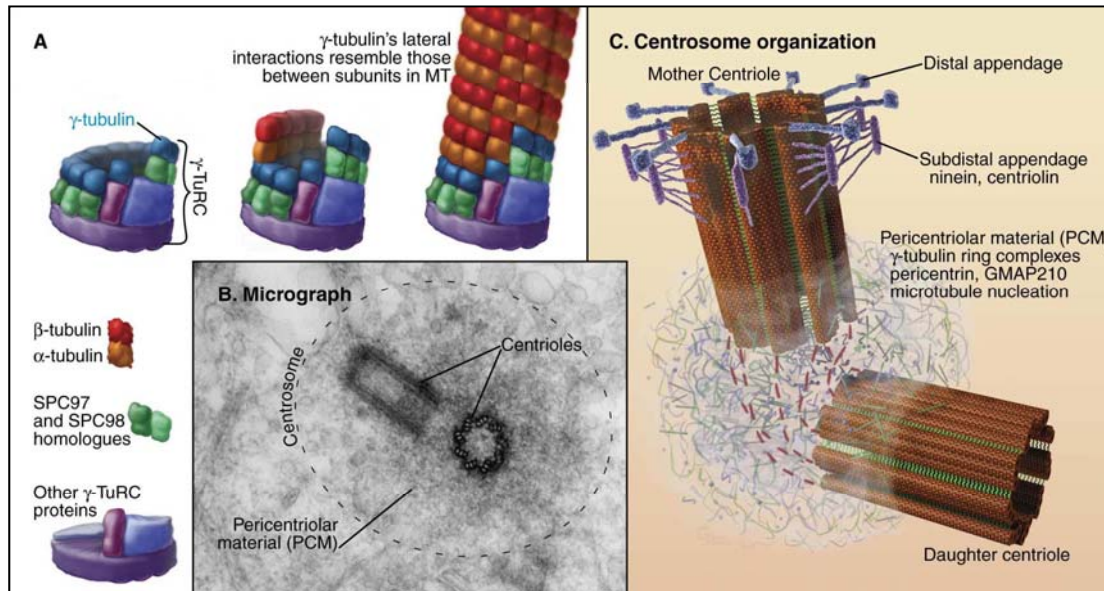
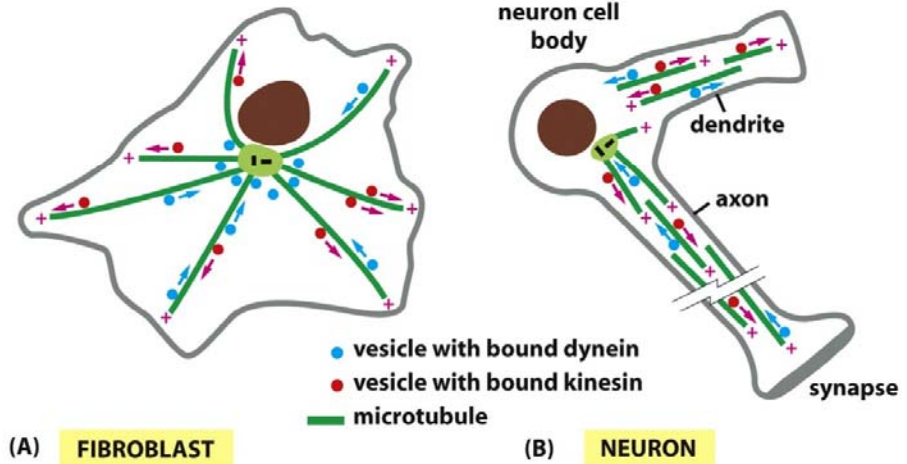
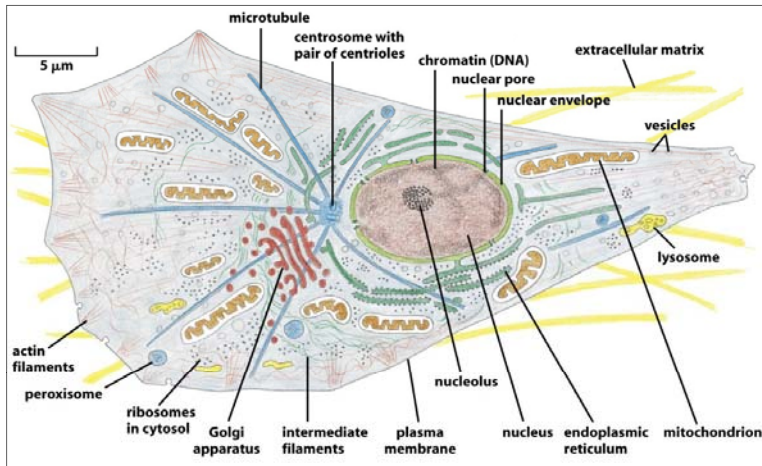
Dynamics and mechanics of the microtubule plus end. Howard J, Hyman AA. *Nature*. 2003 422(6933):753-8.

# Summary: Microtubule

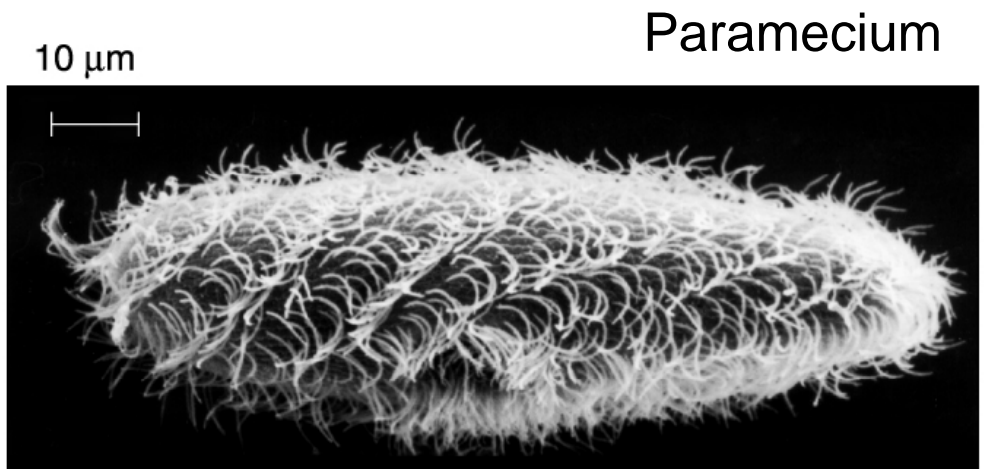
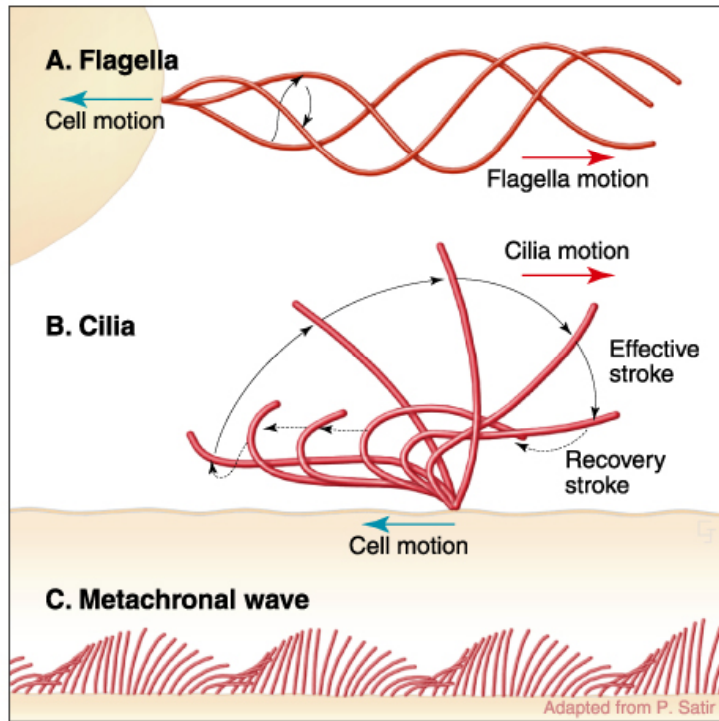
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- Microtubule is relatively rigid (quantification in following lectures).
- Microtubule often forms bundles. Mechanical strength of microtubule networks comes mostly from bundling and crosslinking.
- Microtubule can withstand compression.
- Microtubule can denature easily and is therefore difficult to work with biochemically.

# Centrosome and Centrioles



# Centrioles can Form Basal Bodies for Cilia & Flagella



# Outline

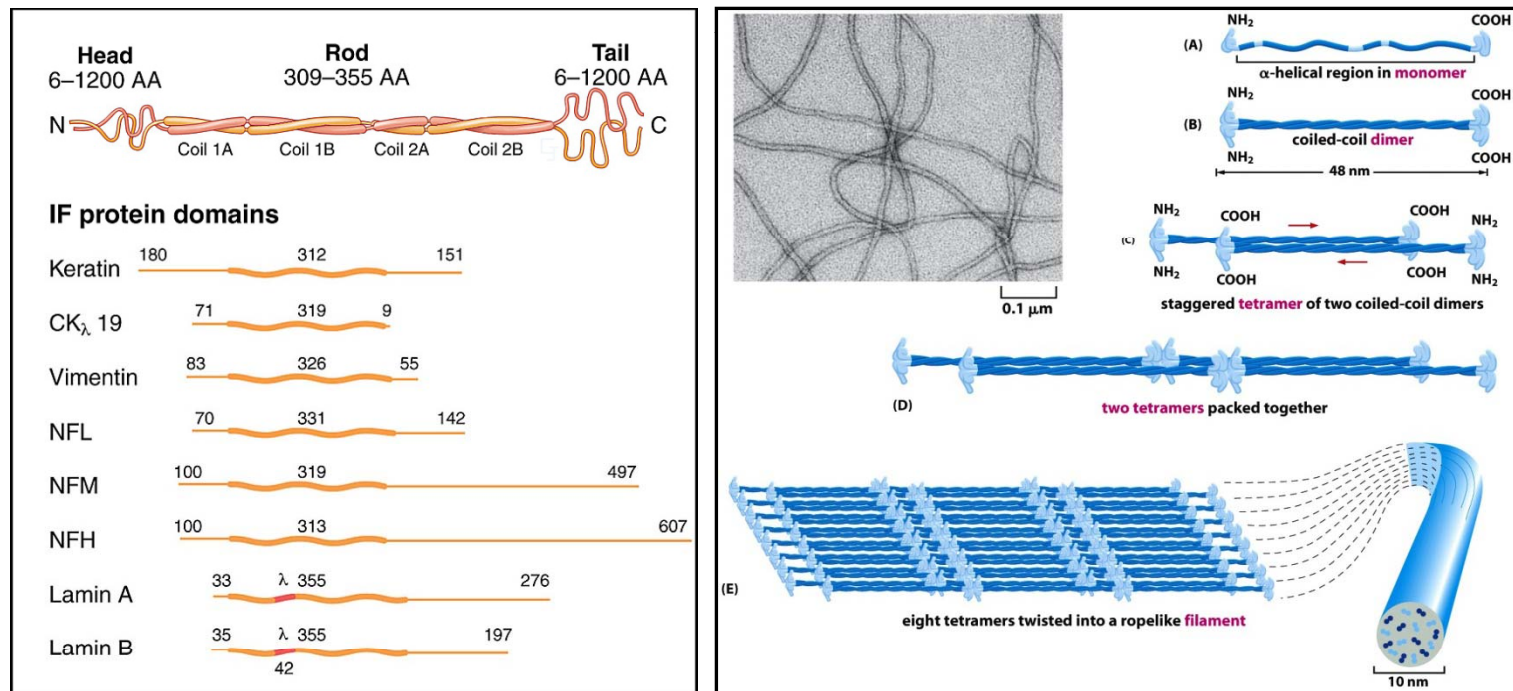
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- **Intermediate filament and its associated proteins**
- An overview of molecular motors



# Intermediate Filament

- So named because of its diameter in striated muscles (diameter ~10nm).
- Its core structure is an  $\alpha$ -helical coiled coil.
- N- and C-terminal domains vary considerably in size.



# Intermediate Filament Classification

**Table 35-1**

## CLASSIFICATION OF INTERMEDIATE FILAMENT PROTEINS BASED ON ROD DOMAIN SEQUENCES

| Class | Type                 | Genes | Molecule                                     | Distribution                           | Diseases   |
|-------|----------------------|-------|--|--|--|
| I     | Acidic keratin       | >15   | 40–65 kD, obligate heterodimer with class II | Epithelial cells                       | Blistering skin, corneal dystrophy, brittle hair and nails   |
| II    | Basic keratin        | >15   | 51–68 kD, obligate heterodimer with class I  | Epithelial cells                       | Similar to class I   |
| III   | Desmin               | 1     | 53 kD, homopolymers                          | Muscle cells                           | Cardiac and skeletal myopathies  |
|       | GFAP                 | 1     | 50 kD, homopolymers                          | Glial cells                            | Alexander disease; mouse null viable   |
|       | Peripherin           | 1     | 57 kD  | Peripheral > CNS neurons               |  |
|       | Synemin              | 1     | 190 kD, interacts with other class III IFs   | Muscle cells                           |  |
|       | Vimentin             | 1     | 54 kD, homopolymers and heteropolymers       | Mesenchymal cells                      | Mouse null viable  |
| IV    | Neurofilament        |       |  |  |  |
|       | NFL                  | 1     | Obligate heteropolymers with NFM, NFH        | Neurons                                | Mouse null viable; neuropathies  |
|       | NFM                  | 1     | Obligate heteropolymers with NFL, NFH        | Neurons                                |  |
|       | NFH                  | 1     | Obligate heteropolymers with NFL, NFM        | Neurons                                | Mutations a risk factor in amyotrophic lateral sclerosis   |
|       | $\alpha$ -Internexin | 1     | 55 kD, homopolymers                          | Embryonic neurons                      |  |
| V     | Lamins               | 4     | 7 Isoforms, 62–72 kD, homodimers             | Animal, plant nuclei                   | Cardiomyopathy, lipodystrophy, one form of Emery-Dreifuss muscular dystrophy, two forms of progeria plus many others |
| VI    | Nestin               | 1     | 230 kD, homopolymers                         | Embryonic neurons, muscle, other cells |  |

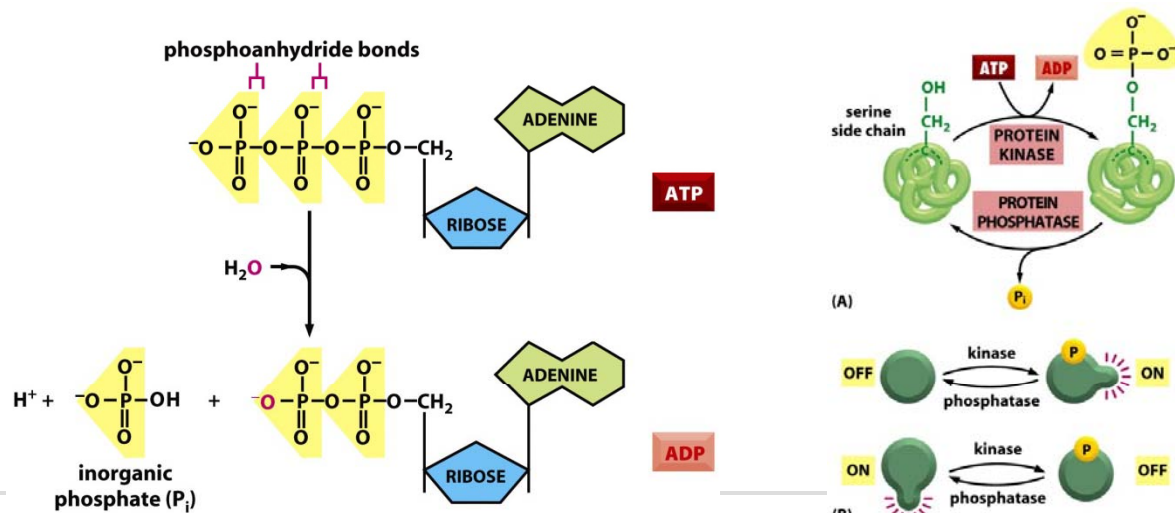
IF, intermediate filament; NFH, neurofilament heavy; NFL, neurofilament light; NFM, neurofilament medium.

Reference: Omary MB, Coulombe PA, McLean WHI: Intermediate filament proteins and their associated diseases. New Engl J Med 351:2087–2100, 2004.



# Intermediate Filament

- Biochemically most stable among the three cytoskeletal filaments.
- Assembly disassembly regulated by phosphorylation
  - Phosphorylation: reaction in which a phosphate group is covalently coupled to another molecule.
  - Kinase: an enzyme that catalyzes the addition of phosphate groups
  - Phosphatase: an enzyme that catalyzes the removal of phosphate groups
- Phosphorylation may stabilize or destabilize intermediate filaments.



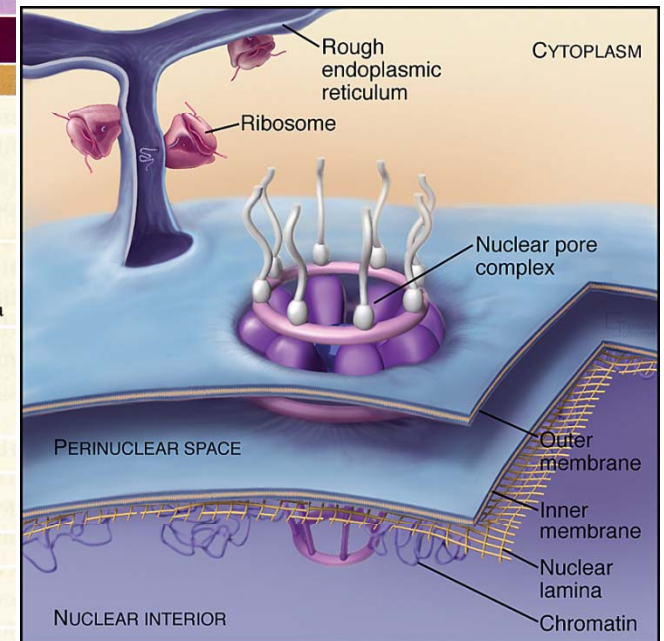
# Intermediate Filament Associated Proteins

Table 35-2

## PROTEINS ASSOCIATED WITH INTERMEDIATE FILAMENTS

| Name                    | Molecule   | Distribution  | Partners  | Diseases  |
|-------------------------|--|---|---|---|
| <b>Plakins</b>          |  |   |   |   |
| BPAG-1                  | Multiple splice isoforms (a, b, c, n) with ABDs and plakin domains $\pm$ spectrin and plakin repeats | a: Hemidesmosomes<br>b: Muscle, cartilage<br>c: Epithelial hemidesmosomes<br>n: Neurons | IFs, MTs, actin   | Autoimmune bullous pemphigoid   |
| Desmoplakin             | Two splice isoforms with plakin and coiled-coil domains and plakin repeats                           | Desmosomes  | IFs; cadherin and other desmosome proteins                    | Autoimmune pemphigus; genetic striate palmoplantar keratoderma              |
| Plectin                 | Multiple splice isoforms; ABD, plakin domain and plakin repeats                                      | Most tissues except neurons   | IFs, actin, MTs, spectrin, $\beta 4$ integrin                 | Autoimmune pemphigus; genetic epidermolysis bullosa with muscular dystrophy |
| <b>Epidermal</b>        |  |   |   |   |
| Filaggrin               | Ten 37-kD filaggrins cut by proteolysis from profilaggrin  | Cornified epithelia   | Aggregates keratin  | ?   |
| <b>Lamin Associated</b> |  |   |   |   |
| LAP1                    | 57-70 kD isoforms  | Integral nuclear membrane proteins  | Binds laminin to nuclear envelope                             |   |
| LAP2                    | 50 kD  | Integral nuclear membrane protein   | Binds laminin to nuclear envelope                             |   |
| LBR                     | 73 kD  | Integral nuclear membrane protein   | Binds laminin to nuclear envelope                             | Pelger-Huët anomaly; Greenberg skeletal dysplasia                           |
| Emerin                  | 34 kD  | ? Peripheral protein of the inner nuclear membrane                                      | ? Nucleates and binds actin filaments to the nuclear envelope | Emery-Dreifuss muscular dystrophy   |

ABD, actin binding domain; IFs, intermediate filaments; MTs, microtubules.





## Intermediate filament: orange

# Intermediate Filaments

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- Intermediate filaments are flexible but stable.
- Primary assumed function of intermediate filaments is to prevent excessive stretching.
- In general, much less is known about intermediate filaments compared to actin and microtubule.

# An Example: Microtubule, Intermediate Filament & Organelles in a Frog Axon

Cross-linker System between Neurofilaments, Microtubules, and Membranous Organelles in Frog Axons Revealed by the Quick-freeze, Deep-etching Method

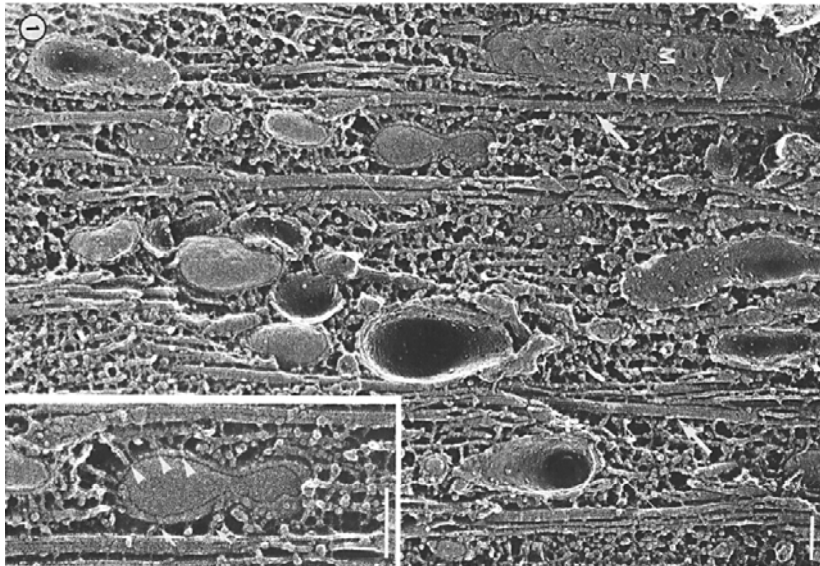
NOBUTAKA HIROKAWA

*Department of Physiology and Biophysics, Washington University School of Medicine, St. Louis, Missouri 63110*



Nobutaka Hirokawa  
University of Tokyo

N. Hirokawa, J. Cell Biol. 94:129, 1982



Bar: 0.1 $\mu$ m

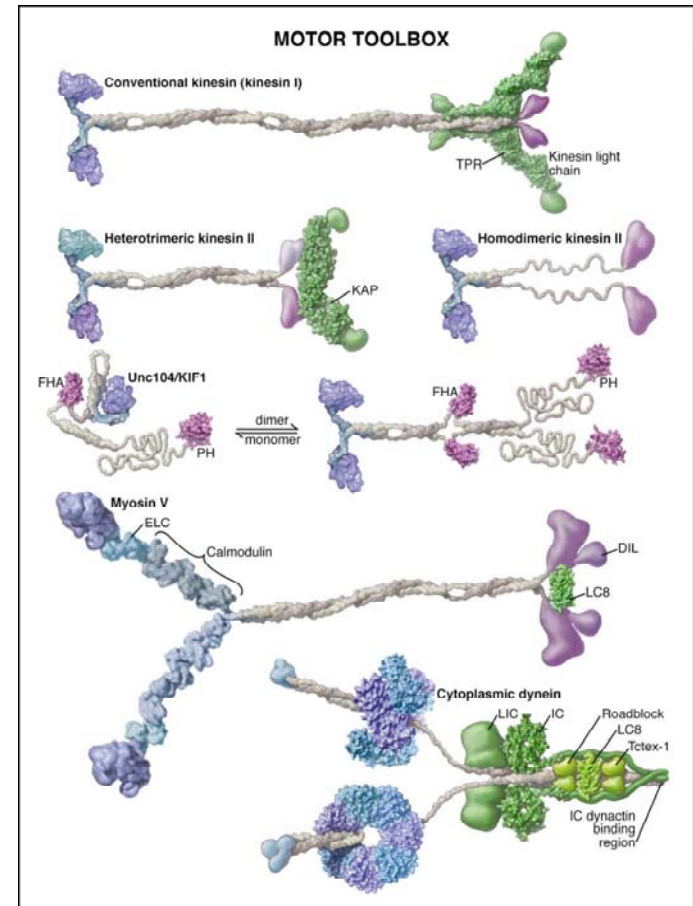
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# Overview of Molecular Motors

- Myosin binds and walks on actin.
- Kinesin and dynein binds and walks on microtubule.

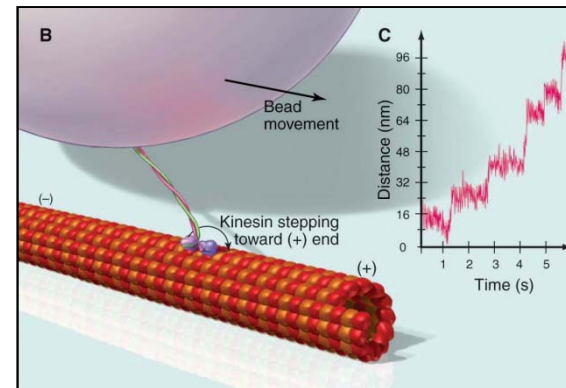
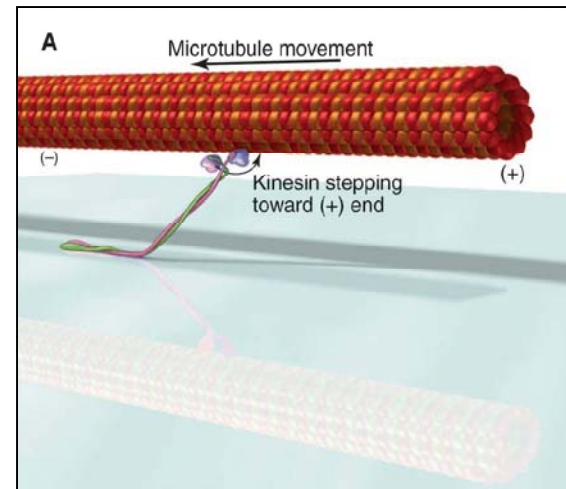


Vale RD, *Cell*, 112:467,2003



# Molecular Motors and Motility Assays

- Actin motor
  - myosin
  - Usually for short-distance movement
- Microtubule motors
  - kinesin
  - dynein
  - Usually for long-distance movement
- In vitro motility assays
  - bead assay
  - microtubule sliding assay





# Bead Motility Assay Video

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kinesin-coated bead moves along a microtubule  
Block Lab, Stanford

# Many Molecules Generate Intracellular Force & Motion

| Table 36-1  |             |               |                                   |  |
|---|-------------|---------------|-----------------------------------|--|
| EXAMPLES OF MECHANOCHEMICAL ATPases AND OTHER SYSTEMS |             |               |                                   |  |
| Families  | Track       | Direction     | Cargo                             | Energy                                     |
| <b>ATPases</b>  |             |               |                                   |  |
| <i>Myosins</i>  |             |               |                                   |  |
| Muscle myosin   | Actin       | Barbed end    | Myosin filament                   | ATP  |
| Myosin II   | Actin       | Barbed end    | Myosin, actin                     | ATP  |
| Myosin I  | Actin       | Barbed end    | Membranes                         | ATP  |
| Myosin V  | Actin       | Barbed end    | Organelles                        | ATP  |
| Myosin VI   | Actin       | Pointed end   | Endocytic vesicles                | ATP  |
| <i>Dyneins</i>  |             |               |                                   |  |
| Axonemal  | Microtubule | Minus end     | Microtubules                      | ATP  |
| Cytoplasmic   | Microtubule | Minus end     | Membranes, chromosomes            | ATP  |
| <i>Kinesins</i>                                       |             |               |                                   |  |
| Conventional  | Microtubule | Plus end      | Membranes, intermediate filaments | ATP  |
| Ncd   | Microtubule | Minus end     | ? Microtubules                    | ATP  |
| <b>Other Mechanochemical Systems</b>                  |             |               |                                   |  |
| <i>Polymerases</i>                                    |             |               |                                   |  |
| Ribosome  | mRNA        | 5' to 3'      | None                              | GTP  |
| DNA polymerase  | DNA         | 5' to 3'      | None                              | ATP  |
| RNA polymerase  | DNA         | 5' to 3'      | None                              | ATP  |
| <i>Conformational System</i>                          |             |               |                                   |  |
| Spasmin/centrin                                       | None        | None          | Cell, basal body                  | Ca <sup>2+</sup>                           |
| <i>Polymerizing Systems</i>                           |             |               |                                   |  |
| Actin filaments                                       | None        | Barbed end    | Membranes                         | ATP  |
| Microtubules  | None        | Plus end      | Chromosomes                       | GTP  |
| Worm sperm MSP  | None        | Not polar     | Cytoskeleton                      |  |
| <i>Rotary Motors</i>                                  |             |               |                                   |  |
| Bacterial flagella                                    | None        | Bidirectional | Cell                              | H <sup>+</sup> or Na <sup>+</sup> gradient |
| F-type ATPase   | None        | Bidirectional | None                              | H <sup>+</sup> or ATP                      |
| V-type ATPase pump                                    | None        |               | None                              | ATP  |

mRNA, messenger RNA; MSP, major sperm protein.

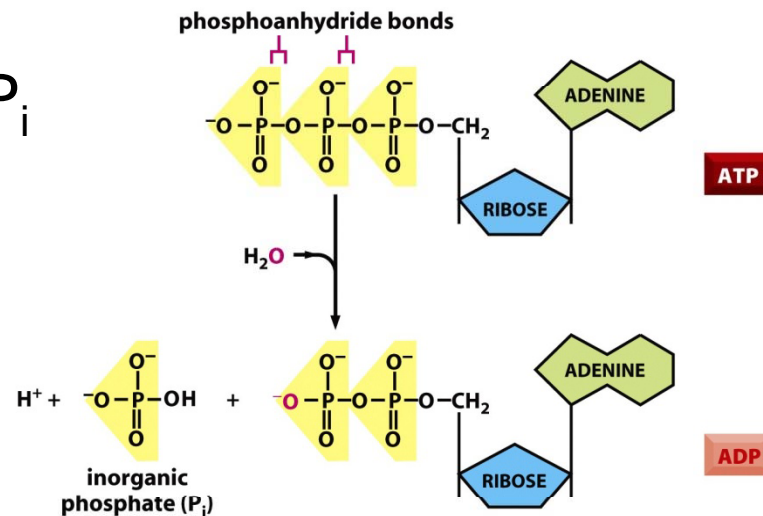
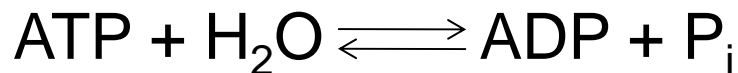
# Relations Between Molecular Motors and Cytoskeleton Polymers

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- Interactions between motors and cytoskeletal polymers are dynamic and complex.
- Cytoskeletal polymers provide dynamic tracks for molecular motors to walk on.
- Molecular motors active interacts with cytoskeletal polymers.  
For example,
  - Molecular motors transport cytoskeletal polymers, especially in neurons.
  - Molecular motors, e.g. MCAK, regulate cytoskeletal polymer dynamics.

# Molecular Motors Are ATP-Hydrolysis Enzymes

- Molecular motors convert chemical energy derived from ATP hydrolysis directly into mechanical work.
- ATP (adenosine triphosphate) hydrolysis



# Motor Behavior Parameters

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- Parameters that characterizing motor behaviors
  - processivity: run-length, number of steps
  - step size
  - stall force
- Myosin is nonprocessive.
- Kinesin and dynein are both processive. Processivity of dynein is weaker.
- Motors walk nano-meter scale steps of specific lengths.
- Stall force is on the pico-Newton level.

# Analyzing Motor Movement at Nanometer Resolution

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- Nanometer-resolution measurement of step sizes
  - First implemented in late 1980's
  - Based on fitting of point spread function

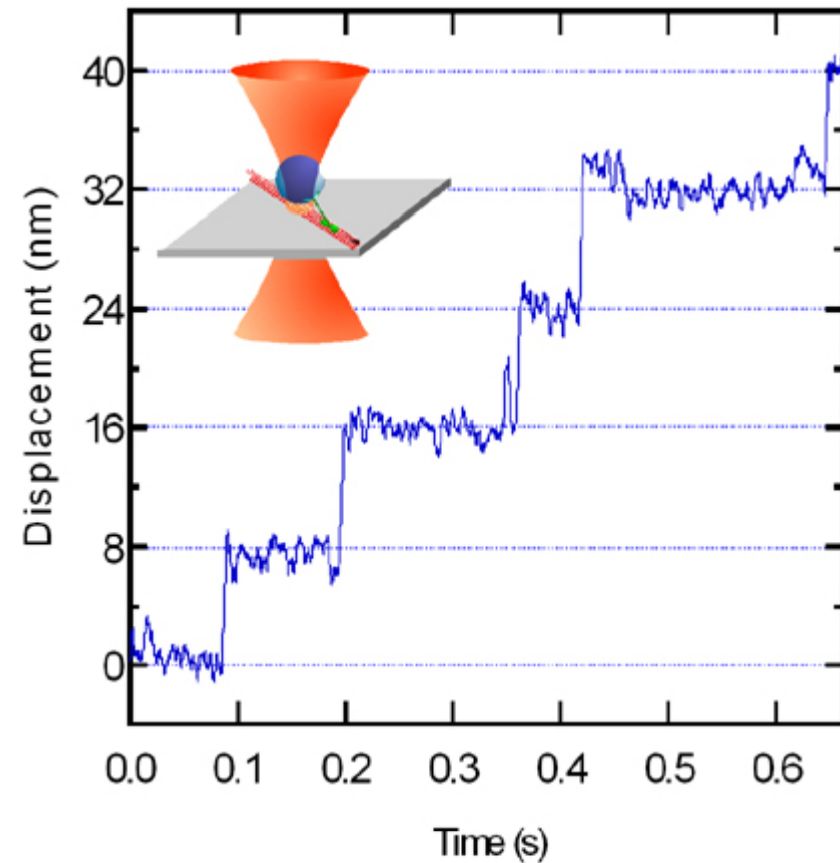
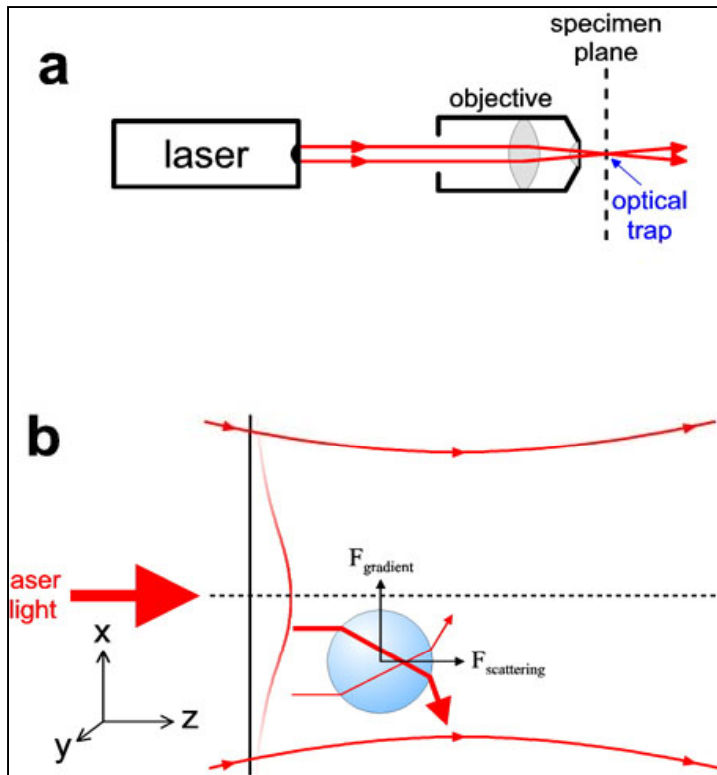
Tracking kinesin-driven movements with nanometre-scale precision. Gelles J, Schnapp BJ, Sheetz MP. *Nature*. 331:450-3 (1988).

- Further improved by many others
  - Up to 1nm resolution

Ahmet Yildiz, Paul R.Selvin. Fluorescence Imaging with One Nanometer Accuracy (FIONA): Application to Molecular Motors, *Accounts of Chemical Research*, 38(7), 574-82 (2005)

# Analyzing Motor Force at Piconewton Resolution

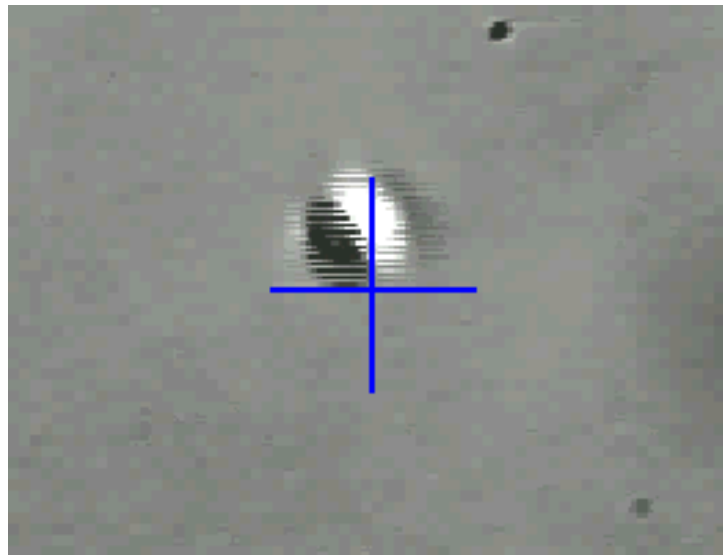
- Optical tweezer is used to generate and measure motor stall force.



From Steven Block Lab webpage  
<http://www.stanford.edu/group/blocklab/>

# Laser Force Trap Video

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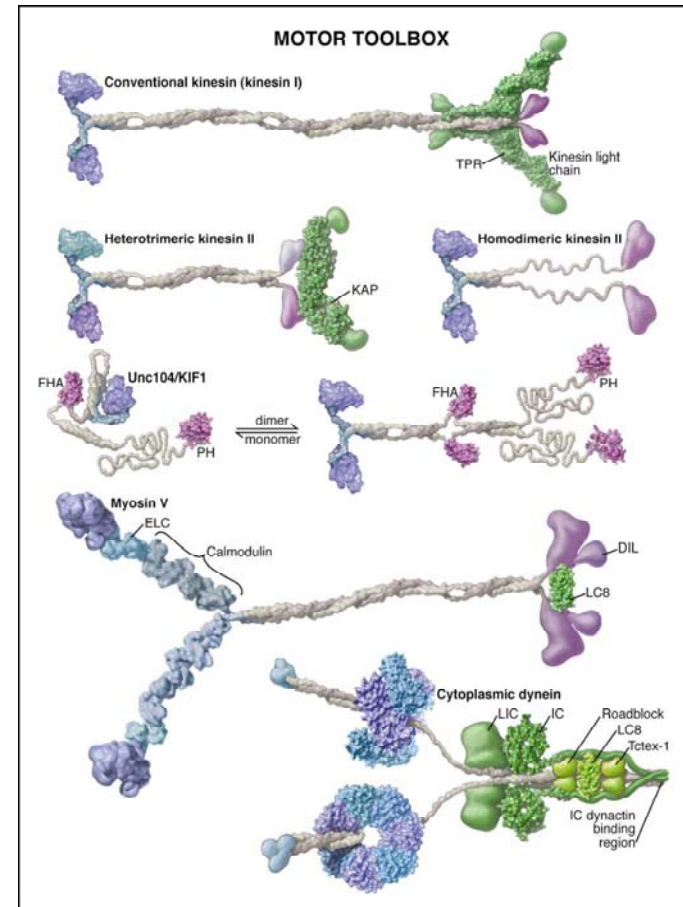


kinesin-coated bead moves in a force trap  
Block Lab, Stanford



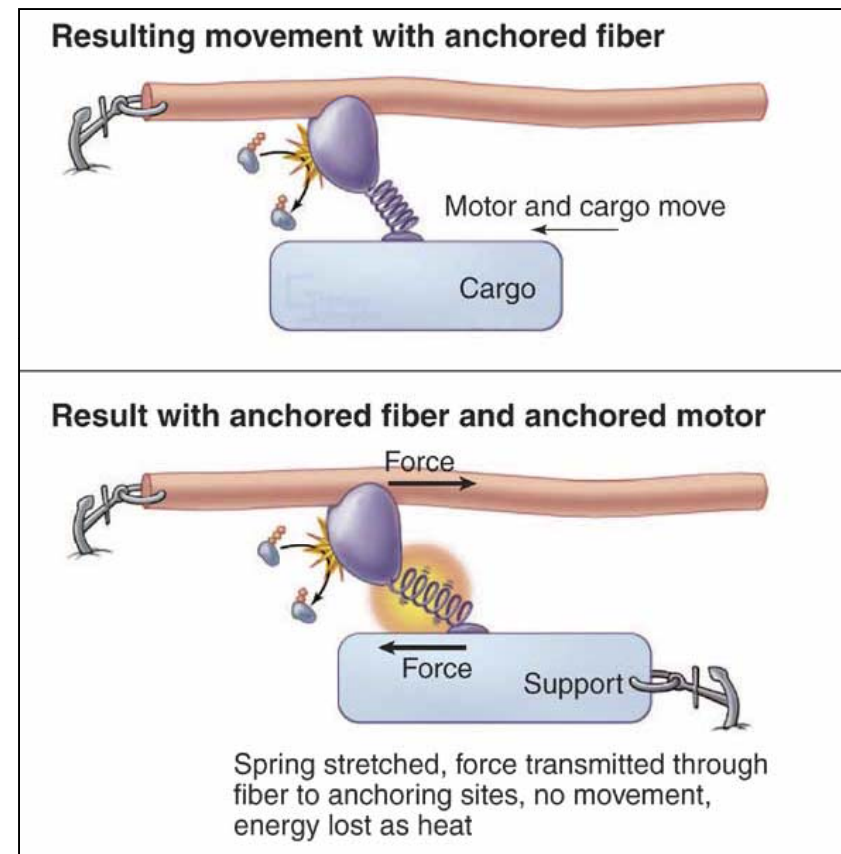
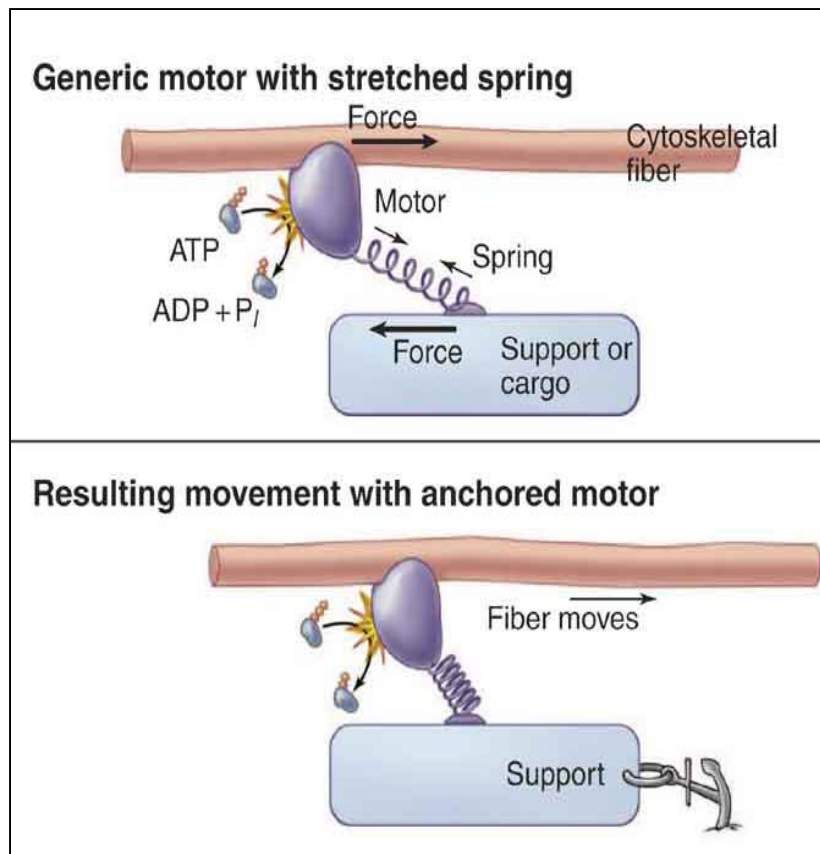
# General Motor Structure

- Motor (head) domain
  - Produces force and motion
- Tail domain
  - Adapts to different cargoes



Vale RD, *Cell*, 112:467,2003

# Different Motility Schemes



# Required Reading

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- Chapters 2 & 16

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**Questions ?**